REVIEW

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New insights into biomarkers and risk stratification to predict hepatocellular cancer



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Abstract

Hepatocellular carcinoma (HCC) is the third major cause of cancer death worldwide, with more than a doubling of incidence over the past two decades in the United States. Yet, the survival rate remains less than 20%, often due to late diagnosis at advanced stages. Current HCC screening approaches are serum alpha-fetoprotein (AFP) testing and ultrasound (US) of cirrhotic patients. However, these remain suboptimal, particularly in the setting of underlying obesity and metabolic dysfunction-associated steatotic liver disease/steatohepatitis (MASLD/MASH), which are also rising in incidence. Therefore, there is an urgent need for novel biomarkers that can stratify risk and predict early diagnosis of HCC, which is curable. Advances in liver cancer biology, multi-omics technologies, artificial intelligence, and precision algorithms have facilitated the development of promising candidates, with several emerging from completed phase 2 and 3 clinical trials. This review highlights the performance of these novel biomarkers and algorithms from a mechanistic perspective and provides new insight into how pathological processes can be detected through blood-based biomarkers. Through human studies compiled with animal models and mechanistic insight in pathways such as the TGF-β pathway, the biological progression from chronic liver disease to cirrhosis and HCC can be delineated. This integrated approach with new biomarkers merit further validation to refine HCC screening and improve early detection and risk stratification.

Keywords Liver cancer, Cirrhosis, Biomarker, Early diagnosis, Risk stratify

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Background

Worldwide, nearly 800,000 deaths from liver cancer were reported in 2022, mostly from hepatocellular carcinoma (HCC) which accounts for 70% of liver cancers (Bray et al. 2024). Recent advances in curative treatments include liver resection, transplantation, and locoregional therapies for early HCC, and for advanced HCC, newer combinations of molecular-targeted agents (MTA) with immune checkpoint blockade (Suzuki et al. 2024). Yet, the 5-year survival remains dismal at 15–20%, which underscores the critical need for improved early detection and risk stratification for HCC.

Major risk factors for HCC include chronic viral hepatitis (HBV and HCV), alcohol use, diabetes, obesity, metabolic dysfunction-associated steatotic liver disease/



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Fig. 1 High risk populations and current clinical approaches for HCC surveillance



Fig. 2 Integrated approaches for functional biomarker studies in HCC that capture ongoing biology in the liver: TGF-β pathway

steatohepatitis (MASLD/MASH) (Konyn et al. 2021; Cho et al. 2023; Oiu et al. 2024), and hereditary disorders such as hemochromatosis (Atkins et al. 2020). These conditions can lead to progressive liver injury characterized by inflammation, necrosis, and regeneration (cirrhosis) (Alberts et al. 2022; Barton et al. 2018; Flemming et al. 2021). Current HCC screening guidelines primarily recommend AFP testing and ultrasound (US) for high-risk patients with chronic HBV infection and/or cirrhosis (Fig. 1). Studies have shown that the combination of AFP and US significantly enhances sensitivity for the detection of early-stage HCC (Tzartzeva et al. 2018). Although biannual screening using US plus AFP has shown promise in HBV patients (Zhang et al. 2004), reducing HCC mortality in this group by 37%, its limitations are compounded by the widespread use of antiviral therapies and the rising prevalence of obesity and MASLD/MASH (Esfeh et al. 2020), for whom screening guidelines are less well-defined.

Challenges in identifying robust markers that stratify risk and predict HCC include molecular heterogeneity, multiple etiologies, and diverse pathology (Fig. 2). The development of chromosomal instability with progressive accumulation of genetic and epigenetic alterations is best understood from large-scale multi-genomic human studies paired with animal models and mechanistic insight into HCC. The human Cancer Genome Atlas (TCGA) characterization of multiple cancer types as well as 363 HCC cases (Chen et al. 2018; Cancer Genome Atlas Research Network 2017; Korkut et al. 2018; Liu et al. 2018; Malta et al. 2018) has given new insight into frequent mutational analyses in multiple pathways. For example, the characterization of 363 HCC cases includes WNT signaling (44%), p53 (31%) and Telomerase (*TERT* promoter mutations in 44%), *CDKN2A* silencing in 53% as well as broader genomic alterations in the TGF- β signaling (43%). Moreover, PI3K, Myc, and Met signaling pathways, among others, play an important role and are described in greater detail in this review, together with animal models.

This review explores recent advances in non-invasive biomarkers for HCC diagnosis from the past five years, focusing on circulating biomarkers (proteins, DNA, and RNA), the gut microbiome, and imaging markers. We highlight biologically functional markers, identified through an integrated approach with animal models, that can stratify HCC risk by reflecting ongoing liver pathology, progression from fatty liver disease to cirrhosis, and ultimately, to cancer. The ideal biomarker should have a sensitivity above 85% and specificity above 95% for risk stratification of disease for cancer (Passaro et al. 2024). Here, we emphasize the incorporation of these functional biomarkers with diagnostic algorithms. By examining recent phase 2-3 clinical trials, we address their potential to stratify risk, improve early HCC detection, and improve patient outcomes.

Protein biomarkers development with phase 3 evaluation and more

Alpha-fetoprotein (AFP) is a glycoprotein implicated in multiple aspects of HCC progression, including roles in hepatocyte proliferation, invasion, metastasis, apoptosis, and immune evasion (Chen et al. 2020; H. I. Kim et al. 2022b). A meta-analysis of 41 studies revealed a suboptimal performance for AFP in detecting earlystage HCC (overall sensitivity 49%, specificity 88%) (Singal et al. 2022), while the combinational use of US with AFP improved the sensitivity (74%) but decreased the specificity (83.9%). AFP levels alone are elevated in only two-thirds of HCC patients, and false positives occur frequently in individuals with other liver conditions, limiting AFP's standalone utility (Y. T. Lee et al. 2021b). Temporal measurements with progression of disease may enhance early detection accuracy compared to single measurements (Philips et al. 2021). Additionally, HCC patients with MASLD typically had lower AFP levels compared to those with viral HCCs (Than et al. 2017). Given the modest standalone performance of AFP (sensitivity ranging from \sim 30–50% in phase 3 studies), further studies are required to validate its combined use with novel biomarkers across diverse populations to enhance early-stage detection and risk stratification.

AFP-L3, lens culinaris agglutinin-reactive fraction of AFP, a liver-specific variant of AFP, differentiates increases in AFP from HCCs as opposed to benign liver disease (Lee et al. 2021a) and is potentially useful in cases with intermediate AFP levels (20–200 ng/mL) (Sterling et al. 2007). In patients with cirrhosis, comparable diagnostic sensitivity for HCC were observed between AFP-L3 and AFP in two American and one European prospective phase 3 studies (Beudeker et al. 2023; Singal et al. 2022b; Tayob et al. 2023) (Table 1). The improved sensitivity of AFP-L3 over AFP (46.6% vs. 34.5%) observed in a Latin American cohort of patients with cirrhosis suggests its potential for enhanced HCC detection in this population, though further validation is warranted (Beudeker et al. 2023) (Table 1). A meta-analysis of six studies (n = 2447) found that AFP-L3 has high specificity (92%) but low sensitivity (34%) for early HCC diagnosis (Zhou et al. 2021). Thus, AFP-L3 may be more useful for ruling out HCC in patients with elevated AFP than for early HCC detection.

Des-y-carboxy prothrombin (DCP), also known as Prothrombin Induced by Vitamin K Absence (PIVKA-II), which is significantly elevated in serum of HCC patients (Liebman et al. 1984), has been widely used in Japan and China for HCC diagnosis and surveillance (Kim et al. 2023). Autocrine/paracrine secretion of DCP has been implicated in promoting HCC proliferation and angiogenesis through activation of the JAK/STAT3 and PLCy/ MAPK signaling pathways (Fujikawa et al. 2007; Suzuki et al. 2005). In phase 2 and 3 studies of cirrhotic patients, DCP exhibited lower sensitivity for HCC detection compared to both AFP and AFP-L3 (DCP sensitivity: from 17.6% to 36.2; AFP and AFP-L3 sensitivities: from the 30.4-50%) (Beudeker et al. 2023; El-Serag et al. 2025; Singal et al. 2022b; Tayob et al. 2023) (Table 1). Moreover, diagnostic performance of DCP varies significantly depending on the etiology of liver disease, demonstrating higher sensitivity but lower specificity in patients with viral infections (Hadi et al. 2022; Marrero et al. 2009; Piratvisuth et al. 2023). AFP-L3% with AFP and DCP utilized in GALAD assays enhance HCC detection rates (Chen et al. 2020; Singal et al. 2022b; Tayob et al. 2023) (Table 3).

Protein biomarkers with phase 2 evaluation studies

Osteopontin (**OPN**), a secreted extracellular matrix protein, that interacts with Integrins, functions as a Th1 cytokine, is involved in tissue remodeling (Lund et al. 2009) and intricately linked to the JAK2/STAT3 and PI3K/Akt signaling pathways in HCC, contributing to tumor growth, invasion, and metastasis (Desert et al. 2022; Wu et al. 2022; Yu et al. 2018). Studies also found that increased secretion of OPN contributed to promoting the synthesis of collagen-I in hepatic stellate cells via inducing HMGB1 (Arriazu et al. 2017), which is involved in chronic liver disease (Song et al. 2021). Increased plasma OPN results are similar to AFP (Abu El Makarem et al. 2011; Jang et al. 2016; Simão et al. 2015). A recent Chinese cohort study with 105 cases of chronic hepatitis,

Table 1 Biomarkers

Biomarker	Study type, No. of subjects, and Biomarker development phase	Sensi- tivity (%)	Speci- ficity (%)	Cut-off	AUROC	Ref.	Notes*
AFP	Prospective (n = 2331), phase 3	38.4	90	8.6 ng/mL	0.72	PMID: 38899967 El-Serag et al., 2024	Cirrhosis any etiology, HCC
AFP	Prospective (<i>n</i> = 534), phase 3	34.6– 38.2	90	10.8–11.2 ng/mL	0.71–0.78	PMID: 35124267 Tayob et al. 2023	Cirrhosis any etiology, HCC
AFP	Prospective ($n = 397$), transition from phase 2 to 3	50	90	17.4 ng/mL	0.77	PMID: 34618932 Singal et al. 2022b	Cirrhosis any etiology, HCC
AFP	Prospective (<i>n</i> = 1084), phase 2	42.4	94.9	20 ng/mL	0.844	PMID: 37938100 Piratvisuth et al. 2023	Chronic liver disease (> 77% viral etiology), HCC
AFP	Retro/prospective (<i>n</i> =437), phase 2	43	98	20 ng/mL	0.81	PMID: 32889146 Chalasani et al. 2021	Liver disease any etiology (> 87.4 cirrhosis) and HCC
AFP	Prospective (<i>n</i> = 163), phase 2	75	93.5	14.2 ng/mL	0.869	PMID: 36013482 Hadi et al. 2022	Non-cirrhosis and cirrho- sis with etiology (HBV/ HCV/MASH), HCC
AFP	Prospective (<i>n</i> = 1120), phase 2	44.8	76.1	20 ng/mL	0.692	PMID: 31358576 Cai et al. 2019	HBV, cirrhosis and HCC
AFP	Prospective (<i>n</i> = 288), phase 3	29.2	87.4	20 ng/mL	0.59	PMID: 37708457 Beudeker et al. 2023	European cirrhosis and HCC
AFP	Prospective (<i>n</i> = 284), phase 3	34.5	92.4	20 ng/mL	0.66	PMID: 37708457 Beudeker et al. 2023	Latin America cirrhosis and HCC
AFP-L3	Prospective (n = 2331), phase 3	37.6	90	7.5%	0.61	PMID: 38899967 El-Serag et al., 2024	Cirrhosis any etiology, HCC
AFP-L3	Prospective (<i>n</i> = 534), phase 3	34.6– 41.2	90	8.3-8.4%	0.64–0.81	PMID: 35124267 Tayob et al. 2023	Cirrhosis any etiology, HCC
AFP-L3	Prospective ($n = 397$), transition from phase 2 to 3	46.2	90	11.9%	0.80	PMID: 34618932 Singal et al. 2022b	Cirrhosis any etiology, HCC
AFP-L3	Prospective (<i>n</i> = 288), phase 3	30.4	81.1	10%	0.56	PMID: 37708457 Beudeker et al. 2023	European cirrhosis and HCC
AFP-L3	Prospective (<i>n</i> = 284), phase 3	46.6	91.7	10%	0.69	PMID: 37708457 Beudeker et al. 2023	Latin America cirrhosis and HCC
AFP-L3	Retro/prospective (n=437), phase 2	56	93	10%	0.81	PMID: 32889146 Chalasani et al. 2021	Liver disease any etiology (> 87.4 cirrhosis) and HCC
DCP	Prospective (n=2331), phase 3	30.4	90	2.91 ng/mL	0.75	PMID: 38899967 Cirrhosis any etiol El-Serag et al., 2024 HCC	
DCP	Prospective (<i>n</i> = 534), phase 3	17.6– 23.1	90	1.4–1.5 ng/mL	0.68–0.72	PMID: 35124267 Tayob et al. 2023	Cirrhosis any etiology, HCC
DCP	Prospective (<i>n</i> = 288), phase 3	26.7	87.4	7.5 ng/mL	0.57	PMID: 37708457 Beudeker et al. 2023	European cirrhosis and HCC
DCP	Prospective (<i>n</i> = 284), phase 3	36.2	97.4	7.5 ng/mL	0.65	PMID: 37708457 Beudeker et al. 2023	Latin America cirrhosis and HCC
DCP	Prospective ($n = 397$), transition from phase 2 to 3	34.6	90	5.9 ng/mL	0.70	PMID: 34618932 Singal et al. 2022b	Cirrhosis any etiology, HCC
DCP	Prospective (<i>n</i> = 1084), phase 2	61.3	88.7	20 ng/mL	0.772	PMID: 37938100 Piratvisuth et al. 2023	Chronic liver disease (> 77% viral etiology), HCC
DCP	Retro/prospective (n=437), phase 2	39	93	7.5 ng/mL	0.83	PMID: 32889146 Chalasani et al. 2021	Liver disease any etiology (> 87.4 cirrhosis) and HCC
DCP	Prospective (<i>n</i> = 163), phase 2	90	82.1	36.7 mAU/mL	0.905	PMID: 36013482 Hadi et al. 2022	Non-cirrhosis and cirrho- sis with etiology (HBV/ HCV/MASH), HCC
DCP	Retrospective (<i>n</i> = 186), phase 2	72	71	7 ng/mL	0.715	PMID: 38994169 He et al. 2024	Healthy control and AFP- negative HCC
OPN	Retrospective (<i>n</i> = 322), phase 2	79.2	79.6	-	0.85	PMID: 32043608 Zhu et al. 2020	Chronic hepatitis, cir- rhosis, and HCC
MDK	Meta-analysis of 9 studies	87	86	0.5 ng/mL	0.95	PMID: 31600291 Zhang et al. 2019	НСС

