

Ranked Choice Voting for Representative Transcripts with TRaCE

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Abstract

Summary: Genome sequencing projects annotate protein-coding gene models with multiple transcripts, aiming to represent all of the available transcript evidence. However, downstream analyses often operate on only one representative transcript per gene locus, sometimes known as the canonical transcript. To choose canonical transcripts, TRaCE (Transcript Ranking and Canonical Election) holds an 'election' in which a set of RNA-seq samples rank transcripts by annotation edit distance. These sample-specific votes are tallied along with other criteria such as protein length and InterPro domain coverage. The winner is selected as the canonical transcript, but the election proceeds through multiple rounds of voting to order all the transcripts by relevance. Based on the set of expression data provided, TRaCE can

identify the most common isoforms from a broad expression atlas or prioritize alternative transcripts expressed in specific contexts.

Availability and Implementation:

Transcript ranking code can be found on GitHub at

{{<https://github.com/warelab/TRaCE>}}

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Supplementary information: Additional data are available in the GitHub repository.

Introduction

Genome sequencing projects often use complex, automated annotation pipelines to build reference sets of gene models. These pipelines mask repeats in the assembled genome, align protein and transcript evidence, and build gene models by aggregating overlapping alignments that adhere to known or inferred splice site patterns (Hoff et al. 2019; Campbell et al. 2014; Haas et al. 2003). Before a project releases a set of high-confidence gene models, additional filtering steps may remove transcript models that lack homology or are subject to nonsense-mediated degradation (NMD).

Alternative splicing contributes to the functional diversity of a genome (Black 2003); and new sequencing technology such as PacBio IsoSeq can capture splice variants at an unprecedented scale (Wang et al. 2016; Zhang et al. 2019; Bruijnesteijn et al. 2018). However, this heightened sensitivity can lead to the detection of transcriptional noise, which can be misreported by gene builders as biologically

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3 relevant splice variants. Furthermore, it is
4 possible for partially processed transcripts
5 containing retained introns that neither
6 disrupt the reading frame nor introduce stop
7 codons to be promoted to canonical
8 transcripts (Figure 1).

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10 Comparative gene tree analysis
11 platforms such as Ensembl Compara
12 (Herrero et al. 2016) operate on a single
13 canonical transcript for each gene locus. In
14 the absence of a curated canonical
15 transcript, this is usually defined as the
16 longest transcript with the longest
17 translation, but this definition does not
18 necessarily select the best representative
19 transcript for a gene locus. Subsequently
20 developed techniques have defined
21 canonical isoforms based on expression
22 level, sequence conservation, annotation of
23 functional domains, or some combination of
24 these features (Li et al. 2014; Pruitt et al.
25 2012; Rodriguez et al. 2018; The UniProt
26 Consortium et al. 2016). For example,
27 NCBI's RefSeq Select dataset uses an
28 evidence hierarchy to identify a transcript in
29 each protein-coding human and mouse
30 gene model. The Matched Annotation from
31 NCBI and EMBL-EBI (MANE) project has
32 the goal of providing a unified set of human
33 protein-coding gene annotations, but it is
34 not known if and when such efforts will be
35 applied to other species.

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37 We developed TRaCE (Transcript
38 Ranking and Canonical Election) to choose
39 canonical transcripts based on data typically
40 available at the time of a new genome
41 annotation. In this approach, transcripts are
42 ranked by length, domain coverage, and
43 how well they represent a diverse
44 population of transcriptome RNA-seq data.
45 An 'election' based on ranked-choice voting
46 selects a canonical transcript that is the
47 first- or second-choice transcript for the
48 majority of samples. The election proceeds

through multiple rounds, effectively sorting
all transcripts by relevance. Here we
present the TRaCE algorithm and results
obtained by running TRaCE on *Zea mays*
and *Homo sapiens* gene annotations. In
addition, we describe validation of TRaCE
predictions by manual curation (Tello-Ruiz
et al. 2019) and compare TRaCE to
RefSeq/MANE Select and APPRIS
(Rodriguez et al. 2018) human transcript
classifications.

Methods

The first step in preparing to run TRaCE is
to gather a diverse set of RNA-seq
expression data covering a wide variety of
tissues or conditions to act as 'voters' in the
upcoming elections. The next step is to
align the reads, assemble sample-specific
transcripts, and quantify their expression.
Each reference gene model with multiple
transcripts (candidates) will hold an election
to sort the reference transcripts by
relevance (Figure 2).

