

Low concordance of multiple variant-calling pipelines: practical implications for exome and genome sequencing

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Acknowledgements



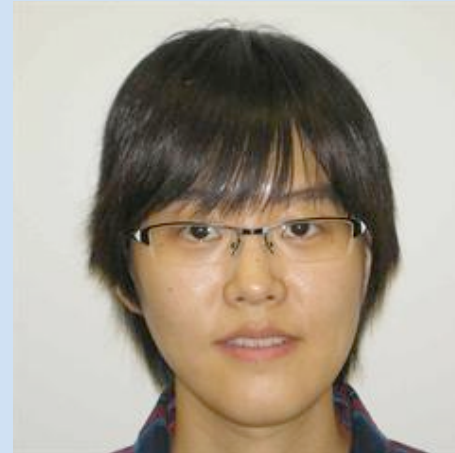
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Jason O'Rawe



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Eric Antoniou
Dick McCombie
Sequencing core facility



Tao Jiang
Guangqing Sun

Results from Exome and WGS requires both Analytic and Clinical Validity

- Analytical Validity: the test is accurate with high sensitivity and specificity.
- Clinical Validity: Given an accurate test result, what impact and/or outcome does this have on the individual person?

Illusions of Certainty. Everything is Probabilistic.

GERD GLIGERENZER

HOW TO KNOW WHEN
NUMBERS DECEIVE YOU

CALCULATED

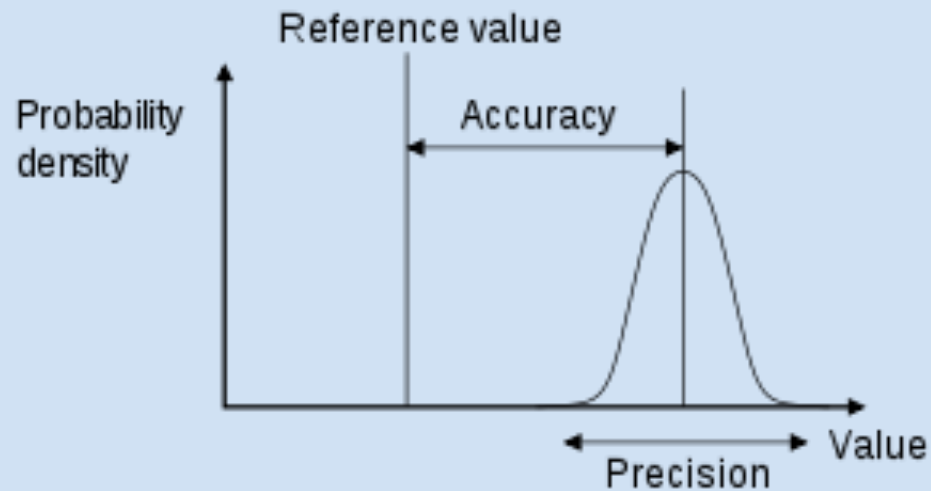
RISKS
RISKS

CALCULATED

NUMBERS DECEIVE YOU

HOW TO KNOW WHEN

GERD GLIGERENZER







High accuracy, but low precision



High precision, but low accuracy

In the fields of [science](#), [engineering](#), [industry](#), and [statistics](#), the **accuracy** of a [measurement](#) system is the degree of closeness of measurements of a [quantity](#) to that quantity's actual (true) [value](#). The **precision** of a measurement system, also called [reproducibility](#) or [repeatability](#), is the degree to which repeated measurements under unchanged conditions show the same [results](#).

http://en.wikipedia.org/wiki/Accuracy_and_precision

	Accurate	Inaccurate (systematic error)
Precise		
Imprecise (reproducibility error)		

Accuracy

$$\text{accuracy} = \frac{\text{number of true positives} + \text{number of true negatives}}{\text{number of true positives} + \text{false positives} + \text{false negatives} + \text{true negatives}}$$