Table 1 (continued)

Biomarker	Study type, No. of subjects, and Biomarker development phase	Sensi- tivity (%)	Speci- ficity (%)	Cut-off	AUROC	Ref.	Notes*
MDK	Meta-analysis of 17 studies	84	82	-	0.89	PMID: 32039435 Lu et al. 2020	HCC
GP73	Retrospective (<i>n</i> = 186), phase 2	75.6	93	103 ng/mL	0.843	PMID: 38994169 He et al. 2024	Healthy control and AFP- negative HCC
GPC3	Retrospective (<i>n</i> = 344), phase 2	80	85	0.0414 ng/mL	0.88	PMID:32087138 Liu et al. 2020	Healthy control and HCC
TGF-β1	Prospective	53.1%	98.9%	50 ug/ g-1 creatinine	0.730	PMID: 9166938 Tsai et al. 1997	Cirrhosis (74.4% viral etiology), HCC
SCCA-IgM	Prospective	89%	50%	130 AU/mL	0.63	PMID:24635038 Pozzan et al. 2014	Cirrhosis, healthy control, and HCC

*Notes refer to the main groups included in the studies, specifically detailing the composition of the control groups

116 of liver cirrhosis, and 101 of HCC showed that serum OPN analyses gave a better AUROC of 0.851 (79.2% sensitivity and 79.6% specificity) compared with AFP (AUROC of 0.683) or DKK1 (AUROC of 0.639) (Zhu et al. 2020) (Table 1). In AFP-negative samples, serum OPN also performed well with an AUROC of 0.838.

Midkine (MDK), a heparin-binding growth factor, activates multiple key pathways such as MAPK, WNT, and TGF-B, leading to increased cancer proliferation, angiogenesis, and metastasis (Du et al. 2022; Sun et al. 2017). A meta-analysis of 9 studies showed that MDK displayed diagnostic efficacy for HCC with a cutoff value of 0.5 ng/mL: an AUROC of 0.95, sensitivity 87%, and specificity 86%) (Zhang et al. 2019) (Table 1). Another systematic meta-analysis of 17 studies further confirmed that MDK showed better performance in diagnosing early-stage HCC than AFP: AUROC, 0.89 vs. 0.52, sensitivity, 84% vs. 44%, specificity, 82% vs. 85% (Lu et al. 2020) (Table 1). Also, MDK showed promising performance in AFP negative HCC: an AUROC of 0.91, sensitivity 89%, and specificity 84%. A recent study validated the functional role of circulating MDK in promoting liver carcinogenesis via activating PI3K/AKT/mTOR signaling (Du et al. 2022), indicating that MDK is a promising biomarker that deserves further validation.

Dickkopf-1 (DKK1) promotes liver cancer invasion and metastasis via β -catenin/MMP7 signaling (Chen et al. 2013). DKK1 genetic deletion impairs HCC cell invasion, proliferation, and tumor development (Seo et al. 2021). Furthermore, analysis of tissue microarray data suggests that DKK1 may serve as a new prognostic predictor for HCC patients, particularly for those with normal AFP levels and those in the early stages of the disease (Yu et al. 2009). In a large-scale multicenter study (*n* = 1284), serum DKK1 levels were significantly elevated in patients with HCC compared to those with cirrhosis or chronic HBV infection(Shen et al. 2012), which displayed complementary diagnostic potential with AFP. For early-stage HCC, DKK1 demonstrated superior diagnostic accuracy (AUROC 0.85 vs. 0.658 for AFP), with higher sensitivity (70.9% vs. 54.4%) and specificity (84.7% vs. 69.3%). Combining DKK1 and AFP further improved performance, achieving 84.9% sensitivity and 77.4% specificity. Similarly, two independent cohorts (n = 90 and n = 80) demonstrated that combination of serum DKK1 and AFP may enhance HCV related HCC diagnostic accuracy (Eldeeb et al. 2020; Fouad et al. 2016). Furthermore, studies have revealed DKK1 promoter hypermethylation in liver tissue from HCV-infected patients with chronic liver disease and cirrhosis preceding HCC development (Umer et al. 2014). Taken together, these studies suggest that DKK1 as a potent inhibitor of WNT pathway may serve as a valuable biomarker for early detection of virus-induced HCC.

Golgi protein-73 (GP73) is a type II Golgi transmembrane protein found significantly elevated in hepatocytes affected by chronic liver diseases and HCC (Gatselis et al. 2020), which acts as a driver oncogene, initiating intraand intercellular signaling cascades such as JAK2/STAT3 and ER stress that enhance the angiogenesis and aggressiveness and reshape the tumor microenvironment of HCC (Chen et al. 2015; Wei et al. 2019; Ye et al. 2024). Cleavage releases GP73 and renders it a potentially useful serum biomarker (Gatselis et al. 2020). Given its unique expression in liver tissue from HCC patients, targeting GP73 could provide a strategy to inhibit angiogenesis with reduced off-target effects, as well as a tool for HCC detection. GP73 shows high specificity for HCC and may offer additional diagnostic value, particularly for AFPnegative patients (75.6% sensitivity, 93% specificity) (He et al. 2024; Zhang et al. 2023). Unfortunately, a 36-study meta-analysis revealed moderate diagnostic accuracy for GP73 in cirrhotic patients possibly because of increased GP73 levels in both cirrhotic and HCC patients (Zhang et al. 2023).

Glypican 3 (GPC-3), a member of the heparan sulfate proteoglycan family, is another oncofetal protein found elevated in hepatocellular carcinoma cells and serum small extracellular vesicles (Sun et al. 2023). GPC3

signals through WNT members and extracellular signalregulated kinase (ERK) pathways (Castillo et al. 2016). Its value may lie in AFP-negative HCC patientswhereGPC-3 displays a sensitivity of 54.6% and a specificity of 76% among AFP-negative patients (AFP<400ug/L) was observed (Li et al. 2013) (Table 1). Combining AFP and GPC3 improved the sensitivity to 88.1%, but the specificity decreased to 82.7% (Liu et al. 2020), potentially as a combination providing the most useful predictors tested so far.

Angiopoietin-2 (ANG2), associated with tumor angiogenesis (Tanaka et al. 1999, 2002), has been shown to outperform AFP in predicting overall survival (OS) in HCC (Llovet et al. 2012). ANG2-blocking antibodies inhibit tumor angiogenesis and metastasis in mice, suggesting its potential role in future therapeutic targeting (Saharinen et al. 2017). Ang-2 levels are associated with advanced HCC, cases with acute renal injury and higher mortality in decompensated cirrhosis, and liver function indicators such as high MELD and Child-Pugh scores, as well as associated with tumor aggressiveness (Ao et al. 2021; Choi et al. 2021).

Viral antigens HBV antigens, such as HBcrAg, represent promising markers due to their direct involvement in liver pathology and carcinogenesis. HBcrAg levels, which are unaffected by nucleotide analog treatment, provide a reliable indicator of viral replication and intrahepatic activity. Higher HBcrAg levels correlate with increased HCC risk, identifying patients with an inactive virus but elevated HCC risk (Chang et al. 2022). In a study with 2666 patients with chronic HBV infection, HBcrAg levels higher than 10KU/ml positively correlated with increased HCC incidence (Tseng et al. 2019).

Squamous Cell Carcinoma Antigen (SCCA) and SCCA-IgM complexes have also emerged as potential markers, with SCCA-IgM showing greater sensitivity and specificity in prognosticating HCC response to therapy. Studies suggest lower SCCA-IgM levels in patients responsive to locoregional therapies, supportive of its diagnostic relevance (Guarino et al. 2017; Pozzan et al. 2014).

DNA/RNA biomarkers

In a recent case-control cohort study with diverse etiologies (n = 558) (Campani et al. 2024), whole-exome sequencing analysis demonstrated that plasma circulating tumor DNA (ctDNA) mutation rates in patients with active HCC were significantly higher (40.2%), compared to that of chronic liver disease control group (1.8%). Consistent with the genomic analysis of liver tissues from TCGA HCC cohort (Cancer Genome Atlas Research Network 2017), the top 5 highest mutations occur in TERT promoter (27.5%), TP53 (21.3%), CTNNB1 (13.1%), PIK3CA (0.2%), and NFE2L2 (0.2%), suggesting these ctDNA mutations may serve as promising non-invasive markers for HCC diagnosis. Another study (n = 609) reported that urine ctDNA biomarkers (TP53, RASSF1a, and GSTP1) combined with serum AFP significantly increased the sensitivity for early-stage HCC detection from 62 to 92% (BCLC stage 0, Kim et al. 2022a).

Global 5-hydroxymethylcytosines (5hmC) contents were significantly decreased in liver tissues from patients with early-stage HCC (Liu et al. 2019), which was associated with HBV infection and decreased translocation enzyme activity. As potential effective epigenomic biomarkers, a 32-gene panel that captures 5hmc signature in cell free DNA (cfDNA) significantly discerned early-stage HCC from non-HCC (AUROC of 0.884) and from a highrisk group with chronic hepatitis B virus infection or liver cirrhosis (AUROC of 0.846) in a cohort of 1204 HCC patients and 1350 controls (chronic liver disease and healthy individuals) (Cai et al. 2019) (Table 3). Another independent study (n = 262) (Cai et al. 2021) expanding the panel to 64-gene 5hmC signatures in cfDNA further increased the performance for HCC diagnosis (AUROC of 0.93). These studies supported that 5hmC markers could serve as a noninvasive tool for early-stage HCC detection among high-risk subjects.

The better performance of plasma methylated DNA markers (MDMs) for HCC diagnosis (Kisiel et al. 2019) has been validated in a phase 2 study (n = 244, AUROC 0.96, sensitivity 95%, specificity 92%), which captures 6-marker changes (HOXA1, EMX1, AK055957, ECE1, PFKP, and CLEC11A normalized by B3GALT6). Recently, in a clinical trial study (NCT03628651), a multiple-target blood-based panel (Chalasani et al. 2021) that combines 4 methylated DNA markers (HOXA1, EMX1, TSPYL5, and B3GALT6) and 2 protein markers (AFP and AFP-L3) outperformed the GALAD score for early-stage diagnosis (AUROC: 0.88 vs. 0.81; sensitivity: 74% vs. 60%, specificity: 90% vs. 86%) (Table 3). The performance of this panel was comparable in patients with virus or non-virus etiologies, and with or without cirrhosis. Moreover, validation of the multi-target panel (HOXA1, TSPYL5, plus AFP and sex) (Chalasani et al. 2022) Using an independent cohort of 156 HCC cases and 245 controls, the multiple target panel that combines methylated DNA markers and protein markers displayed similar performance (AUROC 0.86, sensitivity 72%, specificity 88%) for early-stage HCC diagnosis. These data implied that the multiple target panel may significantly improve early-stage HCC diagnosis.

Several studies highlight circulating microRNAs (miRNA) and exosomal miRNAs could serve as non-invasive biomarkers for HCC surveillance. A serum miR-NAs panel that includes six targets (miR-21, miR-221,

miR-801, miR-1246, miR-26a, and miR-122) displayed clinical value for the early diagnosis of HCC (AUROC of 0.95) (Zhang et al. 2025). More recently, exosomal miRNAs (miR-10b-5p, miR-221-3p, miR-223-3p, miR-21-5p) may effectively distinguish HCC patients from CH/LC control with AUROC of 0.86, sensitivity of 74%, and specificity of 86% (Ghosh et al. 2020). Another independent study reported a similar performance of a panel including five circulating exosomal miRNAs (miR19-3p, miR16-5p, miR30d-5p, miR-451a, miR-223-3p) with AUROC of 0.85, for distinguishing HCC with non-virus etiology and non-HCC control (Boonkaew et al. 2023).