In each election, samples rank the
candidate transcripts based on the
annotation edit distance (AED) to the most
highly expressed overlapping sample-
specific transcripts (Eilbeck et al. 2009).
AED scores range from 0 (perfect
agreement) to 1 (no overlap) and are
calculated from the pairwise similarity of
reference transcripts and aligned evidence
based on the proportion of exonic overlap.
Because there may be insufficient data to
assemble full-length transcripts from
samples in which the gene is expressed at
low levels, the AED score calculation is
restricted to overlapping portions of
candidate transcripts. A maximum AED
score cutoff (default, 0.5) prevents samples
from voting for candidate transcripts with
very little similarity. There are also cutoff

parameters for minimum expression level (default TPM, 0.5) and proportion overlapping (default, 0.5) to filter out some noise in the sample transcriptome data. The election includes additional voters that rank transcripts based on domain coverage, protein length, and transcript length. To avoid overwhelming the length-based voters when running TRaCE with many samples, sample votes are weighted to balance the electorate. Default weights were selected to prioritize functional domain coverage over protein length and total transcript length.

Once each sample voter and the length-based voters have ranked the transcripts, the election proceeds in multiple rounds selecting winners until no candidates remain. In each round, TRaCE tallies votes for top-ranked candidates; and so long as there is a tie for first place, votes for the subsequent rankings are added to the tally.

Results

We ran TRaCE on a pre-release set of *Zea mays* B73 gene models with the set of 10 RNA-seq samples that had already been aligned to the genome as part of the evidence-based gene annotation pipeline (Hufford et al. 2021). The samples were derived from shoot, root, embryo, endosperm, ear, tassel, anther, and three leaf sections (base, middle, and tip). StringTie version 1.3.5 (with the `--rf` flag) was used for transcript assembly and quantification (Pertea et al. 2016) and InterProScan version 5.38-76.0 was run to identify Pfam domains (Mulder and Apweiler 2007). The *Zea mays* B73 V5 annotation set (Zm00001eb) has 15,162 multi-transcript protein-coding gene models; for 5,616 of these (37%), the canonical transcript chosen by TRaCE was not the longest isoform. TRaCE selected canonical

transcripts for the genome annotations of 25 additional maize accessions, 33-38% of which were not the longest isoform (Suppl Table 1).

We used two approaches to validate TRaCE's predictions on maize genes. First, we modified an interactive gene tree viewer, designed to flag problematic gene models by visual inspection of the multiple sequence alignment and domain annotations (Tello-Ruiz et al. 2020). We used this interface to compare maize B73 V5 canonical transcripts (Zm00001eb) selected by TRaCE with the prior set of maize V4 canonical transcripts (Zm00001d) selected by length criteria alone. A random selection of 173 pairs of genes for which the TRaCE canonical was not the longest transcript were evaluated in the gene tree viewer and flagged if the alignment was inconsistent with outgroup orthologs. Genes were flagged if there was a relative gain or loss of conserved sequence within the transcript or at either end. Of these gene pairs, 32% were flagged as problematic in Zm00001d only, 4% in Zm00001eb only, and 5% in both versions (Suppl Table 2). The most common issue in the flagged Zm00001d gene models was gain of sequence due to an intron retention. Thus, according to this approach, TRaCE was selecting better-conserved isoforms than the prior length-based algorithm.

In the second approach, TRaCE predictions were validated by student curators who were given a subset of 48 gene models with two to five transcripts, for which TRaCE's top-ranked isoform was not the longest isoform. The students, who were not aware of TRaCE's output, were asked to rate transcripts as best, good, or poor, based on viewing the gene structure and expression evidence in the Apollo genome browser (Dunn et al. 2019). Each gene

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3 model was curated by at least 3 different
4 students. The transcript ratings were
5 mapped to a score (best 2, good 1, poor -1).
6 Transcript rankings from TRaCE and
7 rankings based on length alone were
8 compared to rankings based on curator
9 scores. For each rank (1-5), we calculated
10 the sum of the curator scores for the
11 associated transcripts. The correlation of
12 these sums between the length-based
13 ranking and the curator-based ranking was
14 0.917, whereas the TRaCE and curator
15 ranking sums had a higher correlation
16 coefficient of 0.985 (Suppl Table 3).