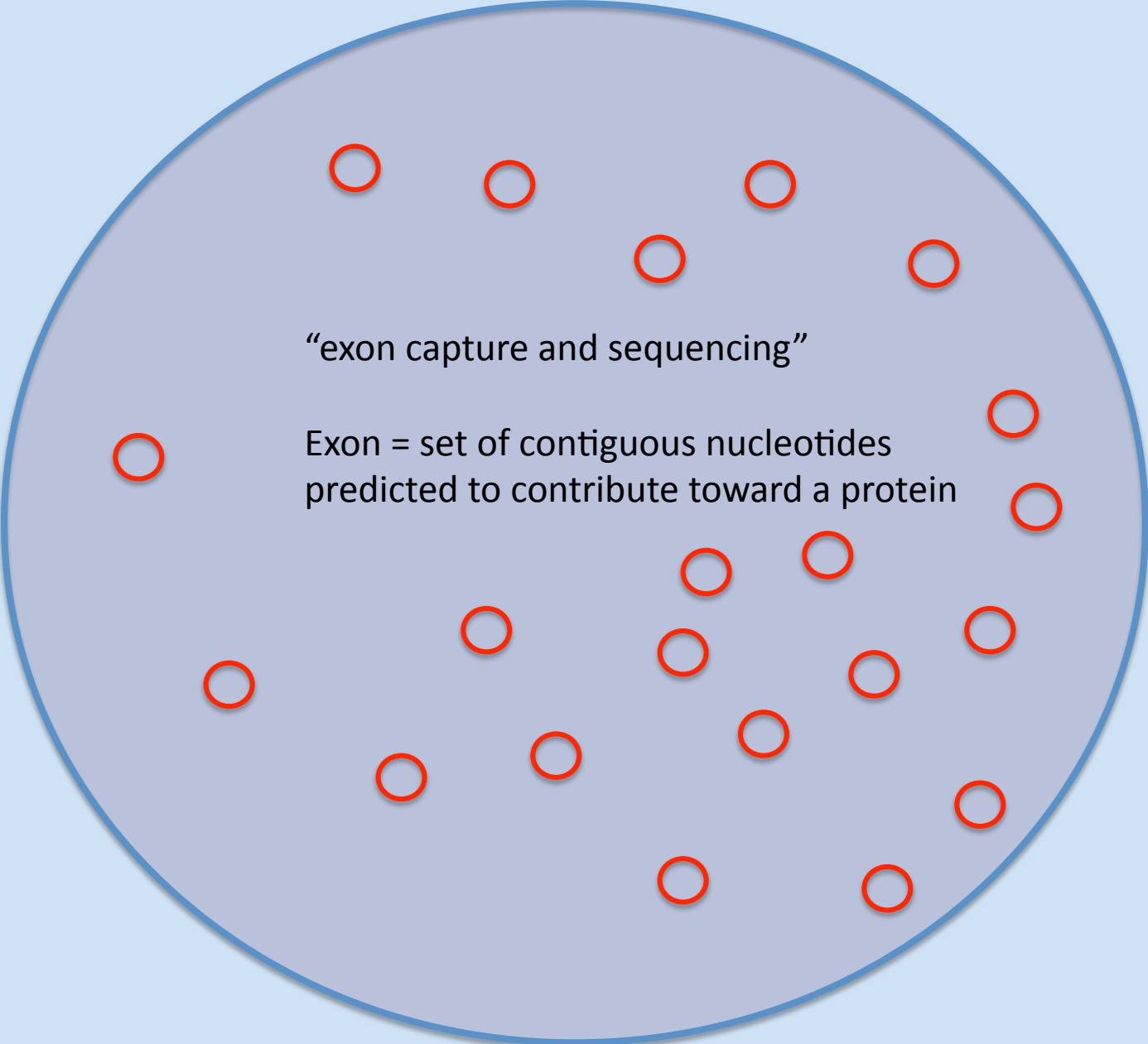
An accuracy of 100% means that the measured values are exactly the same as the given values.

True negative

True positive

“ground truth” Genome from blood of one person
(of course, that is from millions of cells and only blood,
not other tissues)

