RESEARCH ARTICLE

The START domain potentiates HD-ZIPIII transcriptional activity

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Short title: START-dependent potentiation of HD-ZIPIII TFs

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Abstract

The CLASS III HOMEODOMAIN-LEUCINE ZIPPER (HD-ZIPIII) transcription factors (TFs) were repeatedly deployed over 725 million years of evolution to regulate central developmental innovations. The START domain of this pivotal class of developmental regulators was recognized over twenty years ago, but its putative ligands and functional contributions remain unknown. Here, we demonstrate that the START domain promotes HD-ZIPIII TF homodimerization and increases transcriptional potency. Effects on transcriptional output can be ported onto heterologous TFs, consistent with principles of evolution via domain capture. We also show the START domain binds several species of phospholipids, and that mutations in conserved residues perturbing ligand binding and/or its downstream conformational readout, abolish HD-ZIPIII DNA-binding competence. Our data present a model in which the START domain potentiates transcriptional activity and uses ligand-induced conformational change to render HD-ZIPIII dimers competent to bind DNA. These findings resolve a long-standing mystery in plant development and highlight the flexible and diverse regulatory potential coded within this widely distributed evolutionary module.

IN A NUTSHELL

Background: Development has been compared to a ball rolling down a hill. Cells initially have broad potential, but over time, they make decisions based on local cues which set them down a path to differentiation. Along this trajectory, thousands of genes need to be turned off or on in a carefully choreographed manner to ensure specialized identity. This is accomplished in part by transcription factors whose activities are precisely controlled by inputs operating across multiple regulatory levels. Here, we study the regulation of CLASS III HOMEODOMAIN LEUCINE ZIPPER (HD-ZIPIII) proteins, a 725-million-year-old family of transcription factors that were redeployed throughout evolution to impact nearly all aspects of plant development.

Question: HD-ZIPIII proteins contain a START domain, an evolutionarily ubiquitous module that binds lipophilic ligands. The hypothesis thus arose that HD-ZIPIII activity may be controlled by lipid ligand inputs. Remarkably, both the ligands and regulatory properties of the HD-ZIPIII START domain have remained unknown for over twenty years. The goal of our study was to resolve this long-standing mystery.

Findings: We find that the START domain promotes HD-ZIPIII dimerization and increases their transcriptional potency. We also identify several phospholipid ligands bound by the HD-ZIPIII START domain, and show that perturbing ligand binding abolishes DNA-binding competence. Thus, the START domain turns HD-ZIPIII proteins into potent DNA-binding competent transcription factors but only if they can bind and respond to their phospholipid ligands. Our findings further highlight the remarkably flexible and diverse regulatory potential of START domains.

Next steps: Having established START regulatory properties for an HD-ZIPIII protein in *Arabidopsis*, we are now considering our findings from an evolutionary perspective. How and when did these regulatory properties come about? Is there functional divergence of START regulation of HD-ZIPIII activity? And how are START domains able to employ such a diverse set of regulatory mechanisms?

Introduction

Development of multicellular organisms requires the precise control of transcription factor (TF) inputs into their gene regulatory networks. As such, the activity of TFs is highly regulated, often integrating distinct mechanisms across multiple regulatory levels to impact developmental outcomes. In plants, this is exemplified by CLASS III HOMEODOMAIN-LEUCINE ZIPPER (HD-ZIPIII) proteins, an ancient TF family that arose after the divergence of unicellular Chlorophyta but before the emergence of Streptophyte algae and land plants over 725 million years ago (Ariel et al., 2007; Goodstein et al., 2012; Romani et al., 2018). HD-ZIPIII TFs have been repeatedly co-opted throughout plant evolution to regulate key developmental advances (McConnell et al., 2001; Juarez et al., 2004; Prigge et al., 2005;

Kelley et al., 2009; Carlsbecker et al., 2010; Robischon et al., 2011; Sebastian et al., 2015; Yip et al., 2016; Ramachandran et al., 2017; Xu et al., 2019). For instance, in *Arabidopsis thaliana*, HD-ZIPIII genes contribute to vascular specification (Carlsbecker et al., 2010; Ramachandran et al., 2017; Smetana et al., 2019), root and shoot apical meristem maintenance (Prigge et al., 2005; Sebastian et al., 2015; Ramachandran et al., 2017), and the distinction of adaxial tissues in lateral organs (McConnell et al., 2001; Prigge et al., 2005). These innovations parallel the increasing complexity of plant form and were instrumental to the enormous success of land plants (Ariel et al., 2007; Goodstein et al., 2012; Romani et al., 2018).

In keeping with a critical role in development, HD-ZIPIII activity is subject to intricate regulation. HD-ZIPIII transcripts are targeted by small RNAs of the miR166 family, which restricts their accumulation via morphogen-like patterning properties (Skopelitis et al., 2017). Loss of this regulation conditions gain-of-function phenotypes impacting nearly all aspects of plant development (McConnell et al., 2001). At the protein level, HD-ZIPIII activity is modulated in part through interaction with their direct targets, the LITTLE ZIPPER (ZPR) family. ZPR proteins capture HD-ZIPIII TFs into heteromeric complexes lacking DNA-binding potential, creating a negative feedback loop that fine-tunes HD-ZIPIII activity (Wenkel et al., 2007; Kim et al., 2008; Husbands et al., 2016).

An additional layer of regulation is suggested by the fact that HD-ZIPIII proteins contain a START domain (**Fig. 1A**, Ponting & Aravind, 1999). START domains are members of the StARkin superfamily (derived from 'kin of steroidogenic acute regulatory protein (StAR)'), which are present throughout the tree of life (Wong et al., 2016; Dresden et al., 2021). StARkin domains are characterized by an α/β helix-grip fold structure with a deep hydrophobic pocket that accommodates lipophilic ligands such as long-chain fatty acids, sterols, and isoprenoids (Tsujishita et al., 2000; Roderick et al., 2002; Hubbard et al., 2010). Ligand binding induces stereotypical conformational changes that activate StARkin proteins through a diverse set of non-mutually exclusive regulatory mechanisms (reviewed in Dresden et al.,

2021). For instance, StARkin domains can control protein turnover, homomeric and heteromeric complex stoichiometry, subcellular localization, and secondary structure stability (Alpy et al., 2005; Kanno et al., 2007; Park et al., 2009; Du et al., 2012; Schrick et al., 2014; Belda-Palazon et al., 2018; Zhang et al., 2018; Iida et al., 2019; Tillman et al., 2020; Nagata et al., 2021). The presence of a homeodomain and a START domain therefore sparked a long-standing hypothesis that the transcriptional activity of HD-ZIPIII proteins may be controlled by a lipid ligand, in a manner reminiscent of nuclear receptors in mammalian systems (Schrick et al., 2004; Lumba et al., 2010; Sladek, 2011). Remarkably, the role of the HD-ZIPIII START domain remains unclear, despite the essential roles these TFs play in development (McConnell et al., 2001; Prigge et al., 2005).

Potential insights may come from the evolutionarily related HD-ZIPIV family, which also diverged at least 725 million years ago (Ariel et al., 2007; Goodstein et al., 2012; Romani et al., 2018), and whose START domain is required for function (Schrick et al., 2014; Iida et al., 2019; Nagata et al., 2021). Yeast expression analyses indicate that this START domain binds a broad spectrum of metabolites and increases TF stability (Schrick et al., 2014). Recent studies find similar promotive effects on protein stability in plants and identify subcellular localization as an additional HD-ZIPIV START regulatory mechanism controlling epidermal cell fate (Iida et al., 2019; Nagata et al., 2021). These effects are thought to be mediated by binding of epidermally-synthesized ceramides, which would reinforce their tissue-specific activity (Iida et al., 2019; Nagata et al., 2021). Given the long evolutionary divergence of HD-ZIPIII proteins (Ariel et al., 2007; Goodstein et al., 2012; Romani et al., 2018), their functions outside of the epidermis (McConnell et al., 2001; Juarez et al., 2004; Prigge et al., 2005; Kelley et al., 2009; Carlsbecker et al., 2010; Robischon et al., 2011; Sebastian et al., 2015; Yip et al., 2016; Ramachandran et al., 2017; Xu et al., 2019), and the fact that HD-ZIPIII and HD-ZIPIV START domains are not interchangeable in yeast or plant assays (Schrick et al., 2014), the extent to which these observations apply to the HD-ZIPIII START domain is difficult to predict.

We show that addition of the HD-ZIPIII START domain potentiates transcriptional activity by promoting homodimerization and increasing transcriptional potency. Further, mutations that affect ligand binding and likely its downstream conformational response abolish DNA-binding competence, without overt effects on protein stability, subcellular localization, and interaction partners. Thus, the HD-ZIPIII START domain potentiates TF activity through regulatory mechanisms distinct from the evolutionarily related HD-ZIPIV START domain. These findings resolve a long-standing mystery in plant development and highlight the flexible and diverse regulatory potential coded within the ubiquitously distributed StARkin evolutionary module.

Results

The START domain is required for full PHB function

We assessed START-dependent effects on HD-ZIPIII developmental function using *Arabidopsis thaliana* PHABULOSA (PHB) as a representative member. We began by replacing the PHB START domain in a functional YFP-tagged *pPHB:PHB* reporter (Skopelitis et al., 2017) with the 21-nt microRNA166 (miR166) recognition site found within the START domain coding sequence (*pPHB:PHB-Delta*, Rhoades et al., 2002). Whereas the *pPHB:PHB* transgene complements the *phb, phavoluta (phv),* and *corona (cna)* triple mutant phenotype, the *pPHB:PHB-Delta* construct fails to rescue (**Fig. 1B**), despite equivalent accumulation of *PHB* and *PHB-Delta* transcripts (**Supplemental Fig. S1A**). We therefore next introduced a silent mutation into the 21-nt miR166 binding site, abolishing miR166 regulation of PHB and PHB-Delta (*pPHB:PHB*; pPHB:PHB*-Delta*). Loss of miR166 regulation generates a highly sensitive, dosage-dependent readout of HD-ZIPIII activity (McConnell et al., 2001; Mallory et al., 2004; Carlsbecker et al., 2010; Skopelitis et al., 2017), permitting detection of weak or subtle HD-ZIPIII function that might be missed in standard complementation assays. As expected (Mallory et al., 2004), over 90% of miR166-insensitive *pPHB:PHB** primary transformants show PHB gain-of-function phenotypes (**Fig. 1C**). By contrast, *pPHB:PHB*-Delta* transformants are indistinguishable from wild-type

plants (**Fig. 1C**), despite similar ectopic accumulation of protein (**Supplemental Fig. S1B-E**). Further, transcript levels of the HD-ZIPIII direct targets *ZPR3* and *ZPR4* are strongly upregulated in *pPHB:PHB** lines but are indistinguishable from wild-type in *pPHB:PHB*-Delta* transformants (**Fig. 1D**). Thus, the START domain is required for PHB to fulfill its developmental function. Moreover, as PHB-Delta shows no obvious effects on protein stability or subcellular localization (**Supplemental Fig. S1B-E,** and see below), the HD-ZIPIII START domain employs regulatory mechanisms distinct from those of the evolutionary related HD-ZIPIV START domain.