Molecular pathways with new insight from animal models

In an ideal situation, a single simple model that replicates the spectrum of HCC from cirrhosis should provide rapid new insight into biologically relevant markers that could stratify risk for HCC. These new insights can be provided from animal models. While no single animal model replicates HCC progression, commonly used preclinical models for HCC include cell lines, organoids, patient-derived xenografts, scaffold-based models, those induced by chemotoxic agents, special diets, genetic modifications, and tumor cell transplantation (He et al. 2015; Zabransky et al. 2023). c-MYC which is overexpressed in up to 70% of viral and alcohol-related human HCCs (Schlaeger et al. 2008) lends itself as a strong GEM model, in which dual (albumin-driven) AEG-1 and Myc overexpression- mice develop aggressive HCCs and lung metastases (Srivastava et al. 2015). GEM models expressing an activated form of β -catenin, the downstream effector of the Wnt pathway, or harboring a liver-specific Apc knockout (KO) showed hepatomegaly or HCC after a long latency (Colnot et al. 2004). In liver-specific p53 KO model through Cre–Lox recombination, the AlfpCre⁺Trp53^{$\Delta 2-10/\Delta 2-10$} mice develop liver cancer in 14 months (Katz et al. 2012). A liver-specific Setd2 depletion model, finding that Setd2 deficiency is sufficient to trigger spontaneous HCC formation (Li et al. 2021). c-MET levels are raised in 20–48% of human HCC samples and represents a potentially therapeutic target (Adebayo Michael et al. 2019; Tao et al. 2017; Venepalli and Goff 2013; You et al. 2011). With Pten expression being reduced in up to 50% and activated mutant forms of PIK3CA in 4% of hepatocellular tumors, liver-specific knockout of Pten in mice develop steatosis and late-onset liver cancer (Horie et al. 2004). Those models provide new insight into biologically relevant biomarkers for HCC.

Proteins reflecting liver pathophysiology: findings on TGF- β pathway modulation

Animal models combined with analyses of human genomics could ideally provide the most relevant biologically functional biomarkers (Dhanasekaran et al. 2025). For instance, a genomic, epigenomic, and transcriptomic landscape of 44 TGF- β pathway genes and 50 downstream target genes of the pathway in 9,125 patients across all 33 TCGA PanCancer Atlas tumor types (Korkut et al. 2018) revealed that 40% of the cancers carry TGF- β -Smad pathway gene alterations with a common transcription signature; the genomic alterations affect expression of metastatic and epidermal-mesenchymaltransition (EMT) genes; the pathway is most frequently aberrant in Liver and GI cancers, which exhibited 113 of the 176 hotspot mutations identified in the overall cohort.

In cancer, the TGF- β pathway plays apparently contradictory roles, either suppressing (early) or (later) promoting tumor growth. Mouse models of hepatocellular cancers indicate a primarily an early tumor-suppressive role (Chen et al. 2016; David et al. 2016; Katz et al. 2016). Examples include mouse models of HCC with haploinsufficiency of Tgfbr2, Tgfb, and intercrosses between Smad3/4 with the adaptor Sptbn1 (Tgfbr2-'-, Smad4^{+/-}Sptbn1^{+/-} and Smad3^{+/-}Sptbn1^{+/-} on a C57BL/6 background) (Biswas et al. 2004; Gough et al. 2021a; Gu et al. 2020; Z. Wang et al. 2021b). More recently, we have uncovered obesity-driven HCC in our mouse models with disruption of TGF-β signaling and loss of aldehyde dehydrogenase 2 (Aldh2) (Rao et al., 2021; Yang et al. 2024). ALDH2 detoxifies cells of lipid end productsreactive aldehydes such as 4-HNE that accumulate with a high-fat diet, and the Aldh2^{-/-}Sptbn1^{+/-} mice provided new insight into the role of obesity in promoting HCC (Yang et al. 2024).

Taking this further, elevated TGF-B1 levels in human HCC tissue are associated with poor prognosis and immune suppression, marking it as a potential target for immunotherapy (Gough et al. 2021; Jin et al. 2022). Both plasma and urine TGF-B1 levels are higher in patients with HCC than in those with cirrhosis, display comparable diagnostic ability as AFP to discriminate HCC from cirrhosis (Song et al. 2002; Tsai et al. 1997). TGFBR2 is a transmembrane protein that plays a crucial role in regulating TGF-β signaling, which is closely associated with the progression of liver cirrhosis and HCC. Reduced TGFBR2 levels have been observed in liver tissue from HCV-HCC compared with HCV-related cirrhosis patients and healthy subjects, which were significantly correlated with aggressive features of HCC (Abu El-Makarem et al. 2022). A multi-cohort study demonstrated a significant reduction in serum TGFBR2 levels in HCCs compared to cirrhotic liver tissues (Zaidi et al. 2022). Also, circRNA-TGFBR2 has been observed to promote HCC progression via regulating autophagy (Wang et al. 2023), implying a role in risk stratification of HCC. Thus, additional studies are necessary to investigate the potential biomarker value of circulating TGFBR2 in HCC.

Myostatin (MSTN, or GDF8) is a member of the transforming growth factor beta (TGF- β) superfamily and may prove to be a promising biologically relevant marker, in part from its role as an autocrine inhibitor of muscle growth, contributing to muscle wasting in patients with sarcopenia, which are major issues in cirrhosis and HCC. Sarcopenia is prevalent in up to 40% of cirrhotic patients, particularly with alcoholic liver disease or Child-Pugh class C, linked to a high risk of mortality (Cui et al. 2023; Tantai et al. 2022). Recent studies have identified a causal relationship between sarcopenia and increasing risk of HCC in European populations (Cao et al. 2024), implying that MSTN, the primary mediator of sarcopenia, is a promising biomarker candidate for HCC risk stratification. Consistent with this, a multicenter prospective study found a two-fold increase in serum myostatin levels significantly predicted a higher risk of HCC development in patients with alcoholic cirrhosis (Kim et al. 2020). However, lower MSTN levels were observed in patients with acute decompensation and acute-on-chronic liver failure (ACLF) (Ruiz-Margáin et al. 2023). Therefore, multiple timepoint assays or longitudinal studies are necessary for predicting HCC risk and stratifying the value of MSTN.

IL-6 is a proinflammatory cytokine that is significantly elevated in both liver tissue and serum of patients with HCC (Kao et al. 2015), specifically in progressive sarcopenia and advanced HCC stage (Choi et al. 2020; Myojin et al., 2022). A systematic meta-analysis of 18 studies demonstrated higher IL-6 levels in HCC patients compared with hepatitis and cirrhosis patients and healthy controls (Shakiba et al. 2018). Serum high mobility group box 1 protein (HMGB1) is a proinflammatory molecule that induces inflammatory cytokine production of TNF- α and IL-6 (Chen et al. 2022; Tripathi et al. 2019). Elevated HMGB1 levels in HCC liver tissue (Liu et al. 2012), are associated with poor prognosis HCC. High mobility group box 2 (HMGB2), closely related to HMGB1 is overexpressed in HCC cells (Kwon et al. 2010; Lu et al. 2023), via activating signaling pathways such as ERK, PI3K/ AKT, and Wnt/ β -catenin. HMGB2 is involved in stellate cell activation, and serum HMGB2 levels are increased in patients with liver fibrosis and cirrhosis (Huang et al. 2023). Elevated HMGB2 is associated with poor prognosis of HCC patients. Collagen type I α 1 (COL1A1) is often overexpressed in cancers, influencing cell proliferation, metastasis, apoptosis, and cisplatin resistance, with high levels linked to poor patient prognosis (Li et al. 2022). COL1A1 is implicated in epithelial-to-mesenchymal transition (EMT) and stemness in HCC (Ding et al. 2024; Ma et al. 2019). COL1A1 levels are higher in HBV-positive cirrhosis and HCC (Mohamed et al. 2021), reflecting a potential for risk stratification of HCC risk in Hepatitis B virus infected patients.

Gut Microbiome

The liver-gut axis plays a crucial role in liver disease progression and carcinogenesis. Recent studies indicate that specific gut bacteria such as Bacteroides, Streptococcus, and Veillonella are enriched in patients with HCC, especially in non-viral HCC, and may serve as potential biomarkers (Jinato et al. 2024) (Table 2). The presence of viable bacteria within liver tissue further indicates the contributive roles of the gut microbiome in HCC pathophysiology, potentially opening new avenues for noninvasive diagnosis and therapeutic intervention (Huang et al. 2022). Circulating microbial signatures are another emerging subject in the cancer field and are thought to be partially derived directly from the gut via bacterial translocation (You et al. 2022). Similarly, oral Cyanobacteria may be independently associated with HCC risk, possibly via direct impact on the tumor-promoting effects of microcystins and other hepatotoxins and their disruptive influence on lipid metabolism. A 2021 study identified oral Cyanobacteria as an independent risk factor for HCC through bacterial 16 S rRNA sequences in oral samples from 90 HCC cases and 90 controls -part of a larger U.S. case-control study of HCC among patients diagnosed from 2011 to 2016 (Hernandez et al. 2022; Song et al. 2023) (Table 2). Elevated levels of gut bacteria such as Dialister, Veillonella, and Eubacterium, along with their associated metabolites, have been linked to early HCC recurrence (Zheng et al. 2023). Furthermore, increased abundance of Veillonella has shown potential for differentiating HCC from cirrhosis (Lapidot et al. 2020). We have observed altered microbiomes in our mouse models with disruption of TGF- β signaling that develop spontaneous HCC and other gastrointestinal cancers (Gu et al. 2020; Z. Wang et al. 2021b). Interestingly, our group and others have observed that these mutant mice do not develop cancers in a germ-free environment (Gu et al. 2020; Maggio-Price et al. 2006). As a novel potential diagnostic tool for HCC, even though the performance of gut microbiome is currently limited in scale and lacking extensive sample validation, the predictive model using gut microbiome together with AFP demonstrated better accuracy (AUROC: 0.9811 vs. 0.8505) (Yang et al. 2023), suggesting its potential complementary effect to the serum testable markers. Future studies should provide new insights into the role of the microbiome in the setting of altered mutational profiles in HCC.

Diagnostic algorithms GALAD score

The GALAD score was developed in 2015 and incorporates Gender, Age, and three biomarkers: AFP, AFP-L3%, and DCP to improve the detection of HCC, specifically in patients with chronic liver disease. In phase 2 studies, the GALAD score shows promising results (Table 3). In

Table 2 Microbiome

Biomarkers	Samples	Sample Size	Findings	Diagnosis/Prognosis Potential	Reference
Gut Microbiome	Fecal	16 healthy controls, 33 patients with viral-HCC (17 and 16 cases with hepatitis B virus (HBV) and hepatitis C virus (HCV) infection, respective- ly), and 18 patients with NBNC-HCC	Bacteroides, Streptococcus, Ruminococcus gnavus group, Veillonella, and Erysipelatoclos- tridium ↑ Romboutsia, UCG-002, Lachno- spiraceae NK4A-136, Eubacteri- um hallii group, Lachnospiraceae ND-3007 group, Erysipelotricha- ceae UCG-003, and Bilophila↓	Identify a gut microbiota signa- ture in differentiating between viral-related HCC (Viral-HCC) and non-hepatitis B-, non-hepatitis C-related HCC (NBNC-HCC)	PMID: 38183473 Jinato et al. 2024
Gut Microbiome	Fecal	124 patients diagnosed with HBV- associated HCC and 82 HBV-related hepatitis, and 86 healthy volunteers	Dialister, Veillonella, Eubacterium coprostanoligenes group, and Lactobacillus genus ↑ Bacteroides↓	TNM, AST, Veillonella, and Strep- tococcus pneumoniae used in a model for the prognosis/early recurrence of HBV induced HCC, AUROC: 0.78	PMID: 37778742 Zheng et al. 2023
Gut Microbiome	Fecal	30 HCC-cirrhosis patients, 38 cir- rhotic patients without HCC, and 27 age- and body mass index [BMI]- matched healthy volunteers	Veillonella and Scardovia ↑ Lachnospira, Ruminococcus, and Butyricicoccus ↓	Demonstrates the potential of fecal microbes as tools for non- invasive diagnosis or microbi- ome-oriented interventions in HCC-cirrhosis	PMID: 32546668 Lapidot et al. 2020
Oral Microbiome	oral	90 HCC cases and 90 controls with oral samples obtained from a larger population-based case-control study of 673 patients with HCC and 1 166 controls	Cyanobacteria positively associated with HCC	Novel evidence that oral Cyano- bacteria may be an independent risk factor for HCC	PMID: 34697061 Hernan- dez et al. 2022

↑: Upregulated in HCC/Early Recurrence ↓: Downregulated in HCC/No Early Recurrence/ Healthy Controls

Sequencing method used for the above studies: 16 S rRNA sequencing

a cohort of patients with chronic liver disease (n = 1084), primarily viral in etiology, the score achieved a high sensitivity of 71.8%, specificity of 90%, and an AUROC of 0.907, underscoring its effectiveness in this group (Piratvisuth et al. 2023). A second phase 2 retro/prospective study with 437 patients demonstrated an AUROC of 0.87, further supporting the diagnostic utility of the GALAD score in a population with a high prevalence of cirrhosis (Chalasani et al. 2021). In a large, phase 3 prospective cohort (n = 2331) with cirrhosis of any etiology, the GALAD score exhibited a sensitivity of 40% and specificity of 90%, with an AUROC of 0.76 (El-Serag et al., 2024). A prospective transition study between phase 2 and phase 3 with a cohort of 397 cirrhotic patients showed the advantage of longitudinal assessments of GALAD over single-timepoint scores for HCC diagnosis, with AUROCs of 0.83 and 0.78, respectively, further underscoring the added benefit of longitudinal monitoring (Singal et al. 2022b). While AFP-L3 has been found contributed negligibly in GALAD (Hou et al. 2025; Johnson et al. 2014), two phase 2 prospective studies (n = 1006 and n = 1142) found that the GAAD score (combining sex, age, AFP, DCP) performed as sensitive (90% and 93.7%) and specific (85.3% and 83.1%) as the GALAD algorithm (sensitivity 93%, specificity 83.3%) in differentiating HCC from chronic liver disease (Hou et al. 2025; Piratvisuth et al. 2023).