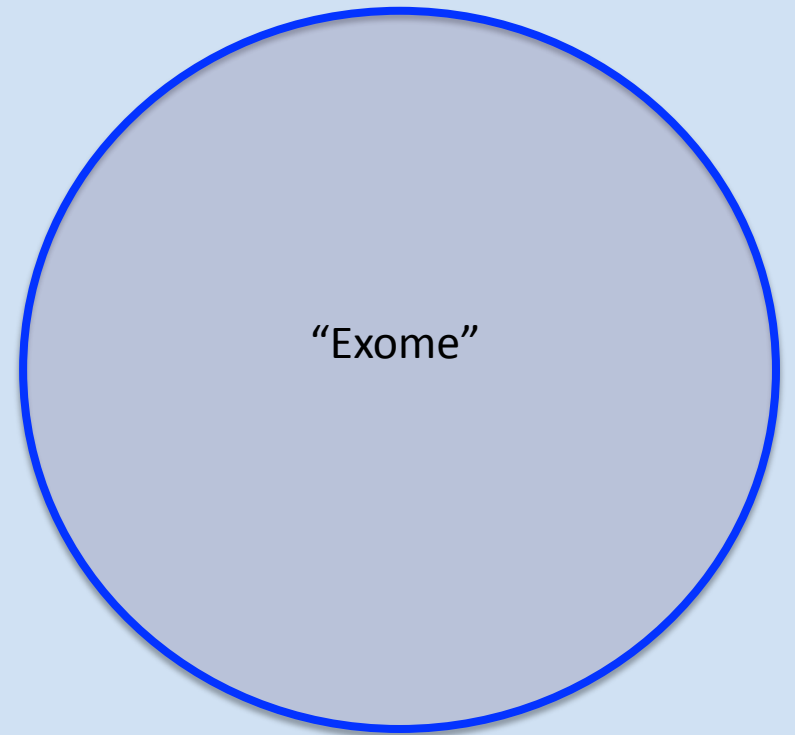
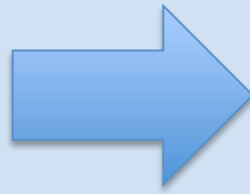
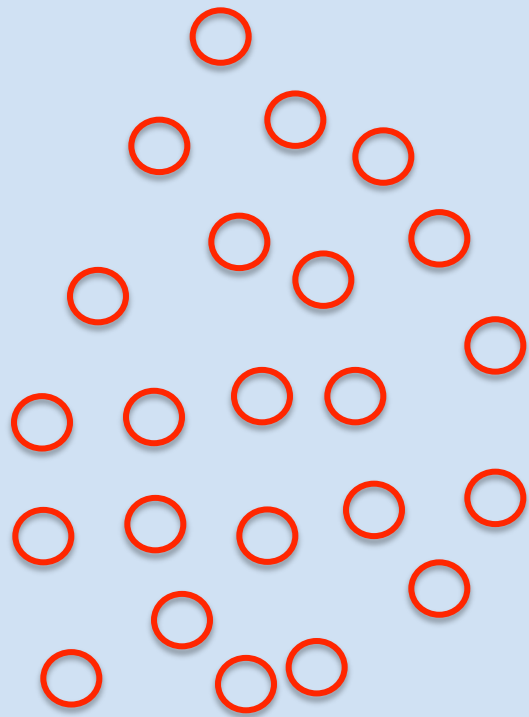
~3 billion nucleotides



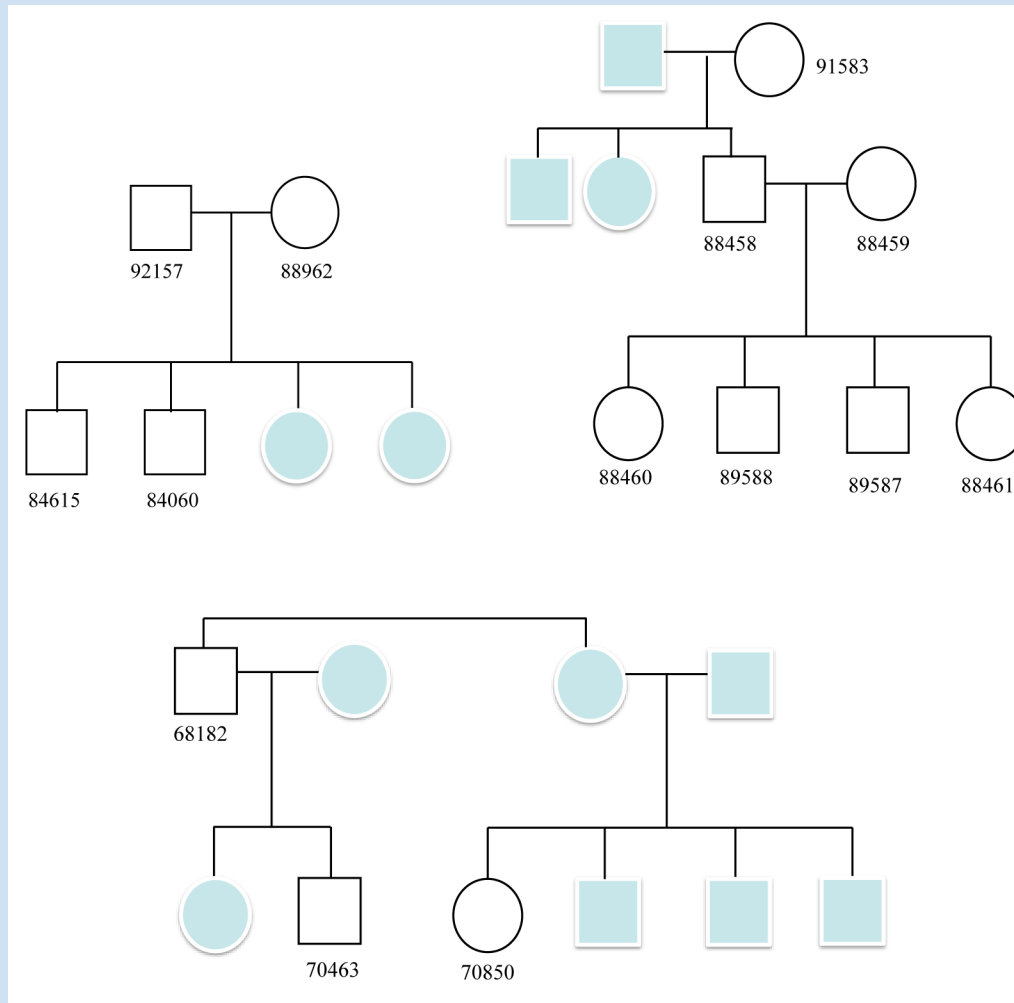
“exon capture and sequencing”