The START domain promotes PHB dimerization

One frequently used StARkin regulatory mechanism is modulation of homomeric stoichiometry (Fujii et al., 2009; Yin et al., 2009; Hubbard et al., 2010; Gatta et al., 2015; Prashek et al., 2017; Tillman et al., 2020; Sanchez-Solana et al., 2021). To test whether the START domain impacts PHB homodimerization, we used single-molecule pull down (SiMPull), which we previously adapted for plant systems (Husbands et al., 2016). We first determined the maturation frequency of the monomeric citrine yellow fluorescent protein (YFP) variant in Arabidopsis to calibrate the frequency of two-step photobleaching events into a quantitative assessment of homomeric stoichiometry (**Figs. 2A** and **S2,** Jain et al., 2011; Jain et al., 2014; Aggarwal et al., 2014; Husbands et al., 2016**)**. Subsequent analyses of over 3300 protein complexes showed that ~80% of wild-type PHB proteins are present as dimers (**Fig. 2B**). This frequency resembles that of strongly homodimeric proteins in animal systems (Jain et al., 2011; Jain et al., 2014), and predicts PHB functions primarily as a homodimer. By contrast, PHB-Delta has a dimerization frequency of 15% – about twice the frequency with which two-step photobleaching events are observed by chance (**Fig. 2B** and **Supplemental Fig. S2B**). Thus, the PHB START domain promotes either the formation or the maintenance of PHB homodimers (**Fig. 2B**). As HD-ZIPIII proteins require dimerization to bind DNA (Sessa et al., 1998), this effect on homodimerization provides a potential explanation for PHB-Delta failing to activate the normal PHB developmental program (**Figs. 1B-D**).

The START domain enhances PHB transcriptional potency

One consequence of poor dimerization of PHB-Delta is that it may obscure possible additional contributions from the START domain to HD-ZIPIII TF function. We therefore turned to a short-term estradiol-inducible overexpression system to increase the total number of PHB-Delta dimers available for molecular assays of TF activity. Importantly, estradiol induction does not change the subcellular localization or dimerization frequencies of PHB or PHB-Delta seen at native expression levels (**Supplemental Figs. S3A-S3D**). Both variants can also be induced to similar levels and show comparable protein stability (**Supplemental Figs. S3A** and **S3E**), making this approach suitable for identifying additional START-dependent effects.

Using short-term estradiol-inductions, we first tested whether PHB proteins lacking the START domain are capable of binding to DNA using chromatin immunoprecipitation (ChIP). PHB occupies multiple palindromic HD-ZIPIII binding sites (Sessa et al., 1998) in the regulatory regions of its *ZPR3* and *ZPR4* direct targets (**Figs. 2C** and **2D**). Similarly, enrichment at these sites was also detected for PHB-Delta (**Figs. 2C** and **2D**), indicating this variant retains the capacity to bind to DNA.

Given this outcome, we next gauged the effect of the START domain on transcriptional potency. Here, the short-term estradiol-induction system has the added benefit that it provides a direct quantitative readout of transcriptional activity while avoiding confounding consequences of morphological changes and regulatory feedback (Wenkel et al., 2007; Kim et al., 2008). As expected, *ZPR3* and *ZPR4* transcript levels are strongly upregulated upon induction of PHB (**Fig. 3A**). *ZPR3* and *ZPR4* targets are also upregulated upon induction of PHB-Delta (**Fig. 3A**). Intriguingly, these transcripts are upregulated to between one-half and one-third the levels seen for PHB, despite equivalent induction of *PHB** and *PHB*-Delta* (**Fig. 3A**). As PHB variants are not differentially stable (**Supplemental Figs. S1B-E** and **S3E**), and ChIP experiments indicate near equivalent promoter occupancy (**Fig. 2D**), these data suggest deletion of the START domain, in addition to reducing the frequency of dimers, significantly reduces their

transcriptional potency. This idea is supported by two orthogonal lines of evidence. First, virtually identical results were obtained using co-transfection assays in *Nicotiana benthamiana*, which show PHB-Delta fails to fully activate both endogenous *ZPR* targets and a *pZPR3:3x-NLS-RFP* reporter (**Supplemental Fig. S4**). Second, the phenotypic severity of transformants constitutively overexpressing PHB-Delta (*p35S:PHB*-Delta*) is markedly lower than the *p35S:PHB** control (**Supplemental Fig. S5A**). *ZPR3* and *ZPR4* transcripts in *p35S:PHB*-Delta* lines also accumulate to between one-half and one-third the levels seen in *p35S:PHB** seedlings, despite equivalent accumulation of *PHB** and *PHB*-Delta* transcripts (**Supplemental Fig. S5B**). These complementary assays support augmenting of transcriptional potency as an additional HD-ZIPIII START regulatory mechanism.

START regulatory properties are transferable onto heterologous TFs

The idea that the PHB START domain potentiates TF activity and is not strictly required for TF identity *per se* is in line with principles of TF evolution by domain capture (Lynch & Wagner, 2008; de Mendoza et al., 2013; Jarvela & Hinman et al., 2015; Sebe-Pedros et al., 2017). Relevant to this, plant genomes encode HD-ZIP TFs that lack a START domain (Ariel et al., 2007; Goodstein et al., 2012; Romani et al., 2018). These related HD-ZIP members provide a unique opportunity to definitively test how addition of a START domain impacts TF output. We therefore inserted the PHB START domain downstream of the homeodomain and leucine zipper motifs of the HD-ZIPI family member ATHB12, partially recapitulating HD-ZIPIII architecture (ATHB12-START). ATHB12 is a known activator of transcription and is sufficient to drive expression of reporters placed downstream of multimeric HD-ZIPI binding sites (**Fig. 3B**, Lee et al., 2012). Remarkably, addition of the HD-ZIPIII START domain increases ATHB12 transcription activity by a factor of three over unmodified ATHB12 (**Fig. 3B**). This effect on transcriptional potency parallels effects seen for PHB and PHB-Delta in Arabidopsis as well as equivalent co-transfection assays in *N. benthamiana* (**Figs. 3A** and **S4**). Thus, the START domain is necessary for HD-ZIPIII dimers to achieve full transcriptional potency, and its addition is sufficient to confer this increase in potency onto heterologous TFs.

The START domain does not affect heteromeric stoichiometry

At a molecular level, StARkin domains undergo stereotypical conformational changes in response to ligand binding (Tsujishita et al., 2000; Roderick et al., 2002). In the *apo* form, StARkin domains reveal an open ligand-binding pocket, and upon ligand interaction the pocket is sealed via conformational changes. In addition to changes in tertiary structure, this creates a new interaction surface that, for a subset of StARkin proteins, mediates recruitment of additional protein partners to exert StARkin-dependent regulatory effects Fujii et al., 2009; Yin et al., 2009; Hubbard et al., 2010; Gatta et al., 2015; Prashek et al., 2017; Tillman et al., 2020; Sanchez-Solana et al., 2021). To test this possibility, we performed quantitative mass spectrometry on proteins that co-immunoprecipitate with PHB (IP-MS; **Supplemental Fig. S6**). Among the proteins significantly enriched across five PHB IP-MS replicates were multiple members of the BRAHMA-containing Switch/Sucrose Non-fermentable (SWI/SNF) chromatin-remodeling complex. These data point to specificity of the IP-MS and suggest HD-ZIPIII proteins facilitate transcription via direct modification of chromatin. Further supporting specificity of the IP-MS, the known HD-ZIPIII interacting partners ZPR1 and ZPR3 (Wenkel et al., 2007; Kim et al., 2008; Husbands et al., 2016) also co-immunoprecipitate with PHB. Finally, significant enrichments were detected for the lipid-binding proteins OLEOSIN1 (OLEO1), OLEO5, DYNAMIN-RELATED PROTEIN 1C (DRP1C), DRP1E, and ANNEXIN 4 (ANNAT4), which is of potential interest given the lipid-binding nature of StARkin domains (**Supplemental Fig. S6** and **Supplemental Data Set 1**). Protein-protein interactions are, however, not mediated via the START domain as IP-MS shows that PHB-Delta binds the same interaction partners (**Supplemental Data Set 2**), and SiMPull co-localization analyses confirm that interaction with ZPR3 is unchanged (**Supplemental Fig. S7**). These findings thus argue against START-mediated regulation of HD-ZIPIII complex stoichiometry at the level of interaction partners.

Taken together, complementary phenotypic and molecular analyses reveal the START domain is required for PHB developmental function, but unlike its homolog in the HD-ZIPIV TFs, does not impact protein stability or subcellular localization. Instead, the presence of a START domain in PHB potentiates TF activity by promoting homodimerization and by increasing transcriptional potency. These effects seem to be mediated by interactions between domains of PHB, as the START domain does not appear to determine its spectrum of interaction partners.

Mutating conserved START residues abolishes PHB DNA-binding competence

Following the idea that, like other StARkin domains (Prashek et al., 2017; Tillman et al., 2020), the HD-ZIPIII START domain influences intramolecular domain-to-domain interactions, mutations perturbing this property may exert effects on TF function distinct from those observed after full deletion of the domain. To assess this possibility, we first performed homology modeling of the PHB START domain using I-TASSER and AlphaFold2 to identify conserved functional residues to target via mutagenesis. Both algorithms indicate the START domain is distinct from sterol- or isoprenoid-binding StARkin domains such as the abscisic acid (ABA) receptor (Yin et al., 2009), and instead resembles mammalian phosphatidylcholine transfer protein (PC-TP; **Fig. 4A**, Roderick et al., 2002). START domains like PC-TP contain several highly conserved residues that mediate the ligand-directed conformational change (Ponting & Aravind, 1999; Tsjuishita et al., 2000; Roderick et al., 2002; Baker et al., 2007). Accordingly, two short amino acid stretches (RDFTWLR and RAEMK), centered around three such arginine residues (Ponting & Aravind, 1999; Tsjuishita et al., 2000; Roderick et al., 2002; Baker et al., 2007), were selected for mutagenesis (SDmut; **Figs. 4A** and **S8**). Importantly, these mutations are predicted to minimally affect protein folding (RMSD 0.291; **Fig. 4A**), and circular dichroism confirmed that wild-type, SDmut, and PC-TP START domains purified from *E. coli* adopt virtually identical secondary structures (**Fig. 4B**).

Identical mutations were then introduced into the functional YFP-tagged *pPHB:PHB* reporter and its miR166-insensitive counterpart *pPHB:PHB**, creating *pPHB:PHB-SDmut* and *pPHB:PHB*-SDmut*,

respectively. The *pPHB:PHB-SDmut* transgene failed to complement the *phb phv cna* triple mutant phenotype (**Fig. 4C**), suggesting the selected residues are indeed required for PHB function. Supporting this, *pPHB:PHB*-SDmut* primary transformants did not show phenotypes in the more-sensitive dose-dependent, gain-of-function assay (**Fig. 4D**), and *ZPR3* and *ZPR4* transcript levels in these lines were indistinguishable from the wild-type (**Fig. 4E**). These effects are again not explained by changes in protein stability, sub-cellular localization, or interacting partners, as these properties are comparable for PHB, PHB-Delta, and PHB-SDmut (**Supplemental Figs. S1B-E, S3, S7,** and **Supplemental Data Set 2**). By contrast, SiMPull showed that homodimeric PHB-SDmut complexes are present at ~40% frequency in the population (**Fig. 5A**), approximately twice that seen for PHB-Delta but half that seen for wild-type PHB (**Fig. 2B**).