Doylestown algorithm

The Doylestown algorithm incorporates AFP and other laboratory markers (age, gender, alkaline phosphatase and alanine aminotransferase levels) and has demonstrated enhanced specificity and sensitivity over AFP alone (Mehta et al. 2018; Wang et al. 2016) (Table 3), especially at early detection time points. Further development may enhance its utility, particularly in racial and ethnic minority populations where HCC disparities persist. In a prospective study involving 120 patients with cirrhosis of various etiologies, the Doylestown algorithm achieved a sensitivity of 50% and a specificity of 90%, with an AUROC of 0.98 (Mehta et al. 2018) (Table 3). This high AUROC suggests that the Doylestown algorithm may outperform traditional single biomarker approaches, particularly in early HCC detection, where sensitivity and specificity are critical.

Hepatocellular carcinoma early detection screening

Hepatocellular Carcinoma Early Detection Screening (HES) is a screening technique that combines AFP with age alanine aminotransferase and platelets. HES has been evaluated across multiple cohorts to assess its diagnostic effectiveness in detecting HCC among patients with cirrhosis (Table 3). In a phase 2–3 transition study involving 397 patients with cirrhosis of varying etiologies, the HES score demonstrated a sensitivity of 34.6% with a specificity of 90%, yielding an AUROC of 0.71, highlighting its moderate diagnostic accuracy in this group (Singal et

Table 3 Biomarker panel and algorithms

Biomarker panel	Study type, No. of subjects, and Biomarker develop- ment phase	Sensitiv- ity (%)	Speci- ficity (%)	Cut-off	AUROC	Ref.	Notes*
GALAD	Prospective (n = 534), phase 3	30.0-32.4	90	(-0.03)-0	0.75–0.79	PMID: 35124267 Tayob et al. 2023	Cirrhosis any etiology, HCC
GAAD (combing age, sex, AFP, DCP)	Prospective (n = 1084), phase 2	71.8	90	2.57	0.907	PMID: 37938100 Piratvisuth et al. 2023	Chronic liver disease (> 77% viral etiology), HCC
GALAD, Single-timepoint	Prospective (<i>n</i> = 397), transi- tion from phase 2 to 3	53.8	90	-0.33	0.78	PMID: 34618932 Singal et al. 2022b	Cirrhosis any etiology, HCC
GALAD, longitudinal	Prospective ($n = 397$), transi- tion from phase 2 to 3	69.2	90	-0.33	0.83	PMID: 34618932 Singal et al. 2022b	Cirrhosis any etiology, HCC
GALAD	Prospective (n = 288), phase 3	65.8	71.7	-0.63	0.69	PMID: 37708457 Beudeker et al. 2023	European cirrhosis and HCC
GALAD	Prospective (n = 284), phase 3	69.8	82.9	-0.63	0.76	PMID: 37708457 Beudeker et al. 2023	Latin America cirrhosis and HCC
GALAD	Prospective (n=2331), phase 3	40	90	-0.38	0.76	PMID: 38899967 El-Serag et al., 2024	Cirrhosis any etiology, HCC
GALAD	Retro/prospective (<i>n</i> =437), phase 2	72	86	-0.63	0.87	PMID: 32889146 Chalasani et al. 2021	Liver disease any etiology (> 87.4 cirrhosis) and HCC
Doylestown	Prospective ($n = 120$)	50	90	0.5	0.98	PMID: 30169533 Mehta et al. 2018	Cirrhosis with any etiology
HES	Prospective ($n = 397$), transi- tion from phase 2 to 3	34.6	90	10.05	0.71	PMID: 34618932 Singal et al. 2022b	Cirrhosis any etiology, HCC
HES	Prospective (n = 534), phase 3	36.7-41.2	90	7.94–8.03	0.76–0.82	PMID: 35124267 Tayob et al. 2023	Cirrhosis any etiology, HCC
HES V2.0	Prospective (n=2331), phase 3	47.2	90	1.27	0.77	PMID: 38899967 El-Serag et al., 2024	Cirrhosis any etiology, HCC
5hmc markers (wd-score)	Prospective (n = 1120), phase 2	82.7	67.4	27.9	0.846	PMID: 31358576 Cai et al. 2019	HBV, cirrhosis and HCC
Multi-target panel (methyl- ated DNA plus protein)	Retro/prospective (n=437), phase 2	80	90	67	0.92	PMID: 32889146 Chalasani et al. 2021	Liver disease any etiology (>87.4 cirrhosis) and HCC

*Notes refer to the main groups included in the studies, specifically detailing the composition of the control groups

al. 2022). In a larger phase 3 cohort of 534 patients, the HES score showed improved sensitivity, ranging from 36.7 to 41.2%, with a consistently high specificity of 90% and AUROC values between 0.76 and 0.82, indicating enhanced performance in a broader population (Tayob et al. 2023).

The updated HES v2.0, which incorporates AFP-L3 and DCP in addition to the original HES components, was evaluated in a large phase 3 prospective cohort of 2,331 patients with cirrhosis. This newer version showed an increased sensitivity of 47.2% while maintaining the specificity at 90% and achieving an AUROC of 0.77 (El-Serag et al., 2024). These results suggest that HES v2 provides a modest improvement over the original HES score in detecting HCC, especially in more extensive and diverse patient populations with cirrhosis, potentially enhancing its utility in clinical practice for early HCC detection.

Imaging markers MRI

Magnetic Resonance Imaging (MRI) is highly effective for detecting hepatocellular carcinoma (HCC). A metaanalysis of 15 studies involving 2,807 patients showed that MRI demonstrated high diagnostic accuracy with a pooled per-patient sensitivity of 86% and specificity of 94%, while per-lesion sensitivity was 77%. This diagnostic performance was consistent across different MRI protocols, both with and without contrast enhancement, and was superior to ultrasound, which had a sensitivity of 53% (Gupta et al. 2021). Similar findings were reported from another study involving 22 studies and 1685 patients, mentioning that multi-sequence noncontrast MRI (NC-MRI) achieved a pooled per-patient sensitivity of 86.8% and specificity of 90.3%. NC-MRI also maintained high sensitivity for detecting smaller lesions (<2 cm) at 77.1%, compared to 88.5% for lesions > 2 cm (Chan et al. 2022). The application of deep learning to interpret MRI images is rapidly advancing, achieving

high diagnostic performance and potentially aiding less experienced radiologists in early HCC detection.

Vibration-controlled transient elastography (VCTE)

Liver stiffness measurement (LSM) by vibration-controlled transient elastography (VCTE, FibroScan) which is a non-invasive diagnostic biomarker of liver fibrosis, is a promising method for HCC risk stratification in cirrhosis. Recently, a retrospective study (n = 1850)reported that the HCC risk in HCV cirrhotic patients after a sustained virological response (SVR) was positively correlated with the increase of LSM, especially for those patients with LSM above 10 kPa (John et al. 2024). Another Swedish multi-center cohort study (n = 14414)further supported that increased LSM was significantly associated with increased HCC risk for patients with cirrhosis and chronic liver diseases across different etiologies (Hegmar et al. 2024). An LSM-based machine learning algorithm displayed superior performance for stratifying 5-year HCC risk among patients with chronic liver disease (AUROC of 0.89), which was separately validated in the Hong Kong (n = 2732) and Europe (n = 2384) cohorts, and was significantly better than other existing HCC risk scores such as aMAP score, Toronto HCC risk index, and 7 hepatitis B-related risk scores (Lin et al. 2024).

Al-enhanced imaging and biomarker integration

Artificial intelligence (AI)-enhanced CT scans improve diagnostic accuracy, achieving sensitivities up to 89.4% in complex cases. AI-assisted imaging holds the potential for automated HCC detection, particularly for earlystage disease, thus supporting more timely and accurate diagnosis. A deep-learning AI system trained on CT images from 7,512 patients, validated and achieved an area under the receiver-operating characteristic curve (AUROC) of 0.887 and 0.883 for the internal and external test sets, respectively. The AI system demonstrated high accuracy (81.0% for the internal test set and 81.3% for the external test set) and high sensitivity (78.4% for the internal test set and 89.4% for the external test set) (Wang et al. 2021a).

AI-based analysis has also refined our tissue-based assessment of TGFBR2 in cirrhotic versus HCC tissue samples, demonstrating reduced TGFBR2 levels as a promising biomarker for HCC detection (Zaidi et al. 2022). Our AI-enhanced model improved accuracy (sensitivity of 0.7, specificity of 0.54) and revealed a reduction in TGFBR2 in HCC compared to cirrhotic tissue, highlighting its potential as a diagnostic tool.

Cost-effectiveness of biomarker-based screening

Several studies and models suggest HCC surveillance using magnetic resonance imaging (MRI) and/ or ultrasound is cost-effective in patients with compensated cirrhosis (Goossens et al. 2017; Nahon et al. 2022), particularly when considering quality-adjusted life years (QALY) gained. Latest cost-effectiveness analysis supports the potential viability of future biomarker-based HCC screening (Singal et al. 2024). Both ultrasound/AFP and biomarker-based screening strategies are cost-effective compared to no screening at a willingness-to-pay threshold of \$150,000/QALY. However, biomarker-based screening demonstrated a lower incremental cost-effectiveness ratio and has been favored in a greater proportion of simulations. Sensitivity analyses reveal that screening adherence, costs, and sensitivity for early-stage HCC detection influenced the cost-effectiveness of the evaluated strategies. The cost of screening per qualityadjusted life years decreases with increasing HCC risk, making it crucial to accurately stratify patients and consider factors like etiology and disease stage when making screening decisions.

Limitations in current research

Several methodological challenges exist across HCC biomarker studies. Many studies rely on derivation samples without independent validation, leading to optimistic AUROC estimates that may not generalize to broader populations. Additionally, multi-center studies often fail to account for center effects, which can introduce confounding variables. Model calibration and handling of outliers are also inconsistently reported, potentially skewing diagnostic accuracy. Combinatorial strategies for the surveillance and diagnosis of HCC, exemplified by the GALAD and HES algorithms, which incorporate AFP alongside additional biomarkers, have shown considerable promise following rigorous phase 3 prospective validations. Nonetheless, the multitude of variables-including protein markers, clinical characteristics, and varied analytical methodologies-can lead to the development of predictive models that may demonstrate comparable performance metrics but differ significantly in their applicability across distinct geographical regions, etiological contexts, and stages of disease. While the GALAD and HES biomarker panel demonstrate improved HCC detection sensitivity in patients with chronic liver disease (Piratvisuth et al. 2023; Chalasani et al. 2021), their performance (sensitivity, specificity, and AUROC) decreases substantially in high-risk cirrhotic cohorts (Tayob et al. 2023; EI-Serag et al., 2024; Beudeker et al. 2023), implying the need for further refinement and validation in specific high-risk populations. Furthermore, ongoing biomarker studies under phase 3 face similar challenges, such as incomplete cohort data, potential selection bias during sample acquisition, and limitations in sample sizes for both discovery and validation cohorts. The inherent heterogeneity of HCC, encompassing a

wide range of underlying etiologies and risk factors, further complicates the quest for a single, universally applicable biomarker. This complexity highlights the imperative for a comprehensive approach to HCC diagnosis and monitoring, one that integrates a variety of biomarkers, particularly those with biological relevance, and clinical data to enhance the specificity and sensitivity of detection across diverse patient populations. Adherence to rigorous epidemiological standards, such as STROBE guidelines, could improve the reliability and applicability of future research. It should be noted that the final effectiveness of those potential HCC surveillance strategy will be further challenged as the US-based screening when applicated in resources-limited settings (Parikh et al. 2023), such as lacking experienced health providers, varied image visualization and biomarker test performance, and with patients lacking up-to-date knowledge and low adherence.