The diagram shows a large light blue circle representing a cell. Inside this circle is a darker blue circle representing the nucleus. Scattered throughout the cell are approximately 20 small red circles, which represent exons. The text “exon capture and sequencing” is centered within the cell.

Exon = set of contiguous nucleotides
predicted to contribute toward a protein



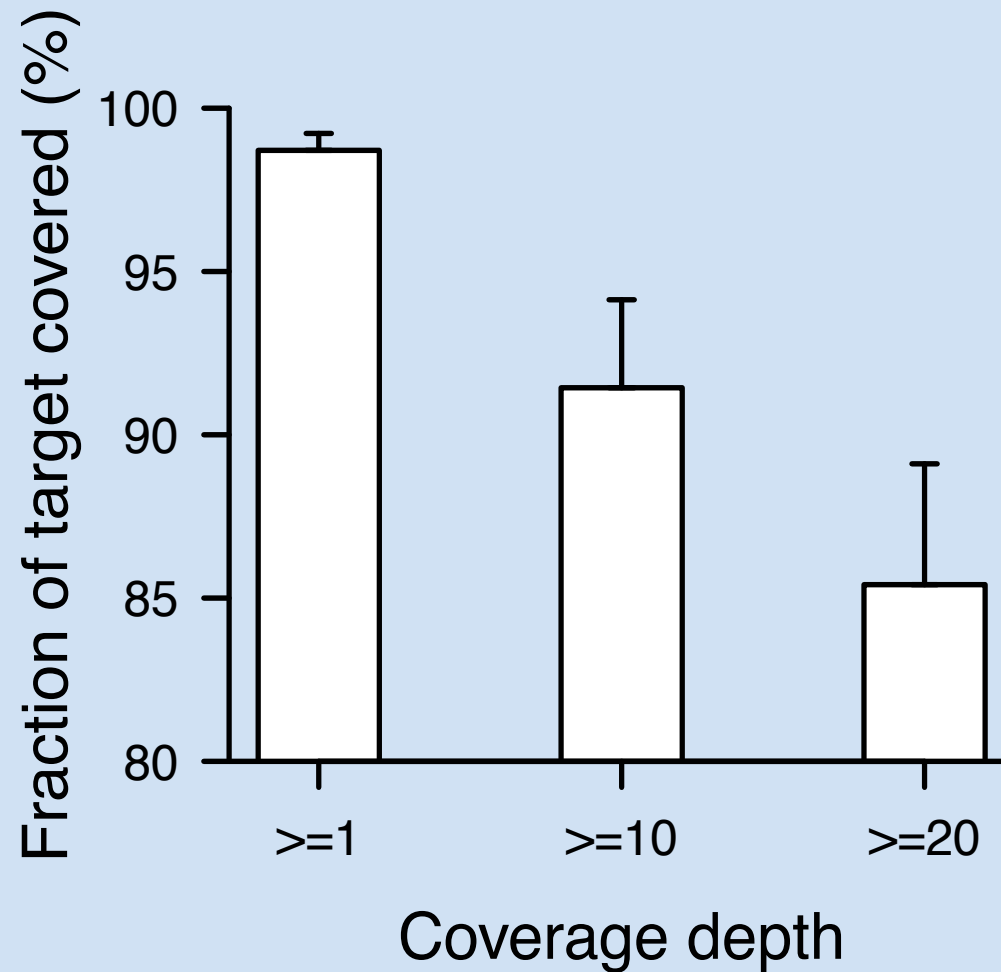
Chose to sequence 15 “exomes”