We then tested whether PHB-SDmut dimers retain DNA-binding capability. Interestingly, no enrichment of PHB-SDmut was detected in the regulatory regions of the *ZPR3* or *ZPR4* loci using ChIP (**Fig. 5B**), indicating these mutations abolish PHB DNA-binding competence. Consistent with this, PHB-SDmut fails to activate *ZPR3* and *ZPR4* in short-term estradiol-induction assays in Arabidopsis as well as in co-transfection assays in *N. benthamiana* (**Figs. 5C** and **S4**). Moreover, addition of this non-functional START domain to ATHB12 abolishes its transcriptional activity (ATHB12-SDmut; **Fig. 5D**). Thus, mutating the START domain, and deleting the START domain entirely, leads to proteins with distinct biochemical properties: PHB-SDmut dimerizes relatively well but is unable to bind DNA, whereas PHB-Delta rarely dimerizes but those dimers that do form retain DNA-binding competence and have reduced transcriptional potency.

The PHB START domain binds PC and mutating PC-binding residues abolishes DNA-binding

Structural modeling and mutational analyses propose the PHB START domain may be controlled by phospholipid ligands (**Figs. 4A**, **5**, and **S8**). We therefore tested whether the PHB START domain interacts with a set of ligands similar to those bound by PC-TP. Depending on context, these include

members of the PC, phosphatidylethanolamine (PE), and phosphatidylglycerol (PG) phospholipid classes (de Brouwer et al., 2001; Sablin et al., 2009; Zhai et al., 2017). To this end, recombinant PHB START domain protein was incubated with liposomes generated from Arabidopsis total lipid extracts, re-purified by affinity chromatography, and then subjected to LC-MS at two independent lipidomics centers. Prior to incubation with plant-derived liposomes, recombinant PHB START protein co-purified with bacterial PE and PG species (**Supplemental Figs. S9A, S9B, Supplemental Data Set 3**). Binding of these "fortuitous ligands" has also been reported for other recombinant PC-binding proteins including PC-TP (de Brouwer et al., 2001), steroidogenic factor 1 (SF-1, Sablin et al., 2009), and liver receptor homolog-1 (LRH-1, Zhai et al., 2017). After incubation with plant-derived liposomes (**Supplemental Fig. S9C**), additional lipids were significantly enriched across five PHB START LC-MS replicates (**Supplemental Data Sets 4–6**). These include five species of PC, two of which are preferred ligands for PC-TP (**Fig. 6A**; de Brouwer et al., 2001). Further, membrane-overlay assays indicate this binding is not occurring at the surface of the START domain (**Supplemental Fig. S9D**), drawing further parallels between the HD-ZIPIII and PC-TP START domains.

We therefore created a new PHB START variant with mutations in seventeen residues analogous to those contacting PC within the PC-TP binding pocket (DLPCmut; **Supplemental Figs. S8, S9E**, Roderick et al., 2002). The substitutions chosen have minimal effects on overall structure (RMSD 0.372; **Figs. 6B**), and include six amino acids with bulkier side chains that partially occlude the ligand binding pocket (**Supplemental Figs. S9F-G**). To test the impact of these mutations, we performed PC-binding assays alongside wild-type, SDmut, and PC-TP START domains after first confirming that all purified START domains have nearly indistinguishable secondary structures (**Figs. 4B, 6C**). Wild-type and PC-TP START domains both showed strong binding to PC-conjugated beads (**Fig. 6F**). By contrast, binding affinity of SDmut START domain was indistinguishable from the MBP negative control, indicating that, in addition to their predicted effects on START conformational change, these substitutions abolish PC binding. Loss of PC-binding capacity was also observed with DLPCmut recombinant START protein (**Fig. 6F**), and

although PHB proteins containing the DLPCmut START domain variant (PHB-DLPCmut) are nuclear-localized in plants, these fail to activate *ZPR3* and *ZPR4* (**Fig. 6D; Supplemental Fig. S9H**). Moreover, no enrichment of PHB-DLPCmut was detected at these loci using ChIP assays (**Fig. 6E**). Taken together, these data propose species of PC as promising candidate ligands for the HD-ZIPIII START domain and show that mutations in amino acids that affect ligand-binding and possibly ligand-mediated conformational change result in loss of HD-ZIPIII DNA-binding competence (**Figs. 4B** and **6E**).

Discussion

HD-ZIPIII TFs are principal regulators of key developmental innovations throughout plant evolution. Although molecularly cloned over twenty years ago (McConnell et al., 2001), the contribution of their START domain remained elusive. We show that the START domain promotes TF homodimerization and increases transcriptional potency. These effects are mediated solely through intramolecular protein changes and can be ported onto other TFs. These findings are particularly intriguing when considered through the lens of TF evolution by domain capture. Basic HD-ZIP proteins and minimal START proteins resembling PC-TP are both present in unicellular algae (Goodstein et al., 2012; Romani et al., 2018), whereas HD-ZIPIII architecture arose after the divergence of *Chlorophyta*, but before the emergence of *Chlorokybus atmophyticus* over 725 million years ago (Wang et al., 2020). Capture of a START domain by HD-ZIPIII antecedents may thus have augmented their transcriptional output. Presumably this also placed their activity under control of a ligand such as PC (**Fig. 6A**). This supposition is consistent with our data. We find that perturbing START domain residues that affect ligand-binding and likely the downstream conformational change abolish DNA-binding competence (**Figs. 5B** and **6E**, Ponting & Aravind, 1999; Tsjuishita et al., 2000; Roderick et al., 2002; Baker et al., 2007). One potential explanation is that the *apo* form of the START domain holds PHB in a non-DNA-binding conformation. Binding of its ligand, or deletion of the START domain entirely, would relieve this inhibition and permit DNA binding. Taken together, these data, as well as the known properties of StARkin domains

(Tsujishita et al., 2000; Roderick et al., 2002; Santiago et al., 2009; Hubbard et al., 2010; Tillman et al., 2020; Dresden et al., 2021), present a model in which the START domain potentiates HD-ZIPIII TF activity, and uses ligand-induced conformational change to render HD-ZIPIII dimers competent to bind DNA (**Fig. 7**).

Given this model, it is intriguing to speculate whether acquisition of a START domain enabled HD-ZIPIII TFs to more effectively integrate signaling inputs into their gene networks, a critical feature of multicellularity[59]. In addition, as phospholipid accumulation is spatially regulated (Okazaki & Saito, 2014; Stanislas et al., 2018), acquisition of the START domain could have allowed HD-ZIPIII activity to become patterned across groups of cells. This would have clear developmental implications as transcripts of ancestral HD-ZIPIII genes are not targeted by small RNAs of the miR166 family (Floyd et al., 2006). Such a regulatory paradigm draws parallels between HD-ZIPIII TFs and mammalian nuclear receptors (Sladek, 2011; Evans & Mangelsdorf), however future experiments are needed to determine whether START ligands play structural roles or indeed contribute to the intricate spatiotemporal regulation of HD-ZIPIII activity. Either way, the properties of the START domain described here suggest a compelling basis for the emergence of HD-ZIPIII TFs as key drivers of plant morphogenic evolution.

The START domain of the closely related HD-ZIPIV family is similarly required for function, and impacts subcellular localization and protein stability, possibly through binding of epidermally-synthesized ceramides (Schrick et al., 2014; Iida et al., 2019; Nagata et al., 2021). The regulatory properties conferred by the HD-ZIPIII START domain are thus distinct, despite their evolutionary relationship (Ponting & Aravind, 1999; Schrick et al., 2004). Further, the modulation of DNA-binding competence identified here represents a new type of StARkin-directed regulatory mechanism. Our findings, in addition to resolving a long-standing mystery in plant development, highlight the flexible and diverse regulatory potential coded within this widely distributed evolutionary module (Dresden et al., 2021).

Materials and Methods

Plant materials and growth conditions. *Arabidopsis thaliana* (Col-0 ecotype) seedlings and *Nicotiana benthamiana* plants were grown at 22°C under long-day conditions, on soil or 1% agarose plates containing Murashige and Skoog medium supplemented with 1% sucrose. Light bulbs were Phillips (F32T8/TL965/ALTO TG 30PK) with a Kelvin temperature of 6500K and lumens of 2600. Inductions were performed by spraying 10-day-old seedlings with 20 uM B-estradiol in 1% DMSO supplemented with 0.005% Silwet.

Molecular biology and plant transformations. *pPHB:PHB-YFP* and *pPHB:PHB*-YFP* constructs have been described previously (Skopelitis et al., 2017). Using these templates, *pPHB:PHB-Delta-YFP* and *pPHB:PHB*-Delta-YFP* were constructed via Gibson assembly (NEB), replacing the 642bp START domain (496-1137bp from the start codon) with a sensitive (GGGATGAAGCCTGG**T**CCGGAT) or an insensitive version (GGGATGAAGCCTGG**A**CCGGAT) of the 21nt miR166-recognition site (Skopelitis et al., 2017). To create *pPHB:PHB-SDmut-YFP*, we first synthesized a mutated variant of the START domain (Mr. Gene), which replaced amino acids RDFTWLR with GAVVGAG and amino acids RAEMK with VAAGV by including the following nucleotide substitutions: CGTGACTTTTGGACGCTGAGA at position 841-861 bp from the start codon to GGTGCCGTCGTAGGAGCAGGC, and AGAGCTGAAATGAAA at position 961-975 bp from the start codon to GTGGCGGCCGGCGTC. miR166-insensitive *pPHB:PHB*-SDmut-YFP* was then constructed via site-directed mutagenesis (Stratagene), copying the mutations in *pPHB:PHB*-YFP* (Skopelitis et al., 2017). All constructs were shuttled into the pB7GW binary vector via Gateway LR reactions (VIB Ghent; Invitrogen).

To facilitate subsequent stable and inducible overexpression in Arabidopsis, and transient expression in tobacco, *PHB*-YFP*, *PHB*-SDmut-YFP*, and *PHB*-Delta-YFP* cDNAs were reamplified and cloned into pCR8-GW via Gibson assembly (Invitrogen; NEB). Gateway LR reactions then shuttled each cDNA into pEARLYGATE100 (stable overexpression), modified pMDC7 with the UBQ10 promoter in place of G10-90 (inducible overexpression), or p502Ω (transient expression; VIB Ghent). *PHB-DLPCmut*-YFP* cDNA was generated by replacing the wild-type START domain with a gene-synthesized mutant variant (GeneArt) using Gibson cloning (ThermoFisher). A Gateway LR reaction then shuttled *PHB-DLPCmut-YFP* into pMDC7. *ATHB12* was amplified from cDNA generated from seedlings, then cloned into pCR8-GW using Gibson (NEB). Fusions between *ATHB12* and the START domain variants were created by inserting a fragment of *PHB** or *PHB-SDmut* (376-1137bp from the start codon) into *ATHB12* (at

position 366 from the start codon) using Gibson assembly (Invitrogen; NEB). Gateway LR reactions then shuttled these cDNAs into p502Ω (Invitrogen; VIB Ghent).