Conclusions

The landscape of HCC biomarkers is evolving, driven by advances in molecular biology, genomics, and AIenhanced imaging. While traditional biomarkers like AFP and DCP remain valuable, new candidates from the fields of circulating DNA, the gut microbiome, and diagnostic algorithms hold the promise of improved sensitivity and specificity. The molecular heterogeneity and complex signaling pathways underlying HCC present both challenges and opportunities in biomarker development. Advances in research in pathways such as the TGF- β members, together with animal models, have provided valuable insights into HCC pathogenesis, paving the way for biological biomarker strategies (Fig. 2). Our ongoing work in serum proteomics, informed by TGF- β pathway components, provides a new foundation for novel predictive models that enhance risk stratification of HCC patients. Integrating multi-modal data encompassing proteomic and imaging biomarkers with established clinical parameters, coupled with advanced AI-driven analytical approaches, offers a promising avenue for refining risk stratification algorithms and ultimately improving patient outcomes. Future research efforts should prioritize the validation of these biomarkers across large-scale prospective studies assessing their diagnostic performance in cohorts with diverse populations and etiologies. Ultimately, these findings should be translated into precision HCC surveillance and therapeutic strategies tailored to individual risk profiles. Robust collaborations across institutions and industries will be critical to advancing these biomarkers from the bench to the clinic.

Abbreviations

HCC	Hepatocellular carcinoma
AFP	a-fetoprotein
US	Ultrasound

MASLD/MASH	Metabolic dysfunction-associated steatotic liver disease
	steatohepatitis
HBV	Hepatitis B
HCV	Hepatitis C
МАРК	Mitogen-activated protein kinase
PI3K	Phosphoinositide 3-kinase
IGF-β	Iransforming growth factor
TCGA	The human Cancer Genome Atlas
DCP	Des-γ-carboxy prothrombin
GP73	Golgi protein-73
GPC3	Glypican-3
ERK	Extracellular signal-regulated kinase
ANG2	Angiopoietin-2
SCCA	Squamous Cell Carcinoma Antigen
ctDNA	Circulating tumor DNA
COSMIC	Somatic Mutations in Cancer
MSTN	Myostatin
ACLF	Acute-on chronic liver failure
HMGB1	Serum high mobility group box 1 protein
HMGB2	High mobility group box 2
COL1A1	Collagen type I ɑ1
EMT	Epithelial-to-mesenchymal transition
5-hmC	5-Hydroxymethylcytosine
MDM	Methylated DNA marker
cfDNA	Cell-free DNA
BCLC	Barcelona Clinic Liver Cancer
NC-MRI	Non-contrast MRI
VCTE	Vibration-controlled transient elastography
LSM	Liver stiffness measurement
Al	Artificial intelligence

Acknowledgements

Not applicable.

Author contributions

LM contributed to the topic formulation. KL, BM, ES, PG, XX, and LM contributed to the design of the review, sampling of relevant studies, information collection and drafting the article. KL, BM, ES, PG, XX, ARK, SD, HH, and LM revised and edited the article critically for presentation, interpretation, discussion, and implication for future research. The corresponding author attests that all listed authors meet authorship criteria and that no others meeting the criteria have been omitted.

Funding

This work was supported by NIH grants R01AA023146 (L. Mishra), R01CA236591 (L. Mishra), and U01CA230690 (L. Mishra).

Data availability

No datasets were generated or analysed during the current study.

Declarations

Ethics approval and consent to participate Not applicable.

Consent for publication

Not applicable.

Competing interests

The authors declare no competing interests.

Received: 9 December 2024 / Accepted: 1 April 2025 Published online: 23 April 2025

References

Abu El Makarem MA, Abdel-Aleem A, Ali A, Saber R, Shatat M, Rahem DA, et al. Diagnostic significance of plasma osteopontin in hepatitis C virus-related hepatocellular carcinoma. Ann Hepatol. 2011;10(3):296–305. https://doi.org/1 0.1016/S1665-2681(19)31541-8.

- Abu El-Makarem MA, Kamel MF, Mohamed AA, Ali HA, Mohamed MR, Mohamed AEM, et al. Down-regulation of hepatic expression of GHR/STAT5/IGF-1 signaling pathway fosters development and aggressiveness of HCV-related hepatocellular carcinoma: crosstalk with Snail-1 and type 2 transforming growth factor-beta receptor. PLoS ONE. 2022;17(11):e0277266. https://doi.org /10.1371/journal.pone.0277266.
- Adebayo Michael AO, Ko S, Tao J, Moghe A, Yang H, Xu M, et al. Inhibiting Glutamine-Dependent mTORC1 activation ameliorates liver cancers driven by β -Catenin mutations. Cell Metab. 2019;29(5):1135–e11501136. https://doi.org/10.1016/j.cmet.2019.01.002.
- Alberts CJ, Clifford GM, Georges D, Negro F, Lesi OA, Hutin YJ, et al. Worldwide prevalence of hepatitis B virus and hepatitis C virus among patients with cirrhosis at country, region, and global levels: a systematic review. Lancet Gastroenterol Hepatol. 2022;7(8):724–35.https://doi.org/10.1016/s2468-125 3(22)00050-4.
- Ao J, Chiba T, Kanzaki H, Kanayama K, Shibata S, Kurosugi A, et al. Serum angiopoietin 2 acts as a diagnostic and prognostic biomarker in hepatocellular carcinoma. J Cancer. 2021;12(9):2694–701. https://doi.org/10.7150/jca.56436.
- Arriazu E, Ge X, Leung TM, Magdaleno F, Lopategi A, Lu Y, et al. Signalling via the osteopontin and high mobility group box-1 axis drives the fibrogenic response to liver injury. Gut. 2017;66(6):1123–37. https://doi.org/10.1136/gu tjnl-2015-310752.
- Atkins JL, Pilling LC, Masoli JAH, Kuo CL, Shearman JD, Adams PC, et al. Association of hemochromatosis HFE p.C282Y homozygosity with hepatic malignancy. JAMA. 2020;324(20):2048–57. https://doi.org/10.1001/jama.2020.21566.
- Barton JC, McLaren CE, Chen WP, Ramm GA, Anderson GJ, Powell LW, et al. Cirrhosis in hemochromatosis: independent risk factors in 368 HFE p.C282Y homozygotes. Ann Hepatol. 2018;17(5):871–9. https://doi.org/10.5604/01.30 01.0012.3169.
- Beudeker BJB, Fu S, Balderramo D, Mattos AZ, Carrera E, Diaz J, et al. Validation and optimization of AFP-based biomarker panels for early HCC detection in Latin America and Europe. Hepatol Commun. 2023;7(10). https://doi.org/10.1097/ hc9.00000000000264.
- Biswas S, Chytil A, Washington K, Romero-Gallo J, Gorska AE, Wirth PS, et al. Transforming growth factor beta receptor type II inactivation promotes the establishment and progression of colon cancer. Cancer Res. 2004;64(14):4687–92. https://doi.org/10.1158/0008-5472.CAN-03-3255.
- Boonkaew B, Satthawiwat N, Pinjaroen N, Chuaypen N, Tangkijvanich P. Circulating extracellular Vesicle-Derived MicroRNAs as novel diagnostic and prognostic biomarkers for Non-Viral-Related hepatocellular carcinoma. Int J Mol Sci. 2023;24(22). https://doi.org/10.3390/ijms242216043.
- Bray F, Laversanne M, Sung H, Ferlay J, Siegel RL, Soerjomataram I, et al. Global cancer statistics 2022: GLOBOCAN estimates of incidence and mortality worldwide for 36 cancers in 185 countries. CA Cancer J Clin. 2024;74(3):229– 63. https://doi.org/10.3322/caac.21834.
- Cai J, Chen L, Zhang Z, Zhang X, Lu X, Liu W, et al. Genome-wide mapping of 5-hydroxymethylcytosines in Circulating cell-free DNA as a noninvasive approach for early detection of hepatocellular carcinoma. Gut. 2019;68(12):2195–205. https://doi.org/10.1136/gutjnl-2019-318882.
- Cai Z, Zhang J, He Y, Xia L, Dong X, Chen G, et al. Liquid biopsy by combining 5-hydroxymethylcytosine signatures of plasma cell-free DNA and protein biomarkers for diagnosis and prognosis of hepatocellular carcinoma. ESMO Open. 2021;6(1):100021. https://doi.org/10.1016/j.esmoop.2020.100021.
- Campani C, Imbeaud S, Couchy G, Ziol M, Hirsch TZ, Rebouissou S, et al. Circulating tumour DNA in patients with hepatocellular carcinoma across tumour stages and treatments. Gut. 2024;73(11):1870–82. https://doi.org/10.1136/gutjnl-20 24-331956.
- Cancer Genome Atlas Research Network. Comprehensive and integrative genomic characterization of hepatocellular carcinoma. Cell. 2017;169(7):1327–41. e1323.
- Cao J, Huang Y, Zhu M, Wang Z, Jin Z, Xiong Z. Causal association of sarcopenia with hepatocellular carcinoma risk in European population: a Mendelian randomization study. Front Nutr. 2024;11:1292834. https://doi.org/10.3389/f nut.2024.1292834.
- Castillo LF, Tascón R, Lago Huvelle MR, Novack G, Llorens MC, Dos Santos AF, et al. Glypican-3 induces a mesenchymal to epithelial transition in human breast cancer cells. Oncotarget. 2016;7(37):60133–54. https://doi.org/10.18632/onc otarget.11107.
- Chalasan^T NP, Ramasubramanian TS, Bhattacharya A, Olson MC, Edwards VD, Roberts LR, et al. A novel Blood-Based panel of methylated DNA and protein markers for detection of Early-Stage hepatocellular carcinoma. Clin

Gastroenterol Hepatol. 2021;19(12):2597-e26052594. https://doi.org/10.1016 /j.cgh.2020.08.065.

- Chalasani NP, Porter K, Bhattacharya A, Book AJ, Neis BM, Xiong KM, et al. Validation of a novel multitarget blood test shows high sensitivity to detect early stage hepatocellular carcinoma. Clin Gastroenterol Hepatol. 2022;20(1):173– e182177. https://doi.org/10.1016/j.cgh.2021.08.010.
- Chan MV, Huo YR, Trieu N, Mitchelle A, George J, He E et al. Noncontrast MRI for hepatocellular carcinoma detection: A systematic review and Meta-analysis - A potential surveillance tool?? Clin gastroenterol hepatol. 2022, 20(1), 44– e5642. https://doi.org/10.1016/j.cgh.2021.02.036.
- Chang KC, Lin MT, Wang JH, Hung CH, Chen CH, Chiu SY, et al. HBcrAg predicts hepatocellular carcinoma development in chronic B hepatitis related liver cirrhosis patients undergoing Long-Term effective Anti-Viral. Viruses. 2022;14(12). https://doi.org/10.3390/v14122671.
- Chen L, Li M, Li Q, Wang CJ, Xie SQ. DKK1 promotes hepatocellular carcinoma cell migration and invasion through β-catenin/MMP7 signaling pathway. Mol Cancer. 2013;12:157. https://doi.org/10.1186/1476-4598-12-157.
- Chen X, Wang Y, Tao J, Shi Y, Gai X, Huang F, et al. mTORC1 Up-Regulates GP73 to promote proliferation and migration of hepatocellular carcinoma cells and growth of xenograft tumors in mice. Gastroenterology. 2015;149(3):741–e752714. https://doi.org/10.1053/j.gastro.2015.05.005.
- Chen J, Yao ZX, Chen JS, Gi YJ, Munoz NM, Kundra S, et al. TGF-beta/beta2-spectrin/ CTCF-regulated tumor suppression in human stem cell disorder Beckwith-Wiedemann syndrome. J Clin Invest. 2016;126(2):527–42. https://doi.org/10. 1172/JCI80937.
- Chen J, Zaidi S, Rao S, Chen JS, Phan L, Farci P, et al. Analysis of genomes and transcriptomes of hepatocellular carcinomas identifies mutations and gene expression changes in the transforming growth Factor-β pathway. Gastroenterology. 2018;154(1):195–210. https://doi.org/10.1053/j.gastro.2017.09.007.
- Chen S, Li J, Tan X, Xu Q, Mo Y, Qin H, et al. Clinical role of combining alpha-fetoprotein and lens culinaris agglutinin-reactive fraction of alpha-fetoprotein for hepatocellular carcinoma: evidence from literature and an original study. J Clin Lab Anal. 2020;34(7):e23262. https://doi.org/10.1002/jcla.23262.
- Chen R, Kang R, Tang D. The mechanism of HMGB1 secretion and release. Exp Mol Med. 2022;54(2):91–102. https://doi.org/10.1038/s12276-022-00736-w.
- Cho EEL, Ang CZ, Quek J, Fu CE, Lim LKE, Heng ZEQ, et al. Global prevalence of non-alcoholic fatty liver disease in type 2 diabetes mellitus: an updated systematic review and meta-analysis. Gut. 2023;72(11):2138–48. https://doi.or g/10.1136/gutjnl-2023-330110.
- Choi K, Jang HY, Ahn JM, Hwang SH, Chung JW, Choi YS, et al. The association of the serum levels of myostatin, follistatin, and interleukin-6 with sarcopenia, and their impacts on survival in patients with hepatocellular carcinoma. Clin Mol Hepatol. 2020;26(4):492–505. https://doi.org/10.3350/cmh.2020.0005.
- Choi GH, Jang ES, Kim JW, Jeong SH. Prognostic role of plasma level of angiopoietin-1, angiopoietin-2, and vascular endothelial growth factor in hepatocellular carcinoma. World J Gastroenterol. 2021;27(27):4453–67. https://doi.org/1 0.3748/wjg.v27.i27.4453.
- Colnot S, Decaens T, Niwa-Kawakita M, Godard C, Hamard G, Kahn A, et al. Livertargeted disruption of apc in mice activates beta-catenin signaling and leads to hepatocellular carcinomas. Proc Natl Acad Sci U S A. 2004;101(49):17216– 21. https://doi.org/10.1073/pnas.0404761101.
- Cui Y, Zhang M, Guo J, Jin J, Wang H, Wang X. Correlation between sarcopenia and cirrhosis: a meta-analysis. Front Nutr. 2023;10:1342100. https://doi.org/10.338 9/fnut.2023.1342100.
- David CJ, Huang YH, Chen M, Su J, Zou Y, Bardeesy N, et al. TGF-beta tumor suppression through a lethal EMT. Cell. 2016;164(5):1015–30. https://doi.org/10.1 016/j.cell.2016.01.009.
- Desert R, Ge X, Song Z, Han H, Lantvit D, Chen W, et al. Role of Hepatocyte-Derived osteopontin in liver carcinogenesis. Hepatol Commun. 2022;6(4):692–709. htt ps://doi.org/10.1002/hep4.1845.
- Dhanasekaran R, Suzuki H, Lemaitre L, Kubota N, Hoshida Y. Molecular and immune landscape of hepatocellular carcinoma to guide therapeutic decision-making. Hepatology. 2025;81(3):1038–57. https://doi.org/10.1097/H EP.000000000000513.
- Ding DY, Jiang SY, Zu YX, Yang Y, Gan XJ, Yuan SX, et al. Collagen in hepatocellular carcinoma: A novel biomarker and therapeutic target. Hepatol Commun. 2024;8(7). https://doi.org/10.1097/hc9.00000000000489.
- Du A, Li S, Zhou Y, Disoma C, Liao Y, Zhang Y, et al. M6A-mediated upregulation of circmdk promotes tumorigenesis and acts as a nanotherapeutic target in hepatocellular carcinoma. Mol Cancer. 2022;21(1):109. https://doi.org/10.118 6/s12943-022-01575-z.