17 We also ran TRaCE on human
18 GRCh38 annotations (Frankish et al. 2019)
19 with a diverse panel of 127 samples of
20 human RNA-seq data covering the
21 development of seven major organs (brain,
22 cerebellum, heart, kidney, liver, ovary and
23 testis) from 4 weeks post-conception to
24 adulthood

25 ([https://www.ebi.ac.uk/gxa/experiments/E-](https://www.ebi.ac.uk/gxa/experiments/E-MTAB-6814/Results)
26 [MTAB-6814/Results](https://www.ebi.ac.uk/gxa/experiments/E-MTAB-6814/Results)). Reads were aligned
27 with hisat2 version 2.1.0 (--dta --reorder),
28 transcripts were assembled and quantified
29 with stringtie version 2.1.4 (--conservative),
30 and protein-coding reference transcripts
31 were annotated with Pfam domains using
32 InterProScan version 5.38-76.0 (Pertea et
33 al. 2016; Mulder and Apweiler 2007).

34 The GRCh38 annotation set has
35 13,848 multi-transcript protein-coding gene
36 models that were classified by both APPRIS
37 and MANE Select. The TRaCE canonical
38 was not the longest isoform in 3,717 (27%)
39 of these gene models. For comparison, the
40 principal isoform according to APPRIS and
41 the MANE Select transcript was not the
42 longest isoform in 3,061 (22%) and 4,292
43 (31%) of gene models, respectively. There
44 are 1,202 gene models where APPRIS and
45 MANE Select disagree. In these cases,
46 TRaCE agrees with APPRIS on 408 (34%)

47 genes, MANE Select on 597 (50%) genes,
48 and neither on 197 (16%) genes. On the
49 12,646 multi-transcript gene models where
50 APPRIS and MANE Select agree, TRaCE
51 gives 10,677 (84%) transcripts rank 1, 1470
52 (12%) rank 2, 351 (3%) rank 3, and 148
53 (1%) rank 4 or higher. To assess TRaCE's
54 performance on gene models with many
55 transcripts, we compared TRaCE to
56 APPRIS and MANE Select on the 90% of
57 genes with 2-10 transcripts and the
58 remaining 10% of human protein-coding
59 gene models with 11-151 transcripts. There
60 are 1,399 genes with many transcripts
61 where APPRIS and MANE Select agree. In
62 these cases, TRaCE selects 1,021 (73%) of
63 these as the canonical transcript, 215 (15%)
64 have rank 2, 92 (7%) have rank 3, and 71
65 (5%) have rank 4 or higher. On the 11,247
66 genes with fewer transcripts where APPRIS
67 and MANE Select agree TRaCE assigns
68 9,656 (86%) rank 1, 1,255 (11%) rank 2,
69 259 (2%) rank 3, and 84 (1%) rank 4 or
70 higher. For the initial release of TRaCE, we
71 manually tuned the weights on TRaCE's
72 length-based votes, but future versions may
73 benefit from an automated parameter
74 sweep to minimize these differences.