The *pZPR3:3xNLS-RFP* reporter was created by first cloning the 3.2kb region upstream of the *ZPR3* start codon into pCR8-GW (Invitrogen; NEB), followed by an LR reaction using a modified pGREEN binary vector with a Gateway cassette upstream of *3xNLS-RFP* (Invitrogen). The *HD-ZIPI BS:3xNLS-RFP* reporter was created by first synthesizing a construct comprised of six copies of the HD-ZIPI binding site (CAATTATTG) followed by a minimal 35S enhancer, flanked by attL1/attL2 sites (Mr. Gene). A 10-nt spacer (CATTTCAAGA) was inserted between each binding site to minimize potential stearic hindrance. Finally, an LR reaction was used to shuttle these multimerized binding sites into the pGREEN binary described above (Invitrogen). Cloning primer sequences for all constructs are listed in **Supplemental Data Set 7**. Synthesized sequences are in **Supplemental Data Set 8**.

Transient transfection of *N. benthamiana* was performed using syringe-mediated infiltration (Sparkes et al., 2006). In brief, overnight cultures of *Agrobacterium tumefaciens* were centrifuged, resuspended in 2 to 5mL of room temperature infiltration medium (10 mM MgCl2, 10 mM MES KOH, pH 5.6, 150 mM acetosyringone [Sigma Aldrich], and 1% DMSO), diluted to a working optical density of 1, and infiltrated into third and fourth leaves of 3- to 4-week-old *N. benthamiana* plants.

Homology modeling and sequence alignment. The PHB START domain was modeled with I-TASSER (https://zhanggroup.org/I-TASSER/) and AlphaFold2 (https://colab.research.google.com/github/sokrypton/ColabFold/blob/main/AlphaFold2.ipynb) using default parameters. Sequences were aligned using the ClustalW algorithm in the MEGA X software (v.10.1.7; https://www.megasoftware.net/). The crystal structure of PC-TP (1ln1) was the top template used for threading in I-TASSER. Models were visualized in Pymol.

ChIP and RT-qPCR assays. ChIP assays were performed with 11-day-old estradiol-induced seedlings as previously described (Husbands et al., 2015), using IgG (abcam ab46540) or anti-GFP (abcam ab290) antibodies. ChIP and input DNA samples were assayed by qPCR using iQ SYBR Green Supermix (Bio-Rad). *ZPR3* and *ZPR4* regulatory regions assayed in ChIP were selected based on the presence of HD-ZIPIII binding sites predicted by FIMO (https://meme-suite.org/meme/tools/fimo). All experiments were performed at least three independent times. PCR was performed in duplicate, and enrichments calculated relative to input. Student's *t*-test was used to calculate statistical significance.

Total RNA was extracted from seedlings or infiltrated *N. benthamiana* leaves using Trizol reagent (Gibco BRL). One microgram of RNA was primed with oligo (dT) and reverse transcribed using the SuperScript III first-strand synthesis kit (Invitrogen). Relative quantification values were calculated based on at least three biological replicates, with ΔCt of ACT2 or B-tubulin serving as normalization controls in Arabidopsis or *N. benthamiana*, respectively. Wild-type or uninduced values were set to one and PHB variant values either plotted directly or after further normalization to PHB variant levels. Student's *t*-test was used to calculate statistical significance. ChIP and RT-qPCR primer sequences are listed in **Supplemental Data Set 9**.

Plant imaging. Brightfield images of Arabidopsis seedlings were captured using an SMZ1500 dissecting microscope with NIS Element software (Nikon). Fluorescent images of Arabidopsis seedlings and infiltrated *N. benthamiana* leaves were obtained using the same microscope with the P-FLA2 epi-fluorescent attachment. Heart-stage embryos were dissected, stained with Fluorescent Brightener 28 (Sigma Aldrich), and then imaged using an LSM780 confocal microscope (Zeiss).

Single-molecule imaging and stoichiometric analyses. A detailed protocol of SiMPull with plant tissue has been published (Husbands et al., 2016), however the amount of input tissue can vary between experiments. Here, lysates for SiMPull were prepared from flash frozen tissue comprised of five-to-six 10-day-old Arabidopsis seedlings or 1-2 cm^2 pieces of infiltrated *N. benthamiana* leaves. Note: SiMPull experiments in Arabidopsis were conducted using seedlings expressing PHB variants under native regulatory elements (**Figs. 2A-B**) as well as an estradiol-inducible promoter (**Supplemental Fig. S3D**). A minimum of three independent biological replicates were performed for each SiMPull experiment. To enable calculation of PHB variant dimerization frequencies, as well as the maturation probability of citrine YFP in Arabidopsis, *2x35S:monoYFP* and *2x35S:tdYFP* constructs (described in [18]) were stably transformed and at least four independent lines analyzed.

SiMPull was first performed with 1:150 dilutions of lysates from *2x35S:monoYFP* or *2x35S:tdYFP* seedlings, mixed at ratios described in **Supplemental Fig. S2**. Frequencies of two-step photobleaching events were then scored and used to construct a standard calibration curve ($y = 2.28x - 27$; $R^2 = 0.992$). For stoichiometric analyses of PHB variants, lysates from native promoter lines or 24 hrs estradiol-induced lines were diluted 1:1 or 1:100, respectively, then subject to SiMPull and photobleaching counts. Frequencies of two-step photobleaching events for each genotype were translated into dimerization frequencies via the above calibration curve. Maturation probability was predicted using binomial probability modeling (described in Husbands et al., 2016). Co-localization data for ZPR3-mCherry with

AS2-YFP, PHB-YFP, PHB-SDmut-YFP, or PHB-Delta-YFP were collected and analyzed as described previously (Husbands et al., 2016).

Quantification of PHB variant induction with estradiol. Ten-to-twelve 10-day-old seedlings were induced for 24 hrs with estradiol, flash frozen, then ground in 500 µl freshly prepared lysis buffer (25 mM Tris HCl pH 8, 150 mM NaCl, 1% SDS, 1x cOmplete ULTRA protease inhibitor (Roche), and 1x PhosSTOP (Roche)). Lysates were cleared via 14000x g centrifugation at 4°C, and split evenly for RNA vs protein processing. RNA extraction, cDNA synthesis, and RT-qPCR was carried out as described above. Lysates for protein work were mixed 1:1 with 2x Laemmli sample buffer, and boiled for 1 min. Proteins were resolved via SDS-PAGE, blotted to Hybond ECL membrane (GE Healthcare), and blocked in 5% milk fat. PHB variants were detected via anti-GFP primary antibodies (600-406-215; Rockland Immunochemicals; 1:500 dilution) and anti-rabbit horseradish peroxidase-conjugated secondary antibodies (Jackson Immunoresearch; 1:5000 dilution). Detection of secondary antibodies was performed with SuperSignal West Pico Chemiluminescent Substrate (ThermoFisher Scientific). Blots were scanned and quantified using ImageJ. Four independent biological replicates were performed, each with two technical replicates.

Protein immunoprecipitation and mass spectrometry. One gram of 24 hrs estradiol-induced seedlings was flash frozen, ground under liquid nitrogen, and extracted in 4 ml of freshly prepared Extraction Buffer (50 mM HEPES pH 8, 150 mM NaCl, 0.3% NP-40, 1x cOmplete ULTRA protease inhibitor -EDTA (Roche), 1mM PMSF, 50 uM bortezomib). Lysates were cleared via 14000x g centrifugation at 4°C, pre-cleared with Protein A Dynabeads, and incubated for 1 h with 5µl anti-GFP antibody (abcam ab290). PHB variant complexes were captured using 25µl Protein A Dynabeads and washed 4x with freshly prepared Wash Buffer (50mM HEPES pH 8, 450mM NaCl, 0.1% NP-40, 1x cOmplete ULTRA protease inhibitor -EDTA (Roche), 1mM PMSF, 50 µM bortezomib).

After the final wash, beads were flash frozen in liquid nitrogen and trypsinized via standard protocols (Promega). Peptides were labeled with 8-plex iTRAQ (Ross et al., 2004), and injected into the Orbitrap Velos Pro mass spectrometer. Protein identification and quantification was carried using Mascot 2.4 (Perkins et al., 1999) against the UniProt Arabidopsis sequence database. Enrichments over the uninduced control were calculated for each replicate, mean enrichment was then calculated across the top five replicates, and a Student's *t*-test applied to determine significance. Known false positives were eliminated from subsequent analyses (Van Leene et al., 2015). Interacting partners for PHB-SDmut and PHB-Delta were identified using a similar workflow. Raw data is included in **Supplemental Data Set 10**.

Protein purification, circular dichroism, membrane-overlay, and PC-conjugated bead assays. Wild-type or START mutant variants (496-1137bp from the start codon) were cloned downstream of maltose binding protein (MBP), in a modified pET28b vector (Laha et al., 2015), via Gibson cloning (ThermoFisher). Proteins were induced with 100 µM IPTG at 12ºC for 16 hrs and purified on a Ni^{2+} column according to manufacturer protocols (Qiagen). Proteins were eluted using 125 mM imidazole, desalted and concentrated on a 10 MWCO column (Amersham) into NaCl-free IP buffer (1x PBS pH7.4, 5% glycerol), and quantified next to BSA on a 10% SDS gel.

Circular dichroism was performed with the Jasco J-815 Spectrometer (OSU Biophysical Interaction & Characterization Facility) using 1mg/ml of purified MBP or MBP-START variant proteins, then corrected and converted to molar ellipticity in Excel (Microsoft). Scans were limited to a range of 190-250nm to increase resolution and data were plotted in GraphPad (Prism).

Membrane-overlay assays were done according to manufacturer protocols (ThermoFisher Scientific; P23751). In brief, membranes were blocked for 1h at RT (PBS, 0.1% Tween20, 3% BSA fatty acid free). 150 µg of wild-type START purified protein was added to the membrane in 5ml blocking buffer and incubated for 1h with gentle shaking. Membranes were washed three times with PBS-T and incubated with 1:2000 anti-His-tag antibody (Abiocode M0335-1) for 1h with gentle shaking at RT. Membranes were washed three times and incubated with 1:2000 anti-mouse 2º antibody (ab6789) for 1h. Bound proteins were detected using Hyper HRP Substrate (Takara). PtIns(3,5)P_2 Grip protein (P-3516-3-EC, MoBiTec), detected via anti-GST antibody (Sigma Aldrich, A7340), served as a positive control (not shown).

PC-conjugated bead assays were done according to manufacturer protocols (P-B0PC; Echelon Biosciences) except that 0.1% IGEPAL was used in Buffer System 1. In brief, 1 nmol of purified START domain protein was added to 0.25 nmols of beads equilibrated in Buffer System 1. Protein-bead mixes were rotated at room temperature for 15 mins, then transferred to 4ºC for continued capture overnight. Protein-bead mixes were washed 3x using 250ul Buffer System 1, eluted using 1x Laemmli buffer, and bound proteins separated on an SDS-PAGE gel. Bound proteins and 1:1000 diluted Inputs were detected by western blotting using a 1:1000 HRP-conjugated 6x-His Tag antibody (PA1-23024; ThermoFisher).