Eldeeb MK, Magour GM, Bedair RN, Shamseya MM, Hammouda MA. Study of Dickkopf-1 (DKK-1) in patients with chronic viral hepatitis C-related liver cirrhosis with and without hepatocellular carcinoma. Clin Exp Hepatol. 2020;6(2):85– 91. https://doi.org/10.5114/ceh.2020.95831.

Esfeh JM, Hajifathalian K, Ansari-Gilani K. Sensitivity of ultrasound in detecting hepatocellular carcinoma in obese patients compared to explant pathology as the gold standard. Clin Mol Hepatol. 2020;26(1):54–9. https://doi.org/10.33 50/cmh.2019.0039.

Flemming JA, Djerboua M, Groome PA, Booth CM, Terrault NA. NAFLD and Alcohol-Associated liver disease will be responsible for almost all new diagnoses of cirrhosis in Canada by 2040. Hepatology. 2021;74(6):3330–44. https://doi.org /10.1002/hep.32032.

Fouad YM, Mohamed HI, Kamal EM, Rasek MA. Clinical significance and diagnostic value of serum dickkopf-1 in patients with hepatocellular carcinoma. Scand J Gastroenterol. 2016;51(9):1133–7. https://doi.org/10.3109/00365521.2016. 1172337.

Fujikawa T, Shiraha H, Ueda N, Takaoka N, Nakanishi Y, Matsuo N, et al. Des-gammacarboxyl prothrombin-promoted vascular endothelial cell proliferation and migration. J Biol Chem. 2007;282(12):8741–8. https://doi.org/10.1074/jbc.M 609358200.

Gatselis NK, Tornai T, Shums Z, Zachou K, Saitis A, Gabeta S, et al. Golgi protein-73: A biomarker for assessing cirrhosis and prognosis of liver disease patients. World J Gastroenterol. 2020;26(34):5130–45. https://doi.org/10.3748/wjg.v2 6.i34.5130.

Ghosh S, Bhowmik S, Majumdar S, Goswami A, Chakraborty J, Gupta S, et al. The exosome encapsulated MicroRNAs as Circulating diagnostic marker for hepatocellular carcinoma with low alpha-fetoprotein. Int J Cancer. 2020;147(10):2934–47. https://doi.org/10.1002/ijc.33111.

Goossens N, Singal AG, King LY, Andersson KL, Fuchs BC, Besa C, et al. Cost-Effectiveness of risk Score-Stratified hepatocellular carcinoma screening in patients with cirrhosis. Clin Transl Gastroenterol. 2017;8(6):e101. https://doi.or g/10.1038/ctg.2017.26.

Gough NR, Xiang X, Mishra L. TGF-beta signaling in liver, pancreas, and Gastrointestinal diseases and cancer. Gastroenterology. 2021;161(2):434–e452415. https:/ /doi.org/10.1053/j.gastro.2021.04.064.

Gu S, Zaidi S, Hassan MI, Mohammad T, Malta TM, Noushmehr H, et al. Mutated CEACAMs disrupt transforming growth factor beta signaling and alter the intestinal Microbiome to promote colorectal carcinogenesis. Gastroenterology. 2020;158(1):238–52. https://doi.org/10.1053/j.gastro.2019.09.023.

Guarino M, Di Costanzo GG, Gallotta A, Tortora R, Paneghetti L, Auriemma F, et al. Circulating SCCA-IgM complex is a useful biomarker to predict the outcome of therapy in hepatocellular carcinoma patients. Scand J Clin Lab Invest. 2017;77(6):448–53. https://doi.org/10.1080/00365513.2017.1336569.

Gupta P, Soundararajan R, Patel A, Kumar MP, Sharma V, Kalra N. Abbreviated MRI for hepatocellular carcinoma screening: A systematic review and meta-analysis. J Hepatol. 2021;75(1):108–19. https://doi.org/10.1016/j.jhep.2021.01.041.

Hadi H, Wan Shuaib WMA, Raja Ali RA, Othman H. Utility of PIVKA-II and AFP in differentiating hepatocellular carcinoma from Non-Malignant High-Risk patients. Med (Kaunas). 2022;58(8). https://doi.org/10.3390/medicina58081 015.

He L, Tian DA, Li PY, He XX. Mouse models of liver cancer: progress and recommendations. Oncotarget. 2015;6(27):23306–22. https://doi.org/10.18632/on cotarget.4202.

He L, Zhang C, Liu LL, Huang LP, Lu WJ, Zhang YY, et al. Development of a diagnostic nomogram for alpha-fetoprotein-negative hepatocellular carcinoma based on serological biomarkers. World J Gastrointest Oncol. 2024;16(6):2463–75. https://doi.org/10.4251/wjgo.v16.i6.2463.

Hegmar H, Wester A, Aleman S, Backman J, Degerman E, Ekvall H, et al. Liver stiffness predicts progression to liver-related events in patients with chronic liver disease - A cohort study of 14 414 patients. Liver Int. 2024;44(7):1689–99. http s://doi.org/10.1111/liv.15919.

Hernandez BY, Zhu X, Risch HA, Lu L, Ma X, Irwin ML, et al. Oral cyanobacteria and hepatocellular carcinoma. Cancer Epidemiol Biomarkers Prev. 2022;31(1):221– 9. https://doi.org/10.1158/1055-9965.Epi-21-0804.

Horie Y, Suzuki A, Kataoka E, Sasaki T, Hamada K, Sasaki J, et al. Hepatocyte-specific Pten deficiency results in steatohepatitis and hepatocellular carcinomas. J Clin Invest. 2004;113(12):1774–83. https://doi.org/10.1172/jci20513. Hou J, Berg T, Vogel A, Piratvisuth T, Trojan J, De Toni EN, et al. Comparative evaluation of multimarker algorithms for early-stage HCC detection in multicenter prospective studies. JHEP Rep. 2025;7(2):101263. https://doi.org/10.1016/j.jh epr.2024.101263.

Huang JH, Wang J, Chai XQ, Li ZC, Jiang YH, Li J, et al. The intratumoral bacterial metataxonomic signature of hepatocellular carcinoma. Microbiol Spectr. 2022;10(5):e0098322. https://doi.org/10.1128/spectrum.00983-22.

Huang Y, Liangpunsakul S, Rudraiah S, Ma J, Keshipeddy SK, Wright D, et al. HMGB2 is a potential diagnostic marker and therapeutic target for liver fibrosis and cirrhosis. Hepatol Commun. 2023;7(11). https://doi.org/10.1097/hc9.00000 000000299.

Jang ES, Jeong SH, Kim JW, Choi YS, Leissner P, Brechot C. Diagnostic performance of Alpha-Fetoprotein, protein induced by vitamin K absence, osteopontin, Dickkopf-1 and its combinations for hepatocellular carcinoma. PLoS ONE. 2016;11(3):e0151069. https://doi.org/10.1371/journal.pone.0151069.

Jin X, Zhang S, Wang N, Guan L, Shao C, Lin Y, et al. High expression of TGF-β1 contributes to hepatocellular carcinoma prognosis via regulating tumor immunity. Front Oncol. 2022;12:861601. https://doi.org/10.3389/fonc.2022. 861601.

Jinato T, Anuntakarun S, Satthawiwat N, Chuaypen N, Tangkijvanich P. Distinct alterations of gut microbiota between viral- and non-viral-related hepatocellular carcinoma. Appl Microbiol Biotechnol. 2024;108(1):34. https://doi.org/10 .1007/s00253-023-12845-1.

John BV, Dang Y, Kaplan DE, Jou JH, Taddei TH, Spector SA, et al. Liver stiffness measurement and risk prediction of hepatocellular carcinoma after HCV eradication in veterans with cirrhosis. Clin Gastroenterol Hepatol. 2024;22(4):778– e788777. https://doi.org/10.1016/j.cgh.2023.11.020.

Johnson PJ, Pirrie SJ, Cox TF, Berhane S, Teng M, Palmer D, et al. The detection of hepatocellular carcinoma using a prospectively developed and validated model based on serological biomarkers. Cancer Epidemiol Biomarkers Prev. 2014;23(1):144–53. https://doi.org/10.1158/1055-9965.Epi-13-0870.

Kao JT, Feng CL, Yu CJ, Tsai SM, Hsu PN, Chen YL, et al. IL-6, through p-STAT3 rather than p-STAT1, activates hepatocarcinogenesis and affects survival of hepatocellular carcinoma patients: a cohort study. BMC Gastroenterol. 2015;15:50. ht tps://doi.org/10.1186/s12876-015-0283-5.

Katz SF, Lechel A, Obenauf AC, Begus-Nahrmann Y, Kraus JM, Hoffmann EM, et al. Disruption of Trp53 in livers of mice induces formation of carcinomas with bilineal differentiation. Gastroenterology. 2012;142(5):1229–e12391223. https: //doi.org/10.1053/j.gastro.2012.02.009.

Katz LH, Likhter M, Jogunoori W, Belkin M, Ohshiro K, Mishra L. TGF-beta signaling in liver and Gastrointestinal cancers. Cancer Lett. 2016;379(2):166–72. https:// doi.org/10.1016/j.canlet.2016.03.033.

Kim JH, Kang SH, Lee M, Youn GS, Kim TS, Jun BG et al. Serum myostatin predicts the risk of hepatocellular carcinoma in patients with alcoholic cirrhosis: A multicenter study. Cancers (Basel). 2020, 12(11). https://doi.org/10.3390/can cers12113347.

Kim AK, Hamilton JP, Lin SY, Chang TT, Hann HW, Hu CT, et al. Urine DNA biomarkers for hepatocellular carcinoma screening. Br J Cancer. 2022a;126(10):1432– 8. https://doi.org/10.1038/s41416-022-01706-9.

Kim HI, Lim J, Shim JH. Role of the alpha-fetoprotein response in immune checkpoint inhibitor-based treatment of patients with hepatocellular carcinoma. J Cancer Res Clin Oncol. 2022b;148(8):2069–77. https://doi.org/10.1007/s0043 2-021-03727-y.

Kim DY, Toan BN, Tan CK, Hasan I, Setiawan L, Yu ML, et al. Utility of combining PIVKA-II and AFP in the surveillance and monitoring of hepatocellular carcinoma in the Asia-Pacific region. Clin Mol Hepatol. 2023;29(2):277–92. https:// doi.org/10.3350/cmh.2022.0212.

Kisiel JB, Dukek BA, R V S R K, Ghoz HM, Yab TC, Berger CK, et al. Hepatocellular carcinoma detection by plasma methylated DNA: discovery, phase I pilot, and phase II clinical validation. Hepatology. 2019;69(3):1180–92. https://doi.o rg/10.1002/hep.30244.