2-3 rounds of sequencing at BGI to attain goal of >80% of target region at >20 reads per base pair

Exome Capture Statistics	K24510-84060	K24510-92157-a	K24510-84615	K24510-88962
Target region (bp)	46,401,121	46,401,121	46,401,121	46,257,379
Raw reads	138,779,950	161,898,170	156,985,870	104,423,704
Raw data yield (Mb)	12,490	14,571	14,129	9,398
Reads mapped to genome	110,160,277	135,603,094	135,087,576	83,942,646
Reads mapped to target region	68,042,793	84,379,239	80,347,146	61,207,116
Data mapped to target region (Mb)	5,337.69	6,647.18	6,280.01	4,614.47
Mean depth of target region	115.03	143.25	135.34	99.76
Coverage of target region (%)	0.9948	0.9947	0.9954	0.9828
Average read length (bp)	89.91	89.92	89.95	89.75
Fraction of target covered >=4X	98.17	98.38	98.47	94.25
Fraction of target covered >=10X	95.18	95.90	95.97	87.90
Fraction of target covered >=20X	90.12	91.62	91.75	80.70
Fraction of target covered >=30X	84.98	87.42	87.67	74.69
Capture specificity (%)	61.52	62.12	59.25	73.16
Fraction of unique mapped bases on or near target	65.59	65.98	63.69	85.46
Gender test result	M	M	M	F

Depth of Coverage in 15 exomes > 20 reads per bp in target region



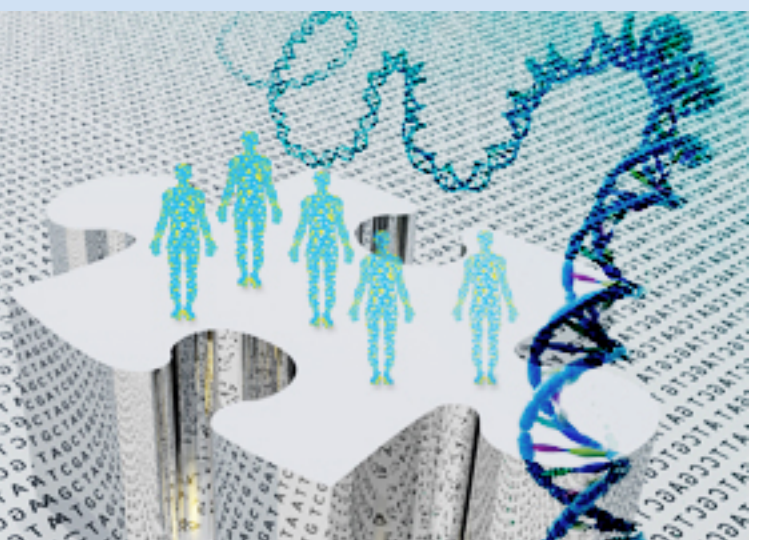
Experimental Design

- Evaluate robustness of variant calling implemented by different bioinformatics analysts.
- Looking at False Positives and False Negatives.
- How reliable are variants that are uniquely called by individual pipelines?
- Are some pipelines better at detecting rare, or novel variants than others?

Human Exome Sequencing Promotion

**50X: \$899/sample 100X: \$1299/sample
(SNP & Indel Included)**

38,000 Exomes Sequenced by BGI to Date



Promotion Details (valid for Americas and Europe customers NOW through MAY 31)

A. The 899 USD/sample package – 50X human exome sequencing

- Agilent SureSelect 50/51M Capture kit
- 100 bp paired-end sequencing on HiSeq 2000
- 5 Gb high quality* sequencing data
- 50X average coverage for target regions guaranteed
- SNP & Indel calling and annotation included

B. The 1299 USD/sample package – 100X human exome sequencing

- Agilent SureSelect 50/51M Capture kit
- 100 bp paired-end sequencing on HiSeq 2000
- 10 Gb high quality* sequencing data
- 100X average coverage for target regions guaranteed
- SNP & Indel calling and annotation included

Table 1. A descriptive summary of the variant calling pipelines included in the comparative analyses.