Liposome production and dynamic light scattering

Total lipids from 35-40g of 21d old Arabidopsis seedling tissue were extracted using Bligh-Dyer (Bligh & Dyer, 1959). Lipids were dried under argon and resuspended in 100% ethanol to 100 mg/ml. Liposomes were generated by rapid injection of ethanol-suspended lipids into IP buffer (1x PBS pH7.4, 150 mM NaCl, 5% glycerol). Liposome size range was determined by dynamic light scattering using the Zetasizer Nano (Malvern).

Lipid immunoprecipitations and mass spectrometry

To identify lipids bound by the START domain in bacteria, lipids from purified recombinant MBP and MBP-START proteins were immediately extracted using Bligh & Dyer (Bligh & Dyer, 1959), dried under argon, and resuspended in chloroform. Lipids were identified using LC-MS performed at the Institut fur Molekulare Physiologie und Biotechnologie der Pflanze (IMBIO) at the University of Bonn, Germany. In brief, lipids were resuspended in 200ul Q-ToF solvent was added (methanol/chloroform/300 mM ammonium acetate, 665:300:35, v/v/v, Welti et al., 2002), and measured using nanoflow direct infusion Q-TOF MS/MS (as described in Gasulla et al., 2013).

To identify lipids bound by the START domain in plant lipid mixtures, 1 mg of recombinant MBP or MBP-START proteins were incubated with 6 mg of liposomes in IP buffer (1x PBS pH 7.4, 150mM NaCl, 5% glycerol) overnight at 4 degrees. IPs were performed in quintuplicate. Proteins were repurified using nickel affinity chromatography, extensively washed, and eluted using 125 mM imidazole. Lipids were extracted using Bligh & Dyer (Bligh & Dyer, 1959), dried under argon, and resuspended in chloroform. Lipids were identified using LC-MS performed at the Kansas Lipidomics Research Center Analytical Laboratory (KLRC; Kansas State University) and the Nutrient & Phytochemical Analytics Shared Resource (NPASR; Ohio State Comprehensive Cancer Center). KLRC used a Waters Xevo TQS mass spectrometer adjusted for SPLASH response factors. 400 ul volumes were injected, ES+ ionization mode was used for all detection of all compounds except lysoPG which used ES-, and data presented are nmol per mg dry weight and normalized to mols of protein per IP. NPASR data was using a SelexION/QTrap 5500 (Sciex) Lipidyzer for CER, LPC, LPE, PC, PE, and SM and a 6550 QTOF MS (Agilent) for FFAs. 150 ul volumes were injected, ESI- ionization mode was used, and data presented are normalized to mols of protein per IP.

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Accession numbers

PHABULOSA – AT2G34710; PHAVOLUTA – AT1G30490; REVOLUTA – AT5G60690; CORONA - AT1G52150; ATHB8 – AT4G32880; LITTLE ZIPPER3 – AT3G52770; LITTLE ZIPPER4 – AT2G36307; ATHB12 – AT3G61890; OLEOSIN1 – AT4G25140; DYNAMIN-RELATED PROTEIN 1C – AT1G14830; DYNAMIN-RELATED PROTEIN 1E – AT3G60190; ANNEXIN 4 – AT2G38750; ORNITHINE TRANSCARBAMYLASE (AT1G75330).

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Author Contributions

A.Y.H and M.T conceived of the research direction and experiments; A.Y.H. performed experiments except: protein purification by A.F., LC-MS by KLRC and NPASR centers, additional ChIP and protein work by C.E.D. and A.S.H, PC-binding assays by C.E.D., and SiMPull by V.A. under the guidance of T.H. A.Y.H and M.T. wrote the manuscript.

Competing interests

The authors declare that they have no competing interests.

References

Aggarwal, V., & Ha, T. (2014). Single-molecule pull-down (SiMPull) for new-age biochemistry: methodology and biochemical applications of single-molecule pull-down (SiMPull) for probing biomolecular interactions in crude cell extracts. *BioEssays*, *36*(11), 1109–1119.

Alpy, F., & Tomasetto, C. (2005). Give lipids a START: the StAR-related lipid transfer (START) domain in mammals. *Journal of cell science*, *118*(Pt 13), 2791–2801.

Ariel, F.D., Manavella, P. A., Dezar, C.A., and Chan, R.L. (2007). The true story of the HD-Zip family. *Trends Plant Sci.*, *12*, 419-26.

Baker, B. Y., Epand, R. F., Epand, R. M., & Miller, W. L. (2007). Cholesterol binding does not predict activity of the steroidogenic acute regulatory protein, StAR. *The Journal of biological chemistry*, *282*(14), 10223–10232.

Belda-Palazon, B., Gonzalez-Garcia, M. P., Lozano-Juste, J., Coego, A., Antoni, R., Julian, J., Peirats-Llobet, M., Rodriguez, L., Berbel, A., Dietrich, D., Fernandez, M. A., Madueño, F., Bennett, M. J., & Rodriguez, P. L. (2018). PYL8 mediates ABA perception in the root through non-cell-autonomous and

ligand-stabilization-based mechanisms. *Proceedings of the National Academy of Sciences of the United States of America*, *115*(50), E11857–E11863.

Bligh, E.G. & Dyer, W.J. (1959). A rapid method of total lipid extraction and purification. *Canadian journal of biochemistry and physiology*, *37*(8), 911–917.

Carlsbecker, A., Lee, J. Y., Roberts, C. J., Dettmer, J., Lehesranta, S., Zhou, J., Lindgren, O., Moreno-Risueno, M. A., Vatén, A., Thitamadee, S., Campilho, A., Sebastian, J., Bowman, J. L., Helariutta, Y., & Benfey, P. N. (2010). Cell signalling by microRNA165/6 directs gene dose-dependent root cell fate. *Nature*, *465*(7296), 316–321.

de Brouwer, A. P., Bouma, B., van Tiel, C. M., Heerma, W., Brouwers, J. F., Bevers, L. E., Westerman, J., Roelofsen, B., & Wirtz, K. W. (2001). The binding of phosphatidylcholine to the phosphatidylcholine transfer protein: affinity and role in folding. *Chemistry and physics of lipids*, *112*(2), 109–119.

de Mendoza, A. et al. Transcription factor evolution in eukaryotes and the assembly of the regulatory toolkit in multicellular lineages. *Proc Natl Acad Sci USA*. **110**, E4858-66. (2013).

Dresden, C. E., Ashraf, Q., & Husbands, A. Y. (2021). Diverse regulatory mechanisms of StARkin domains in land plants and mammals. *Current opinion in plant biology*, *64*, 102148.

Du, X., Qian, X., Papageorge, A., Schetter, A. J., Vass, W. C., Liu, X., Braverman, R., Robles, A. I., & Lowy, D. R. (2012). Functional interaction of tumor suppressor DLC1 and caveolin-1 in cancer cells. *Cancer research*, *72*(17), 4405–4416.

Evans, R. M., & Mangelsdorf, D. J. (2014). Nuclear Receptors, RXR, and the Big Bang. *Cell*, *157*(1), 255–266.

Floyd, S. K., Zalewski, C. S., & Bowman, J. L. (2006). Evolution of class III homeodomain-leucine zipper genes in streptophytes. *Genetics*, *173*(1), 373–388.

Fujii, H., Chinnusamy, V., Rodrigues, A., Rubio, S., Antoni, R., Park, S. Y., Cutler, S. R., Sheen, J., Rodriguez, P. L., & Zhu, J. K. (2009). In vitro reconstitution of an abscisic acid signalling pathway. *Nature*, *462*(7273), 660–664.

Gasulla, F., Vom Dorp, K., Dombrink, I., Zähringer, U., Gisch, N., Dörmann, P., & Bartels, D. (2013). The role of lipid metabolism in the acquisition of desiccation tolerance in Craterostigma plantagineum: a comparative approach. *The Plant journal*, *75*(5), 726–741

Gatta, A. T., Wong, L. H., Sere, Y. Y., Calderón-Noreña, D. M., Cockcroft, S., Menon, A. K., & Levine, T. P. (2015). A new family of StART domain proteins at membrane contact sites has a role in ER-PM sterol transport. *eLife*, *4*, e07253.

Goodstein, D. M., Shu, S., Howson, R., Neupane, R., Hayes, R. D., Fazo, J., Mitros, T., Dirks, W., Hellsten, U., Putnam, N., & Rokhsar, D. S. (2012). Phytozome: a comparative platform for green plant genomics. *Nucleic acids research 40*, D1178–D1186.

Hubbard, K. E., Nishimura, N., Hitomi, K., Getzoff, E. D., & Schroeder, J. I. (2010). Early abscisic acid signal transduction mechanisms: newly discovered components and newly emerging questions. *Genes & development*, *24*(16), 1695–1708.

Husbands, A. Y., Aggarwal, V., Ha, T., & Timmermans, M. C. (2016). In Planta Single-Molecule Pull-Down Reveals Tetrameric Stoichiometry of HD-ZIPIII:LITTLE ZIPPER Complexes. *The Plant Cell*, *28*(8), 1783–1794.

Husbands, A. Y., Benkovics, A. H., Nogueira, F. T., Lodha, M., & Timmermans, M. C. (2015). The ASYMMETRIC LEAVES Complex Employs Multiple Modes of Regulation to Affect Adaxial-Abaxial Patterning and Leaf Complexity. *The Plant cell*, *27*(12), 3321–3335.

Iida, H., Yoshida, A., & Takada, S. (2019). *ATML1* activity is restricted to the outermost cells of the embryo through post-transcriptional repressions. *Development*, *146*(4), dev169300

Jain, A., Arauz, E., Aggarwal, V., Ikon, N., Chen, J., & Ha, T. (2014). Stoichiometry and assembly of mTOR complexes revealed by single-molecule pulldown. *Proceedings of the National Academy of Sciences of the United States of America*, *111*(50), 17833–17838.

Jain, A., Liu, R., Ramani, B., Arauz, E., Ishitsuka, Y., Ragunathan, K., Park, J., Chen, J., Xiang, Y. K., & Ha, T. (2011). Probing cellular protein complexes using single-molecule pull-down. *Nature*, *473*(7348), 484–488.

Jarvela, M.C. & Hinman, V.G. Evolution of transcription factor function as a mechanism for changing metazoan developmental gene regulatory networks. *Evodevo*. **6**, 3 (2015).

Juarez, M. T., Kui, J. S., Thomas, J., Heller, B. A., & Timmermans, M. C. (2004). microRNA-mediated repression of rolled leaf1 specifies maize leaf polarity. *Nature*, *428*(6978), 84–88.

Kanno, K., Wu, M. K., Agate, D. S., Fanelli, B. J., Wagle, N., Scapa, E. F., Ukomadu, C., & Cohen, D. E. (2007). Interacting proteins dictate function of the minimal START domain phosphatidylcholine transfer protein/StarD2. *The Journal of biological chemistry*, *282*(42), 30728–30736.

Kelley, D. R., Skinner, D. J., & Gasser, C. S. (2009). Roles of polarity determinants in ovule development. *The Plant Journal*, *57*(6), 1054–1064.

Kim, Y. S., Kim, S. G., Lee, M., Lee, I., Park, H. Y., Seo, P. J., Jung, J. H., Kwon, E. J., Suh, S. W., Paek, K. H., & Park, C. M. (2008). HD-ZIP III activity is modulated by competitive inhibitors via a feedback loop in Arabidopsis shoot apical meristem development. *The Plant Cell*, *20*(4), 920–933.