Konyn P, Ahmed A, Kim D. Current epidemiology in hepatocellular carcinoma. Expert Rev Gastroenterol Hepatol. 2021;15(11):1295–307. https://doi.org/10.1 080/17474124.2021.1991792.

Korkut A, Zaidi S, Kanchi RS, Rao S, Gough NR, Schultz A, et al. A Pan-Cancer analysis reveals High-Frequency genetic alterations in mediators of signaling by the TGF-β superfamily. Cell Syst. 2018;7(4):422–e437427. https://doi.org/10 .1016/j.cels.2018.08.010.

Kwon JH, Kim J, Park JY, Hong SM, Park CW, Hong SJ, et al. Overexpression of highmobility group box 2 is associated with tumor aggressiveness and prognosis of hepatocellular carcinoma. Clin Cancer Res. 2010;16(22):5511–21. https://do i.org/10.1158/1078-0432.Ccr-10-0825. Lapidot Y, Amir A, Nosenko R, Uzan-Yulzari A, Veitsman E, Cohen-Ezra O, et al. Alterations in the gut Microbiome in the progression of cirrhosis to hepatocellular carcinoma. mSystems. 2020;5(3). https://doi.org/10.1128/mSystems. 00153-20.

Lee HA, Lee YR, Lee YS, Jung YK, Kim JH, An H, et al. Lens culinaris agglutininreactive fraction of alpha-fetoprotein improves diagnostic accuracy for hepatocellular carcinoma. World J Gastroenterol. 2021a;27(28):4687–96. https ://doi.org/10.3748/wjg.v27.i28.4687.

Lee YT, Wang JJ, Luu M, Noureddin M, Kosari K, Agopian VG, et al. The mortality and overall survival trends of primary liver cancer in the united States. J Natl Cancer Inst. 2021b;113(11):1531–41. https://doi.org/10.1093/jnci/djab079.

- Li B, Liu H, Shang HW, Li P, Li N, Ding HG. Diagnostic value of glypican-3 in alpha Fetoprotein negative hepatocellular carcinoma patients. Afr Health Sci. 2013;13(3):703–9. https://doi.org/10.4314/ahs.v13i3.26.
- Li XJ, Li QL, Ju LG, Zhao C, Zhao LS, Du JW, et al. Deficiency of histone methyltransferase SET Domain-Containing 2 in liver leads to abnormal lipid metabolism and HCC. Hepatology. 2021;73(5):1797–815. https://doi.org/10.1002/hep.31 594.
- Li X, Sun X, Kan C, Chen B, Qu N, Hou N, et al. COL1A1: A novel oncogenic gene and therapeutic target in malignancies. Pathol Res Pract. 2022;236:154013. ht tps://doi.org/10.1016/j.prp.2022.154013.
- Liebman HA, Furie BC, Tong MJ, Blanchard RA, Lo KJ, Lee SD, et al. Des-gammacarboxy (abnormal) prothrombin as a serum marker of primary hepatocellular carcinoma. N Engl J Med. 1984;310(22):1427–31. https://doi.org/10.1056 /nejm198405313102204.
- Lin H, Li G, Delamarre A, Ahn SH, Zhang X, Kim BK, et al. A liver Stiffness-Based Etiology-Independent machine learning algorithm to predict hepatocellular carcinoma. Clin Gastroenterol Hepatol. 2024;22(3):602–e610607. https://doi.o rg/10.1016/j.cgh.2023.11.005.
- Liu F, Zhang Y, Peng Z, Gao H, Xu L, Chen M. High expression of high mobility group box 1 (hmgb1) predicts poor prognosis for hepatocellular carcinoma after curative hepatectomy. J Transl Med. 2012;10:135. https://doi.org/10.118 6/1479-5876-10-135.
- Liu Y, Sethi NS, Hinoue T, Schneider BG, Cherniack AD, Sanchez-Vega F, et al. Comparative molecular analysis of Gastrointestinal adenocarcinomas. Cancer Cell. 2018;33(4):721–e735728. https://doi.org/10.1016/j.ccell.2018.03.010.
- Liu J, Jiang J, Mo J, Liu D, Cao D, Wang H, et al. Global DNA 5-Hydroxymethylcytosine and 5-Formylcytosine contents are decreased in the early stage of hepatocellular carcinoma. Hepatology. 2019;69(1):196–208. https://doi.org/1 0.1002/hep.30146.
- Liu S, Wang M, Zheng C, Zhong Q, Shi Y, Han X. Diagnostic value of serum glypican-3 alone and in combination with AFP as an aid in the diagnosis of liver cancer. Clin Biochem. 2020;79:54–60. https://doi.org/10.1016/j.clinbiochem. 2020.02.009.
- Llovet JM, Peña CE, Lathia CD, Shan M, Meinhardt G, Bruix J. Plasma biomarkers as predictors of outcome in patients with advanced hepatocellular carcinoma. Clin Cancer Res. 2012;18(8):2290–300. https://doi.org/10.1158/1078-0432.Cc r-11-2175.
- Lu Q, Li J, Cao H, Lv C, Wang X, Cao S. Comparison of diagnostic accuracy of midkine and AFP for detecting hepatocellular carcinoma: a systematic review and meta-analysis. Biosci Rep. 2020;40(3). https://doi.org/10.1042/bsr20192 424.
- Lu K, Zhao T, Yang L, Liu Y, Ruan X, Cui L, et al. HMGB2 upregulation promotes the progression of hepatocellular carcinoma cells through the activation of ZEB1/vimentin axis. J Gastrointest Oncol. 2023;14(5):2178–91. https://doi.org /10.21037/jgo-23-447.

Lund SA, Giachelli CM, Scatena M. The role of osteopontin in inflammatory processes. J Cell Commun Signal. 2009;3(3–4):311–22. https://doi.org/10.1007/s 12079-009-0068-0.

- Ma HP, Chang HL, Bamodu OA, Yadav VK, Huang TY, Wu ATH et al. Collagen 1A1 (COL1A1) is a reliable biomarker and putative therapeutic target for hepatocellular carcinogenesis and metastasis. Cancers (Basel). 2019, 11(6). https://do i.org/10.3390/cancers11060786.
- Maggio-Price L, Treuting P, Zeng W, Tsang M, Bielefeldt-Ohmann H, Iritani BM. Helicobacter infection is required for inflammation and colon cancer in SMAD3-deficient mice. Cancer Res. 2006;66(2):828–38. https://doi.org/10.115 8/0008-5472.Can-05-2448.
- Malta TM, Sokolov A, Gentles AJ, Burzykowski T, Poisson L, Weinstein JN, et al. Machine learning identifies stemness features associated with oncogenic dedifferentiation. Cell. 2018;173(2):338–e354315. https://doi.org/10.1016/j.c ell.2018.03.034.

- Marrero JA, Feng Z, Wang Y, Nguyen MH, Befeler AS, Roberts LR, et al. Alpha-fetoprotein, des-gamma carboxyprothrombin, and lectin-bound alpha-fetoprotein in early hepatocellular carcinoma. Gastroenterology. 2009;137(1):110–8. https://doi.org/10.1053/j.gastro.2009.04.005.
- Mehta AS, Lau DT, Wang M, Aslam A, Nasir B, Javaid A, et al. Application of the Doylestown algorithm for the early detection of hepatocellular carcinoma. PLoS ONE. 2018;13(8):e0203149. https://doi.org/10.1371/journal.pone.0203 149.
- Mohamed AA, Abo-Amer YE-E, Aalkhalegy A, Fathalla LA, Elmaghraby MB, Elhoseeny MM, et al. COL1A1 gene expression in hepatitis B virus (HBV) related hepatocellular carcinoma (HCC) Egyptian's patients. Open Biomarkers J. 2021;11:108–14. https://doi.org/10.2174/1875318302111010108.
- Myojin Y, Kodama T, Sakamori R, Maesaka K, Matsumae T, Sawai Y, et al. Interleukin-6 is a Circulating prognostic biomarker for hepatocellular carcinoma patients treated with combined immunotherapy. Cancers (Basel). 2022;14(4). https://doi.org/10.3390/cancers14040883.
- Nahon P, Najean M, Layese R, Zarca K, Segar LB, Cagnot C, et al. Early hepatocellular carcinoma detection using magnetic resonance imaging is cost-effective in high-risk patients with cirrhosis. JHEP Rep. 2022;4(1):100390. https://doi.org/10.1016/j.jhepr.2021.100390.
- Parikh ND, Tayob N, Singal AG. Blood-based biomarkers for hepatocellular carcinoma screening: approaching the end of the ultrasound era? J Hepatol. 2023;78(1):207–16. https://doi.org/10.1016/j.jhep.2022.08.036.
- Passaro A, Al Bakir M, Hamilton EG, Diehn M, André F, Roy-Chowdhuri S, et al. Cancer biomarkers: emerging trends and clinical implications for personalized treatment. Cell. 2024;187(7):1617–35. https://doi.org/10.1016/j.cell.202 4.02.041.
- Philips CA, Rajesh S, Nair DC, Ahamed R, Abduljaleel JK, Augustine P. Hepatocellular carcinoma in 2021: an exhaustive update. Cureus. 2021;13(11):e19274. https:/ /doi.org/10.7759/cureus.19274.
- Piratvisuth T, Hou J, Tanwandee T, Berg T, Vogel A, Trojan J, et al. Development and clinical validation of a novel algorithmic score (GAAD) for detecting HCC in prospective cohort studies. Hepatol Commun. 2023;7(11). https://doi.org/10. 1097/hc9.000000000000317.
- Pozzan C, Cardin R, Piciocchi M, Cazzagon N, Maddalo G, Vanin V, et al. Diagnostic and prognostic role of SCCA-IgM serum levels in hepatocellular carcinoma (HCC). J Gastroenterol Hepatol. 2014;29(8):1637–44. https://doi.org/10.1111 /jgh.12576.
- Qiu S, Cai J, Yang Z, He X, Xing Z, Zu J, et al. Trends in hepatocellular carcinoma mortality rates in the US and projections through 2040. JAMA Netw Open. 2024;7(11):e2445525. https://doi.org/10.1001/jamanetworkopen.2024;45525.
- Rao S, Yang X, Ohshiro K, Zaidi S, Wang Z, Shetty K, et al. beta2-spectrin (SPTBN1) as a therapeutic target for diet-induced liver disease and preventing cancer development. Sci Transl Med. 2021;13(624):eabk2267. https://doi.org/10.1126 /scitranslmed.abk2267.
- Ruiz-Margáin A, Pohlmann A, Lanzerath S, Langheinrich M, Campos-Murguía A, Román-Calleja BM, et al. Myostatin is associated with the presence and development of acute-on-chronic liver failure. JHEP Rep. 2023;5(8):100761. ht tps://doi.org/10.1016/j.jhepr.2023.100761.
- Saharinen P, Eklund L, Alitalo K. Therapeutic targeting of the angiopoietin-TIE pathway. Nat Rev Drug Discov. 2017;16(9):635–61. https://doi.org/10.1038/n rd.2016.278.
- Schlaeger C, Longerich T, Schiller C, Bewerunge P, Mehrabi A, Toedt G, et al. Etiology-dependent molecular mechanisms in human hepatocarcinogenesis. Hepatology. 2008;47(2):511–20. https://doi.org/10.1002/hep.22033.
- Seo SH, Cho KJ, Park HJ, Kim H, Lee HW, Kim BK, et al. Dickkopf-1 promotes angiogenesis by upregulating VEGF receptor 2-mediated mTOR/p70S6K signaling in hepatocellular carcinoma. Am J Cancer Res. 2021;11(10):4788–806.
- Shakiba E, Ramezani M, Sadeghi M. Evaluation of serum interleukin-6 levels in hepatocellular carcinoma patients: a systematic review and meta-analysis. Clin Exp Hepatol. 2018;4(3):182–90. https://doi.org/10.5114/ceh.2018.78122.
- Shen Q, Fan J, Yang XR, Tan Y, Zhao W, Xu Y, et al. Serum DKK1 as a protein biomarker for the diagnosis of hepatocellular carcinoma: a large-scale, multicentre study. Lancet Oncol. 2012;13(8):817–26. https://doi.org/10.1016/ s1470-2045(12)70233-4.
- Simão A, Madaleno J, Silva N, Rodrigues F, Caseiro P, Costa JN, et al. Plasma osteopontin is a biomarker for the severity of alcoholic liver cirrhosis, not for hepatocellular carcinoma screening. BMC Gastroenterol. 2015;15:73. https://d oi.org/10.1186/s12876-015-0307-1.
- Singal AG, Haaland B, Parikh ND, Ozbay AB, Kirshner C, Chakankar S, et al. Comparison of a multitarget blood test to ultrasound and alpha-fetoprotein for

hepatocellular carcinoma surveillance: results of a network meta-analysis. Hepatol Commun. 2022a;6(10):2925–36. https://doi.org/10.1002/hep4.2045.