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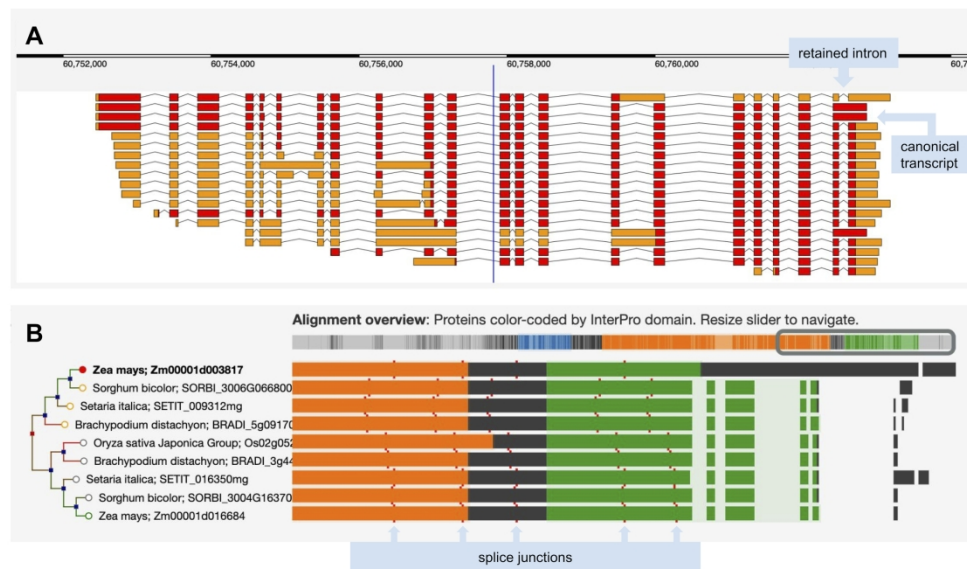
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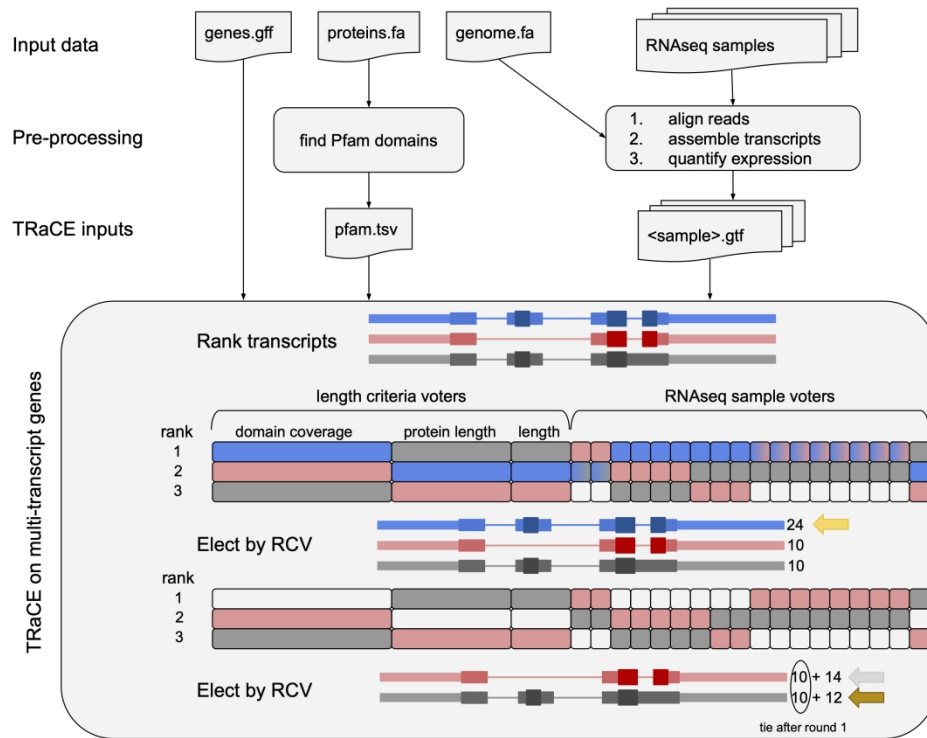
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A) The complex set of transcript models for the *Zea mays* B73 gene *sbe4* (starch branching enzyme4). Red blocks show the predicted coding regions, and orange blocks are untranslated regions. The longest translation contains a retained intron and was selected as the canonical transcript for Compara gene tree analysis. B) The left side shows a portion of the gene tree focused on this maize gene and displaying homologs from *Sorghum bicolor*, *Setaria italica*, *Brachypodium distachyon*, and *Oryza sativa Japonica*. The right side shows regions of protein sequences participating in the multiple sequence alignment, color coded by InterPro domain. The first row shows a unique region relative to other species that derives from the retained intron.

254x190mm (300 x 300 DPI)



Flowchart of preparation of TRaCE inputs and a schematic of the rank-choice voting (RCV) approach to select transcripts for an example gene with three transcripts (blue, red, gray). Exon thickness corresponds to non-coding, coding, and functional regions with Pfam domains. Voters are represented by rectangles, and rank transcripts by length criteria (9, 6, or 3 votes) or AED (1 vote per sample). Eight of the samples rank the red and blue transcripts equally (blue-red gradient), so both get tallied in round 1. RCV selects the blue transcript first with 24 rank 1 votes. After removing the blue votes from consideration, the red and gray transcripts tie with 10 rank 1 votes, but the red transcript is elected with 14 rank 2 votes.

254x190mm (300 x 300 DPI)