Laha, D., Johnen, P., Azevedo, C., Dynowski, M., Weiß, M., Capolicchio, S., Mao, H., Iven, T., Steenbergen, M., Freyer, M., Gaugler, P., de Campos, M. K., Zheng, N., Feussner, I., Jessen, H. J., Van Wees, S. C., Saiardi, A., & Schaaf, G. (2015). VIH2 Regulates the Synthesis of Inositol Pyrophosphate InsP8 and Jasmonate-Dependent Defenses in Arabidopsis. *The Plant cell*, *27*(4), 1082–1097.

Lee, Y. H., Oh, H. S., Cheon, C. I., Hwang, I. T., Kim, Y. J., & Chun, J. Y. (2001). Structure and expression of the Arabidopsis thaliana homeobox gene Athb-12. *Biochemical and biophysical research communications*, *284*(1), 133–141.

Lumba, S., Cutler, S., & McCourt, P. (2010). Plant nuclear hormone receptors: a role for small molecules in protein-protein interactions. *Annual review of cell and developmental biology*, *26*, 445–469.

Mallory, A. C., Reinhart, B. J., Jones-Rhoades, M. W., Tang, G., Zamore, P. D., Barton, M. K., & Bartel, D. P. (2004). MicroRNA control of PHABULOSA in leaf development: importance of pairing to the microRNA 5' region. *The EMBO journal*, *23*(16), 3356–3364.

McConnell, J. R., Emery, J., Eshed, Y., Bao, N., Bowman, J., & Barton, M. K. (2001). Role of PHABULOSA and PHAVOLUTA in determining radial patterning in shoots. *Nature*, *411*(6838), 709–713.

Mikhailov, K. V., Konstantinova, A. V., Nikitin, M. A., Troshin, P. V., Rusin, L. Y., Lyubetsky, V. A., Panchin, Y. V., Mylnikov, A. P., Moroz, L. L., Kumar, S., & Aleoshin, V. V. (2009). The origin of Metazoa: a transition from temporal to spatial cell differentiation. *BioEssays*, *31*(7), 758–768.

Nagata, K., Ishikawa, T., Kawai-Yamada, M., Takahashi, T., & Abe, M. (2021). Ceramides mediate positional signals in *Arabidopsis thaliana* protoderm differentiation. *Development*, *148*(2), dev194969.

Okazaki, Y., & Saito, K. (2014). Roles of lipids as signaling molecules and mitigators during stress response in plants. *The Plant journal*, *79*(4), 584–596.

Park, S. Y., Fung, P., Nishimura, N., Jensen, D. R., Fujii, H., Zhao, Y., Lumba, S., Santiago, J., Rodrigues, A., Chow, T. F., Alfred, S. E., Bonetta, D., Finkelstein, R., Provart, N. J., Desveaux, D., Rodriguez, P. L., McCourt, P., Zhu, J. K., Schroeder, J. I., Volkman, B. F., … Cutler, S. R. (2009). Abscisic acid inhibits type 2C protein phosphatases via the PYR/PYL family of START proteins. *Science*, *324*(5930), 1068–1071.

Perkins, D. N., Pappin, D. J., Creasy, D. M., & Cottrell, J. S. (1999). Probability-based protein identification by searching sequence databases using mass spectrometry data. *Electrophoresis*, *20*(18), 3551–3567.

Ponting, C. P., & Aravind, L. (1999). START: a lipid-binding domain in StAR, HD-ZIP and signalling proteins. *Trends in biochemical sciences*, *24*(4), 130–132.

Prashek, J., Bouyain, S., Fu, M., Li, Y., Berkes, D., & Yao, X. (2017). Interaction between the PH and START domains of ceramide transfer protein competes with phosphatidylinositol 4-phosphate binding by the PH domain. *The Journal of biological chemistry*, *292*(34), 14217–14228.

Prigge, M. J., Otsuga, D., Alonso, J. M., Ecker, J. R., Drews, G. N., & Clark, S. E. (2005). Class III homeodomain-leucine zipper gene family members have overlapping, antagonistic, and distinct roles in Arabidopsis development. *The Plant Cell*, *17*(1), 61–76.

Ramachandran, P., Carlsbecker, A., & Etchells, J. P. (2017). Class III HD-ZIPs govern vascular cell fate: an HD view on patterning and differentiation. *Journal of experimental botany*, *68*(1), 55–69.

Rhoades, M. W., Reinhart, B. J., Lim, L. P., Burge, C. B., Bartel, B., & Bartel, D. P. (2002). Prediction of plant microRNA targets. Cell, 110(4), 513–520.

Robischon, M., Du, J., Miura, E., & Groover, A. (2011). The Populus class III HD ZIP, popREVOLUTA, influences cambium initiation and patterning of woody stems. *Plant physiology*, *155*(3), 1214–1225.

Roderick, S. L., Chan, W. W., Agate, D. S., Olsen, L. R., Vetting, M. W., Rajashankar, K. R., & Cohen, D. E. (2002). Structure of human phosphatidylcholine transfer protein in complex with its ligand. *Nature structural biology*, *9*(7), 507–511.

Romani, F., Reinheimer, R., Florent, S. N., Bowman, J. L., & Moreno, J. E. (2018). Evolutionary history of HOMEODOMAIN LEUCINE ZIPPER transcription factors during plant transition to land. *The New phytologist*, *219*(1), 408–421.

Ross, P. L., Huang, Y. N., Marchese, J. N., Williamson, B., Parker, K., Hattan, S., Khainovski, N., Pillai, S., Dey, S., Daniels, S., Purkayastha, S., Juhasz, P., Martin, S., Bartlet-Jones, M., He, F., Jacobson, A., & Pappin, D. J. (2004). Multiplexed protein quantitation in Saccharomyces cerevisiae using amine-reactive isobaric tagging reagents. *Molecular & cellular proteomics*, *3*(12), 1154–1169.

Sablin, E. P., Blind, R. D., Krylova, I. N., Ingraham, J. G., Cai, F., Williams, J. D., Fletterick, R. J., & Ingraham, H. A. (2009). Structure of SF-1 bound by different phospholipids: evidence for regulatory ligands. *Molecular endocrinology*, *23*(1), 25–34.

Sanchez-Solana, B., Wang, D., Qian, X., Velayoudame, P., Simanshu, D. K., Acharya, J. K., & Lowy, D. R. (2021). The tumor suppressor activity of DLC1 requires the interaction of its START domain with Phosphatidylserine, PLCD1, and Caveolin-1. *Molecular cancer*, *20*(1), 141.

Santiago, J., Dupeux, F., Round, A., Antoni, R., Park, S. Y., Jamin, M., Cutler, S. R., Rodriguez, P. L., & Márquez, J. A. (2009). The abscisic acid receptor PYR1 in complex with abscisic acid. *Nature*, *462*(7273), 665–668.

Schrick, K., Bruno, M., Khosla, A., Cox, P. N., Marlatt, S. A., Roque, R. A., Nguyen, H. C., He, C., Snyder, M. P., Singh, D., & Yadav, G. (2014). Shared functions of plant and mammalian StAR-related lipid transfer (START) domains in modulating transcription factor activity. *BMC biology*, *12*, 70.

Schrick, K., Nguyen, D., Karlowski, W. M., & Mayer, K. F. (2004). START lipid/sterol-binding domains are amplified in plants and are predominantly associated with homeodomain transcription factors. *Genome biology*, *5*(6), R41.

Sebastian, J., Ryu, K. H., Zhou, J., Tarkowská, D., Tarkowski, P., Cho, Y. H., Yoo, S. D., Kim, E. S., & Lee, J. Y. (2015). PHABULOSA controls the quiescent center-independent root meristem activities in Arabidopsis thaliana. *PLoS Genetics*, *11*(3), e1004973.

Sebé-Pedrós, A., Degnan, B.M. & Ruiz-Trillo, I. The origin of Metazoa: a unicellular perspective. *Nat Rev Genet.*, **18**, 498-512 (2017).

Sessa, G., Steindler, C., Morelli, G., & Ruberti, I. (1998). The Arabidopsis Athb-8, -9 and -14 genes are members of a small gene family coding for highly related HD-ZIP proteins. *Plant molecular biology*, *38*(4), 609–622.

Skopelitis, D. S., Benkovics, A. H., Husbands, A. Y., & Timmermans, M. (2017). Boundary Formation through a Direct Threshold-Based Readout of Mobile Small RNA Gradients. *Developmental Cell*, *43*(3), 265–273.e6.

Sladek F. M. (2011). What are nuclear receptor ligands?. *Molecular and cellular endocrinology*, *334*(1-2), 3–13.

Smetana, O., Mäkilä, R., Lyu, M., Amiryousefi, A., Sánchez Rodríguez, F., Wu, M. F., Solé-Gil, A., Leal Gavarrón, M., Siligato, R., Miyashima, S., Roszak, P., Blomster, T., Reed, J. W., Broholm, S., &

Mähönen, A. P. (2019). High levels of auxin signalling define the stem-cell organizer of the vascular cambium. *Nature*, *565*(7740), 485–489.

Sparkes, I. A., Runions, J., Kearns, A., & Hawes, C. (2006). Rapid, transient expression of fluorescent fusion proteins in tobacco plants and generation of stably transformed plants. *Nature protocols*, *1*(4), 2019–2025.

Stanislas, T., Platre, M. P., Liu, M., Rambaud-Lavigne, L., Jaillais, Y., & Hamant, O. (2018). A phosphoinositide map at the shoot apical meristem in Arabidopsis thaliana. *BMC biology*, *16*(1), 20.

Tillman, M. C., Imai, N., Li, Y., Khadka, M., Okafor, C. D., Juneja, P., Adhiyaman, A., Hagen, S. J., Cohen, D. E., & Ortlund, E. A. (2020). Allosteric regulation of thioesterase superfamily member 1 by lipid sensor domain binding fatty acids and lysophosphatidylcholine. *Proceedings of the National Academy of Sciences of the United States of America*, *117*(36), 22080–22089.

Tsujishita, Y., & Hurley, J. H. (2000). Structure and lipid transport mechanism of a StAR-related domain. *Nature structural biology*, *7*(5), 408–414.

V. J. Lynch, G. P. Wagner, Resurrecting the role of transcription factor change in developmental evolution. *Evolution*. **9**, 2131-54 (2008).

Van Leene, J., Eeckhout, D., Cannoot, B., De Winne, N., Persiau, G., Van De Slijke, E., Vercruysse, L., Dedecker, M., Verkest, A., Vandepoele, K., Martens, L., Witters, E., Gevaert, K., & De Jaeger, G. (2015). An improved toolbox to unravel the plant cellular machinery by tandem affinity purification of Arabidopsis protein complexes. *Nature protocols*, *10*(1), 169–187.

Wang, S., Li, L., Li, H., Sahu, S. K., Wang, H., Xu, Y., Xian, W., Song, B., Liang, H., Cheng, S., Chang, Y., Song, Y., Çebi, Z., Wittek, S., Reder, T., Peterson, M., Yang, H., Wang, J., Melkonian, B., Van de Peer, Y., … Liu, X. (2020). Genomes of early-diverging streptophyte algae shed light on plant terrestrialization. *Nature plants*, *6*(2), 95–106.