- Singal AG, Tayob N, Mehta A, Marrero JA, El-Serag H, Jin Q, et al. GALAD demonstrates high sensitivity for HCC surveillance in a cohort of patients with cirrhosis. Hepatology. 2022b;75(3):541–9. https://doi.org/10.1002/hep.32185.
- Singal AG, Chhatwal J, Parikh N, Tapper E. Cost-Effectiveness of a Biomarker-Based screening strategy for hepatocellular carcinoma in patients with cirrhosis. Liver Cancer. 2024;13(6):643–54. https://doi.org/10.1159/000539895.
- Song BC, Chung YH, Kim JA, Choi WB, Suh DD, Pyo SI, et al. Transforming growth factor-beta1 as a useful serologic marker of small hepatocellular carcinoma. Cancer. 2002;94(1):175–80. https://doi.org/10.1002/cncr.10170.
- Song Z, Chen W, Athavale D, Ge X, Desert R, Das S, et al. Osteopontin takes center stage in chronic liver disease. Hepatology. 2021;73(4):1594–608. https://doi.o rg/10.1002/hep.31582.
- Song Y, Lau HC, Zhang X, Yu J. Bile acids, gut microbiota, and therapeutic insights in hepatocellular carcinoma. Cancer Biol Med. 2023;21(2):144–62. https://doi. org/10.20892/j.issn.2095-3941.2023.0394.
- Srivastava J, Siddiq A, Gredler R, Shen XN, Rajasekaran D, Robertson CL, et al. Astrocyte elevated gene-1 and c-Myc cooperate to promote hepatocarcinogenesis in mice. Hepatology. 2015;61(3):915–29. https://doi.org/10.1002/he p.27339.
- Sterling RK, Jeffers L, Gordon F, Sherman M, Venook AP, Reddy KR, et al. Clinical utility of AFP-L3% measurement in North American patients with HCV-related cirrhosis. Am J Gastroenterol. 2007;102(10):2196–205. https://doi.org/10.1111 /i.1572-0241.2007.01405.x.
- Sun B, Hu C, Yang Z, Zhang X, Zhao L, Xiong J, et al. Midkine promotes hepatocellular carcinoma metastasis by elevating Anoikis resistance of Circulating tumor cells. Oncotarget. 2017;8(20):32523–35. https://doi.org/10.18632/onc otarget.15808.
- Sun N, Zhang C, Lee YT, Tran BV, Wang J, Kim H, et al. HCC EV ECG score: an extracellular vesicle-based protein assay for detection of early-stage hepatocellular carcinoma. Hepatology. 2023;77(3):774–88. https://doi.org/10.1002/he p.32692.
- Suzuki M, Shiraha H, Fujikawa T, Takaoka N, Ueda N, Nakanishi Y, et al. Des-gammacarboxy prothrombin is a potential autologous growth factor for hepatocellular carcinoma. J Biol Chem. 2005;280(8):6409–15. https://doi.org/10.1074/j bc.M406714200.
- Suzuki H, Iwamoto H, Tanaka T, Sakaue T, Imamura Y, Masuda A, et al. Fibroblast growth factor Inhibition by molecular-targeted agents mitigates immunosuppressive tissue microenvironment in hepatocellular carcinoma. Hepatol Int. 2024;18(2):610–22. https://doi.org/10.1007/s12072-023-10603-z.
- Tanaka S, Mori M, Sakamoto Y, Makuuchi M, Sugimachi K, Wands JR. Biologic significance of angiopoietin-2 expression in human hepatocellular carcinoma. J Clin Invest. 1999;103(3):341–5. https://doi.org/10.1172/jci4891.
- Tanaka S, Sugimachi K, Yamashita Yi Y, Ohga T, Shirabe K, Shimada M, et al. Tie2 vascular endothelial receptor expression and function in hepatocellular carcinoma. Hepatology. 2002;35(4):861–7. https://doi.org/10.1053/jhep.200 2.32535.
- Tantai X, Liu Y, Yeo YH, Praktiknjo M, Mauro E, Hamaguchi Y, et al. Effect of sarcopenia on survival in patients with cirrhosis: A meta-analysis. J Hepatol. 2022;76(3):588–99. https://doi.org/10.1016/j.jhep.2021.11.006.
- Tao J, Zhang R, Singh S, Poddar M, Xu E, Oertel M, et al. Targeting β-catenin in hepatocellular cancers induced by coexpression of mutant β-catenin and K-Ras in mice. Hepatology. 2017;65(5):1581–99. https://doi.org/10.1002/hep .28975.
- Tayob N, Kanwal F, Alsarraj A, Hernaez R, El-Serag HB. The performance of AFP, AFP-3, DCP as biomarkers for detection of hepatocellular carcinoma (HCC): A phase 3 biomarker study in the united States. Clin Gastroenterol Hepatol. 2023;21(2):415–e423414. https://doi.org/10.1016/j.cgh.2022.01.047.
- Than NN, Ghazanfar A, Hodson J, Tehami N, Coldham C, Mergental H, et al. Comparing clinical presentations, treatments and outcomes of hepatocellular carcinoma due to hepatitis C and non-alcoholic fatty liver disease. QJM. 2017;110(2):73–81. https://doi.org/10.1093/qjmed/hcw151.
- Tripathi A, Shrinet K, Kumar A. HMGB1 protein as a novel target for cancer. Toxicol Rep. 2019;6:253–61. https://doi.org/10.1016/j.toxrep.2019.03.002.
- Tsai JF, Jeng JE, Chuang LY, Yang ML, Ho MS, Chang WY, et al. Clinical evaluation of urinary transforming growth factor-beta1 and serum alpha-fetoprotein as tumour markers of hepatocellular carcinoma. Br J Cancer. 1997;75(10):1460–6. https://doi.org/10.1038/bjc.1997.250.
- Tseng TC, Liu CJ, Hsu CY, Hong CM, Su TH, Yang WT, et al. High level of hepatitis B Core-Related antigen associated with increased risk of hepatocellular carcinoma in patients with chronic HBV infection of intermediate viral load.

Gastroenterology. 2019;157(6):1518–e15291513. https://doi.org/10.1053/j.ga stro.2019.08.028.

- Tzartzeva K, Obi J, Rich NE, Parikh ND, Marrero JA, Yopp A, et al. Surveillance imaging and alpha Fetoprotein for early detection of hepatocellular carcinoma in patients with cirrhosis: A Meta-analysis. Gastroenterology. 2018;154(6):1706– e17181701. https://doi.org/10.1053/j.gastro.2018.01.064.
- Umer M, Qureshi SA, Hashmi ZY, Raza A, Ahmad J, Rahman M, et al. Promoter hypermethylation of Wnt pathway inhibitors in hepatitis C virus - induced multistep hepatocarcinogenesis. Virol J. 2014;11:117. https://doi.org/10.1186 /1743-422x-11-117.
- Venepalli NK, Goff L. Targeting the HGF-cMET Axis in Hepatocellular Carcinoma. Int J Hepatol. 2013, 2013, 341636. https://doi.org/10.1155/2013/341636.
- Wang M, Devarajan K, Singal AG, Marrero JA, Dai J, Feng Z, et al. The Doylestown algorithm: A test to improve the performance of AFP in the detection of hepatocellular carcinoma. Cancer Prev Res (Phila). 2016;9(2):172–9. https://do i.org/10.1158/1940-6207.Capr-15-0186.
- Wang M, Fu F, Zheng B, Bai Y, Wu Q, Wu J, et al. Development of an AI system for accurately diagnose hepatocellular carcinoma from computed tomography imaging data. Br J Cancer. 2021a;125(8):1111–21. https://doi.org/10.1038/s41 416-021-01511-w.
- Wang Z, Hopson LM, Singleton SS, Yang X, Jogunoori W, Mazumder R, et al. Mice with dysfunctional TGF-beta signaling develop altered intestinal Microbiome and colorectal cancer resistant to 5FU. Biochim Biophys Acta Mol Basis Dis. 2021b;1867(10):166179. https://doi.org/10.1016/j.bbadis.2021.166179.
- Wang X, Dong FL, Wang YQ, Wei HL, Li T, Li J. Exosomal circTGFBR2 promotes hepatocellular carcinoma progression via enhancing ATG5 mediated protective autophagy. Cell Death Dis. 2023;14(7):451. https://doi.org/10.1038/s414 19-023-05989-5.
- Wei C, Yang X, Liu N, Geng J, Tai Y, Sun Z, et al. Tumor microenvironment regulation by the Endoplasmic reticulum stress transmission mediator golgi protein 73 in mice. Hepatology. 2019;70(3):851–70. https://doi.org/10.1002/hep.30549.
- Wu Q, Li L, Miao C, Hasnat M, Sun L, Jiang Z, et al. Osteopontin promotes hepatocellular carcinoma progression through inducing JAK2/STAT3/NOX1mediated ROS production. Cell Death Dis. 2022;13(4):341. https://doi.org/10.1 038/s41419-022-04806-9.
- Yang J, He Q, Lu F, Chen K, Ni Z, Wang H, et al. A distinct microbiota signature precedes the clinical diagnosis of hepatocellular carcinoma. Gut Microbes. 2023;15(1):2201159. https://doi.org/10.1080/19490976.2023.2201159.
- Yang X, Bhowmick K, Rao S, Xiang X, Ohshiro K, Amdur RL, et al. Aldehydes alter TGF-beta signaling and induce obesity and cancer. Cell Rep. 2024;43(9):114676. https://doi.org/10.1016/j.celrep.2024.114676.
- Ye J, Gao X, Huang X, Huang S, Zeng D, Luo W et al. Integrating Single-Cell and Spatial transcriptomics to uncover and elucidate GP73-Mediated Pro-Angiogenic regulatory networks in hepatocellular carcinoma. Research (Wash D C). 2024, 7, 0387. https://doi.org/10.34133/research.0387.
- You H, Ding W, Dang H, Jiang Y, Rountree CB. c-Met represents a potential therapeutic target for personalized treatment in hepatocellular carcinoma. Hepatology. 2011;54(3):879–89. https://doi.org/10.1002/hep.24450.
- You L, Zhou J, Xin Z, Hauck JS, Na F, Tang J, et al. Novel directions of precision oncology: Circulating microbial DNA emerging in cancer-microbiome areas. Precis Clin Med. 2022;5(1):pbac005. https://doi.org/10.1093/pcmedi/pbac005.
- Yu B, Yang X, Xu Y, Yao G, Shu H, Lin B, et al. Elevated expression of DKK1 is associated with cytoplasmic/nuclear beta-catenin accumulation and poor prognosis in hepatocellular carcinomas. J Hepatol. 2009;50(5):948–57. https:// doi.org/10.1016/j.jhep.2008.11.020.
- Yu X, Zheng Y, Zhu X, Gao X, Wang C, Sheng Y, et al. Osteopontin promotes hepatocellular carcinoma progression via the PI3K/AKT/Twist signaling pathway. Oncol Lett. 2018;16(4):5299–308. https://doi.org/10.3892/ol.2018.9281.
- Zabransky DJ, Danilova L, Leatherman JM, Lopez-Vidal TY, Sanchez J, Charmsaz S, et al. Profiling of syngeneic mouse HCC tumor models as a framework to understand anti-PD-1 sensitive tumor microenvironments. Hepatology. 2023;77(5):1566–79. https://doi.org/10.1002/hep.32707.
- Zaidi S, Amdur R, Xiang X, Yu H, Wong LL, Rao S, et al. Using quantitative immunohistochemistry in patients at high risk for hepatocellular cancer. Genes Cancer. 2022;13:9–20. https://doi.org/10.18632/genesandcancer.220.
- Zhang BH, Yang BH, Tang ZY. Randomized controlled trial of screening for hepatocellular carcinoma. J Cancer Res Clin Oncol. 2004;130(7):417–22. https://doi.or g/10.1007/s00432-004-0552-0.
- Zhang BH, Li B, Kong LX, Yan LN, Yang JY. Diagnostic accuracy of midkine on hepatocellular carcinoma: A meta-analysis. PLoS ONE. 2019;14(10):e0223514. https ://doi.org/10.1371/journal.pone.0223514.

- Zhang X, Wu LN, Li XQ, Luo X, Liu SW, Zhang L, et al. Whether the golgi protein 73 could be a diagnostic serological marker in hepatocellular carcinoma: a meta analysis. BMC Gastroenterol. 2023;23(1):85. https://doi.org/10.1186/s12876-0 23-02685-8.
- Zhang B, Zhu B, Yu J, Liu H, Zhou Y, Sun G, et al. A combined model of six serum MicroRNAs as diagnostic markers for hepatocellular carcinoma. Clin Chim Acta. 2025;565:119977. https://doi.org/10.1016/j.cca.2024.119977.
- Zheng C, Lu F, Chen B, Yang J, Yu H, Wang D, et al. Gut Microbiome as a biomarker for predicting early recurrence of HBV-related hepatocellular carcinoma. Cancer Sci. 2023;114(12):4717–31. https://doi.org/10.1111/cas.15983.
- Zhou JM, Wang T, Zhang KH. AFP-L3 for the diagnosis of early hepatocellular carcinoma: A meta-analysis. Med (Baltim). 2021;100(43):e27673. https://doi.or g/10.1097/md.00000000027673.
- Zhu M, Zheng J, Wu F, Kang B, Liang J, Heskia F, et al. OPN is a promising serological biomarker for hepatocellular carcinoma diagnosis. J Med Virol. 2020;92(12):3596–603. https://doi.org/10.1002/jmv.25704.

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