Welti, R., Li, W., Li, M., Sang, Y., Biesiada, H., Zhou, H. E., Rajashekar, C. B., Williams, T. D., & Wang, X. (2002). Profiling membrane lipids in plant stress responses. Role of phospholipase D alpha in freezing-induced lipid changes in Arabidopsis. *The Journal of biological chemistry*, *277*(35), 31994–32002.

Wenkel, S., Emery, J., Hou, B. H., Evans, M. M., & Barton, M. K. (2007). A feedback regulatory module formed by LITTLE ZIPPER and HD-ZIPIII genes. *The Plant Cell*, *19*(11), 3379–3390.

Wong, L. H., & Levine, T. P. (2016). Lipid transfer proteins do their thing anchored at membrane contact sites… but what is their thing?. *Biochemical Society transactions*, *44*(2), 517–527.

Xu, Q., Li, R., Weng, L., Sun, Y., Li, M., & Xiao, H. (2019). Domain-specific expression of meristematic genes is defined by the LITTLE ZIPPER protein DTM in tomato. *Communications biology*, *2*, 134.

Yin, P., Fan, H., Hao, Q., Yuan, X., Wu, D., Pang, Y., Yan, C., Li, W., Wang, J., & Yan, N. (2009). Structural insights into the mechanism of abscisic acid signaling by PYL proteins. *Nature structural & molecular biology*, *16*(12), 1230–1236.

Yip, H. K., Floyd, S. K., Sakakibara, K., & Bowman, J. L. (2016). Class III HD-Zip activity coordinates leaf development in Physcomitrella patens. *Developmental biology*, *419*(1), 184–197.

Zhai, G., Song, J., Shu, T., Yan, J., Jin, X., He, J., & Yin, Z. (2017). LRH-1 senses signaling from phosphatidylcholine to regulate the expansion growth of digestive organs via synergy with Wnt/β-catenin signaling in zebrafish. *Journal of genetics and genomics*, *44*(6), 307–317.

Zhang, L., Li, X., Li, D., Sun, Y., Li, Y., Luo, Q., Liu, Z., Wang, J., Li, X., Zhang, H., Lou, Z., & Yang, Y. (2018). CARK1 mediates ABA signaling by phosphorylation of ABA receptors. *Cell discovery*, *4*, 30.

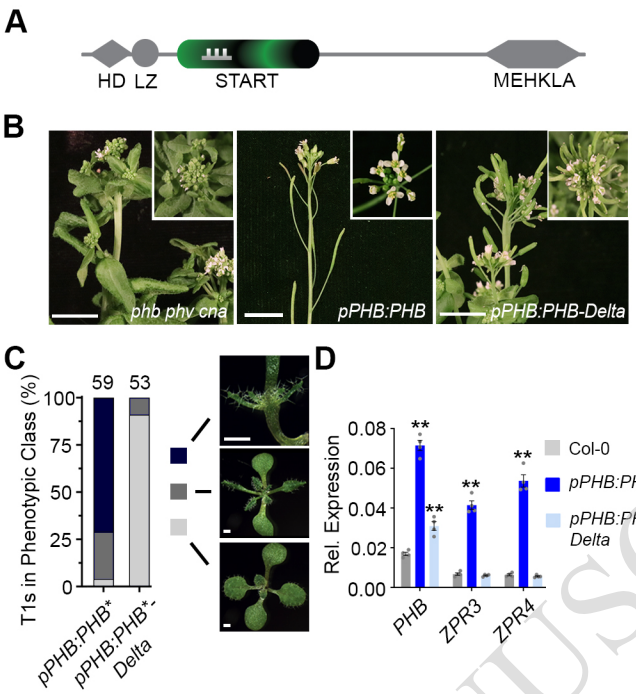

Fig 1. The START domain is required for full PHB function. A, General structure of HD-ZIPIII proteins. Note PHB-Delta retains the miR166-binding site (grey comb) contained within the START domain (green). **B**, The pleiotropic *phb phv cna* triple mutant phenotype has been previously described[9] and includes fasciated stems and abnormal floral organs (left). The *pPHB:PHB* transgene complements the mutant phenotype and plants appear wild-type (middle), whereas *pPHB:PHB-Delta* plants do not (right). **C**, Phenotypic scoring of primary transformants (*n* above bar) carrying miR166-insensitive (*) *pPHB:PHB* or *pPHB:PHB-Delta* constructs. Ectopic accumulation of PHB leads to severe (black) or intermediate (dark grey) gain-of-function phenotypes, whereas plants mis-accumulating PHB-Delta appear wild-type (light grey). **D**, Ectopic *PHB** expression leads to strong upregulation of *ZPR3* and *ZPR4* targets. By contrast, *ZPR3* or *ZPR4* levels are indistinguishable from Col-0 in *pPHB:PHB*-Delta* lines, despite accumulation of this variant above endogenous *PHB* levels. Note: higher PHB transcript levels in *pPHB:PHB** could reflect either a PHB auto-activation mechanism (proposed in [10]) or the greater relative proportion of adaxialized tissues in *pPHB:PHB** seedlings (versus *pPHB:PHB*-Delta)*. *n* = 3 biological replicates. ***P* ≤ 0.01, Student's *t*-test.

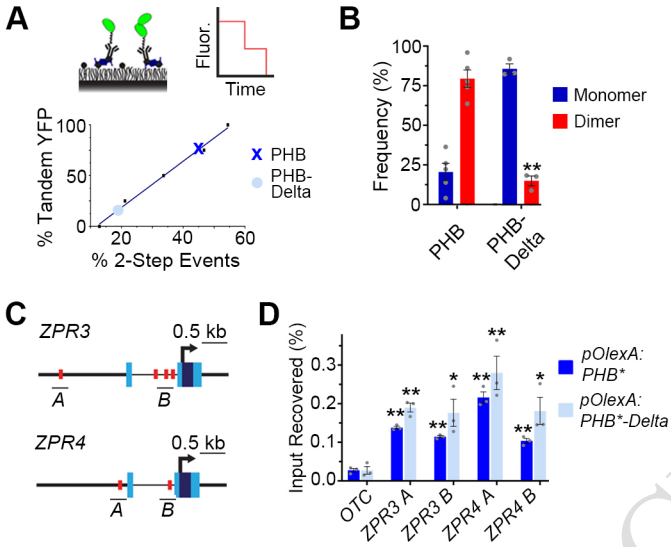

Fig 2. The START domain affects PHB homodimerization. A, SiMPull calibration curve (bottom graph) for monomeric vs dimeric YFP (top left cartoon) translates percentage of two-step photobleaching events (simulated in top right cartoon) to frequency of dimers in the population. PHB (**X**) and PHB-Delta (●) show two-step photobleaching events of 47% and 20%, respectively. **B**, Percentages of two-step photobleaching events translate to dimerization frequencies of ~80% for PHB and ~15% for PHB-Delta. **C**, Schematic representations of *ZPR3* and *ZPR4* showing HD-ZIPIII binding sites identified by FIMO (red boxes), ChIP amplicons (black bars), transcription start sites (arrow), untranslated regions (light-blue boxes), and exons (dark-blue boxes). **D**, PHB and PHB-Delta occupy multiple sites in the regulatory regions of *ZPR3* and *ZPR4* and are significantly enriched over the *ORNITHINE TRANSCARBAMYLASE (OTC)* negative control locus. $n = 3$ biological replicates. $^{*}P \leq 0.05$, $^{**}P \leq 0.01$, Student's *t*-test.

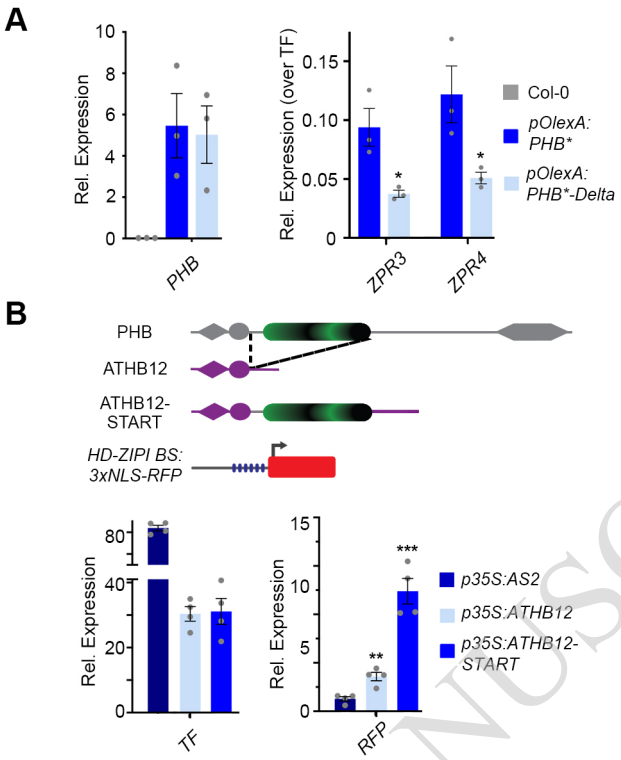

Fig 3. The START domain enhances PHB transcriptional potency and this property is transferable to heterologous TFs. **A**, Relative *ZPR3* and *ZPR4* transcript levels in 24 hrs estradiol-induced *pOlexA:PHB** and *pOlexA:PHB*-Delta* seedlings indicate PHB-Delta is a less-potent transcriptional activator than PHB. **B**, Schematic representation of domain-capture-mimicry constructs detailing insertion of the PHB START domain (green) into ATHB12 (purple) and the design of the *HD-ZIPI BS:3xNLS-RFP* target reporter (top). RT-qPCR following co-transfection with the *HD-ZIPI BS:3xNLS-RFP* target reporter (bottom), shows fusion of the wild-type PHB START domain augments ATHB12 transcriptional potency by a factor of three (bottom). AS2 is included as negative control. *n* = 3 biological replicates. **P* ≤ 0.05, ***P* ≤ 0.01, Student's *t*-test.

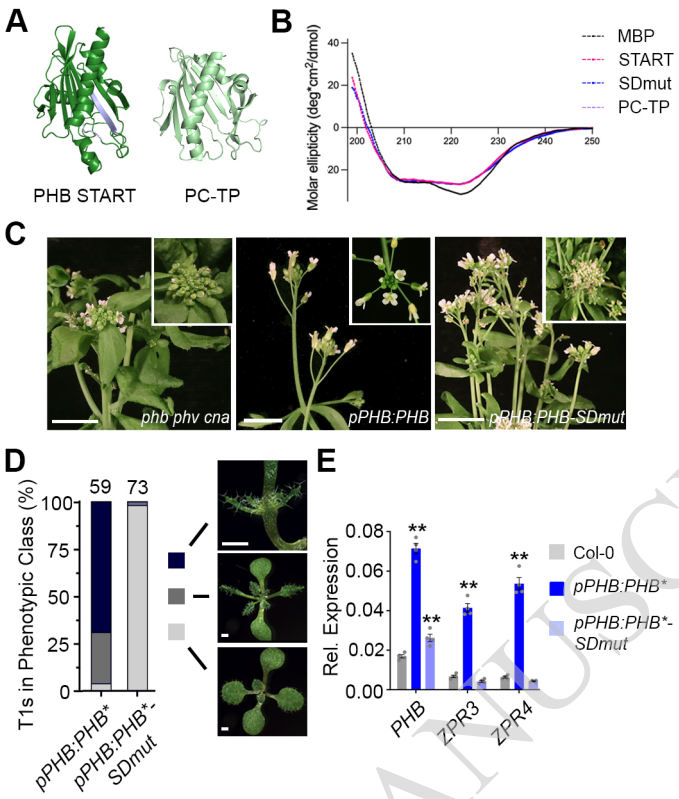

Fig 4. Mutating the START domain perturbs PHB developmental function. A, Homology-guided modeling predicts the PHB START domain (left) closely resembles PC-TP (right). Positions of amino acids mutated in PHB-SDmut are shown in lavender. **B**, Circular dichroism of MBP (negative control), MBP-START, MBP-SDmut, and MBP-PC-TP shows all three recombinant START proteins have virtually identical secondary structures. **C**, The *pPHB:PHB* transgene complements the *phb phv cna* triple mutant (left vs middle) while *pPHB:PHB-SDmut* does not (right). **D**, Phenotypic scoring of primary transformants (*n* above bar) carrying miR166-insensitive *pPHB:PHB** or *pPHB:PHB*-SDmut* constructs. Ectopic accumulation of PHB leads to severe (black) or intermediate (dark grey) gain-of-function phenotypes, whereas plants mis-accumulating PHB-SDmut appear wild-type (light grey). **E**, Ectopic *PHB** expression leads to strong upregulation of *ZPR3* and *ZPR4* targets. By contrast, *ZPR3* or *ZPR4* levels are indistinguishable from Col-0 in *pPHB:PHB*-SDmut* seedlings, despite accumulation of this variant above endogenous *PHB* levels. *n* = 3 biological replicates. ***P* ≤ 0.01, Student's *t*-test. Note: PHB, PHB-SDmut, and PHB-Delta data were collected simultaneously. PHB data from **Fig. 1** are replotted for clarity.

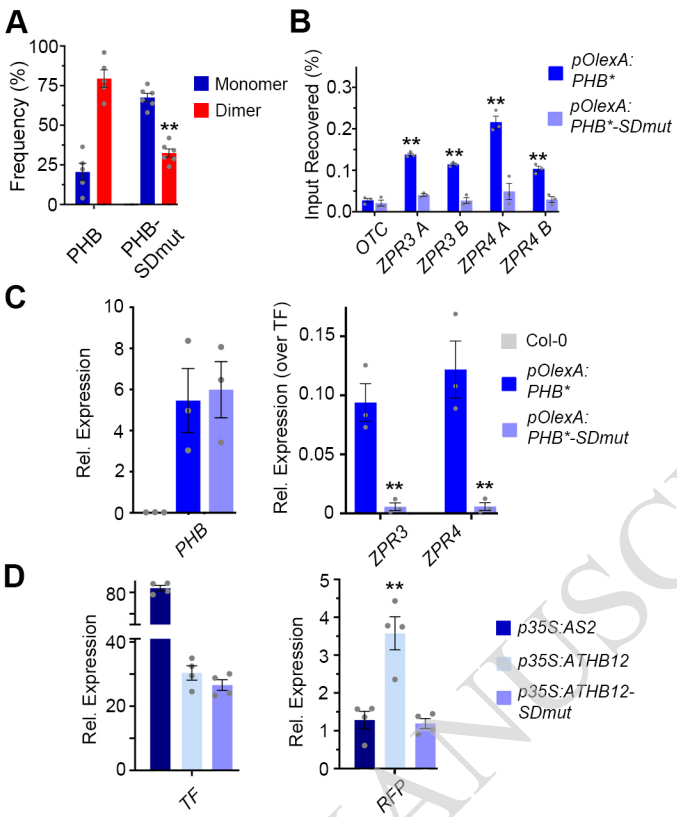

Fig. 5. Mutating PHB START reduces homodimerization and abolishes DNA-binding competence. **A**, Percentages of two-step photobleaching events from SiMPull translate to dimerization frequencies of ~80% for PHB and ~40% for PHB-SDmut. **B**, PHB-SDmut does not bind *ZPR3* or *ZPR4* regulatory regions occupied by PHB. **C**, Unlike PHB, PHB-SDmut cannot activate *ZPR3* or *ZPR4* targets in 24 hr estradiol-induction experiments. **D**, Domain-capture-mimicry experiments demonstrate fusion of SDmut START to ATHB12 (ATHB-SDmut) renders ATHB12 non-functional, as RFP reporter levels are indistinguishable from the AS2 negative control. $n = 3$ biological replicates. $^{**}P \leq 0.01$, Student's *t*-test. Note: PHB, PHB-SDmut, and PHB-Delta data were collected simultaneously. PHB data from **Figs. 2** and **3** are replotted for clarity.

A

KLRC Candidate	Compound	MBP [1]	MBP-START [1]	Enrichment (log2fold)	p-value	Matching NPASR Candidates	MBP [2]	MBP-START [2]	Enrichment (log2fold)	p-value
KLRC-1 **	**PC(34:2)**	0.006	0.015	1.22	3.52E-04	**PC(16:0/18:2)**	34.031	70.243	1.05	1.93E-04
KLRC-2	**PC(34:3)**	0.007	0.017	1.35	4.54E-05	**PC(18:2/16:1)**	0.928	2.140	1.21	2.85E-03
KLRC-3	**PC(36:3)**	0.002	0.004	1.11	2.45E-03	**PC(16:0/20:3)**	6.241	18.189	1.54	7.35E-04
						PC(18:0/18:3)	2.410	6.939	1.53	3.77E-03
						PC(18:1/18:2)	6.466	15.363	1.25	3.20E-03
KLRC-4 **	**PC(36:4)**	0.005	0.012	1.29	1.22E-04	**PC(16:0/20:4)**	5.536	23.420	2.08	8.22E-03
						PC(18:1/18:3)	0.174	0.727	2.06	8.57E-03
KLRC-5	**PC(36:5)**	0.007	0.019	1.35	4.91E-04	**PC(16:0/20:5)**	0.067	0.995	3.90	1.04E-02

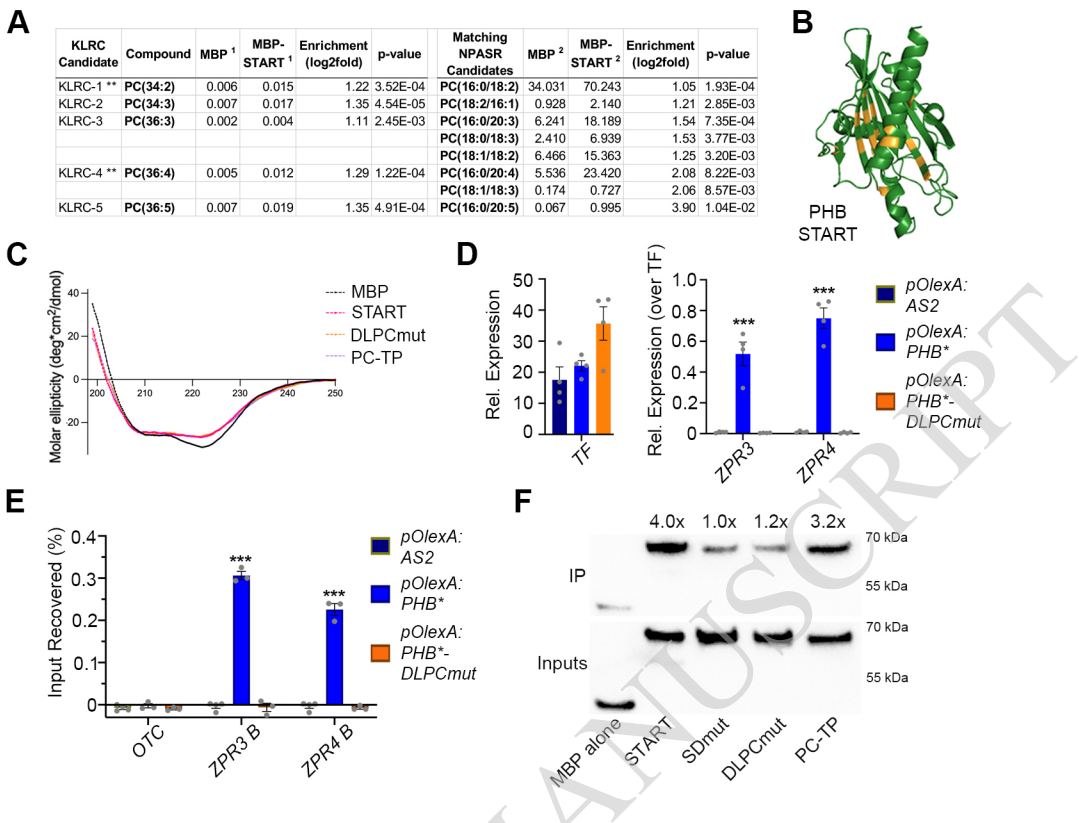

Fig. 6. PHB START domain binds PC and mutating predicted PC-contacting residues abolishes PC-binding and PHB DNA-binding competence. A, LC-MS analysis of lipids bound by recombinant MBP and MBP-START proteins after liposome incubation and repurification. Several species of PC are significantly enriched in MBP-START. Raw signal intensities are reported, ** indicates species preferentially bound by PC-TP[31]. **B**, Homology modeling of DLPCmut START variant showing position of mutations (orange). **C,** Circular dichroism of MBP, MBP-START, MBP-DLPCmut, and MBP-PC-TP showing all three recombinant START proteins have virtually identical secondary structures. **D**, Unlike PHB, PHB-DLPCmut cannot activate *ZPR3* or *ZPR4* targets in 24 h estradiol-induction experiments. **E**, PHB-DLPCmut does not bind *ZPR3* or *ZPR4* regulatory regions occupied by PHB. **F**, MBP fusions of PHB-START and PC-TP strongly bind to PC-conjugated beads, whereas binding of MBP-SDmut and MBP-DLPCmut is indistinguishable from that of the MBP negative control. IP enrichments were calculated by first normalizing each protein to its input then to the MBP alone IP/Input value. *n* = 3 or 4 biological replicates. ***$P \leq 0.001$, Student's *t*-test.

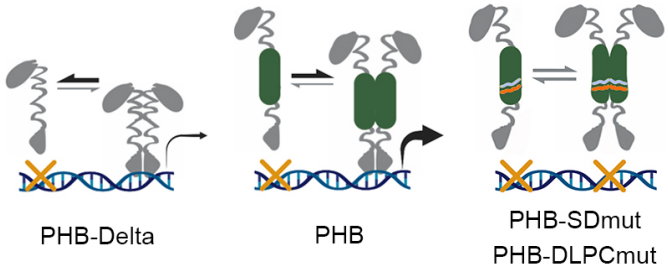

Fig. 7. The START domain potentiates HD-ZIPIII transcriptional activity. Without a START domain, HD-ZIPIII TFs make fewer and less transcriptionally potent dimers (left), while mutations that prevent ligand-binding at the START domain abolish DNA binding (right). START in green; mutations in lavender and orange. These data propose a model in which a functional START domain promotes HD-ZIPIII dimerization and transcriptional potency but requires a ligand for these TFs to bind DNA (middle).