Pipeline name	Alignment method	Variant-calling module	Description of variant-calling algorithm
SOAP	SOAPaligner version 2.21/ BWA version 0.5.9	SOAPsnr version 1.03/ SOAPindel version 2.01	SOAP uses a method based on Bayes' theorem to call consensus genotype by carefully considering the data quality, alignment, and recurring experimental errors [22].
GATK version 1.5	BWA version 0.5.9	UnifiedGenotyper version 1.5	GATK employs a general Bayesian framework to distinguish and call variants. Error correction models are guided by expected characteristics of human variation to further refine variant calls [19].
SNVer version 0.2.1	BWA version 0.5.9	SNVer version 0.2.1	SNVer uses a more general frequentist framework, and formulates variant calling as a hypothesis-testing problem [25].
GNUMAP version 3.1.0	GNUMAP version 3.1.0	GNUMAP version 3.1.0	GNUMAP incorporates the base uncertainty of the reads into mapping analysis using a probabilistic Needleman-Wunsch algorithm [24].
SAMtools version 0.1.18	BWA version 0.5.9	mpileup version 0.1.18	SAMtools [20] calls variants by generating a consensus sequence using the MAQ model framework, which uses a general Bayesian framework for picking the base that maximizes the posterior probability with the highest Phred quality score.

Accuracy



Precision

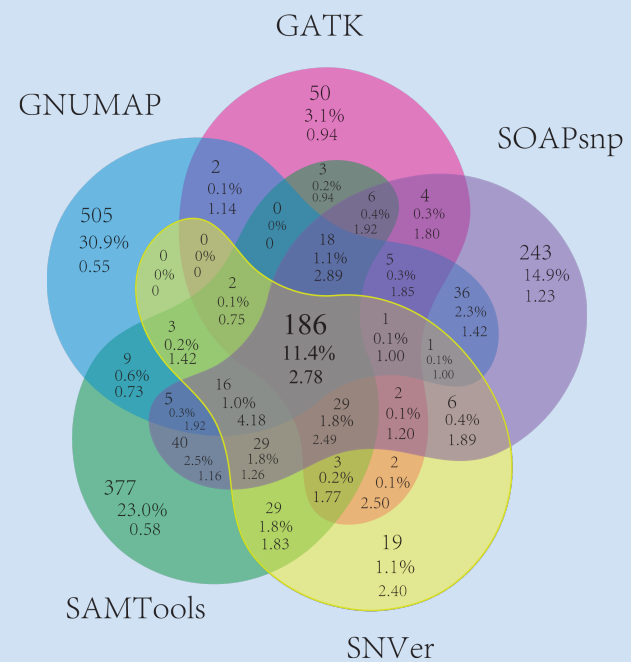
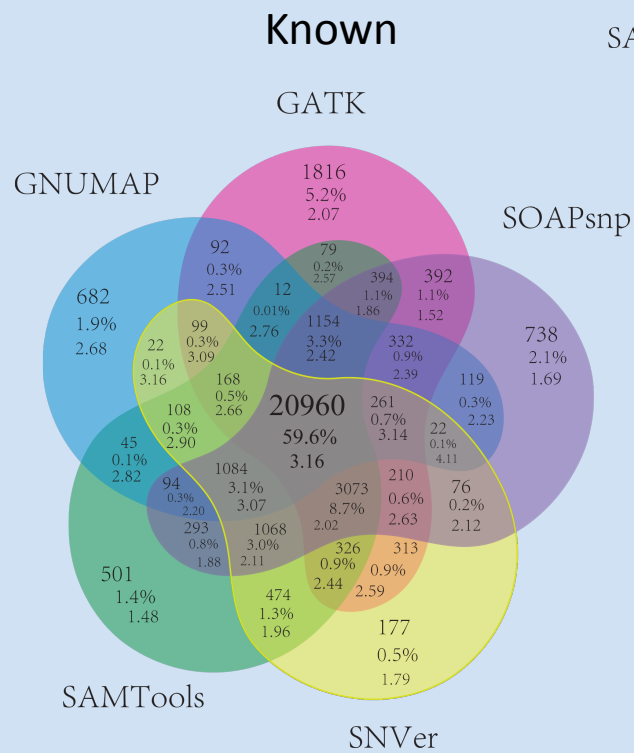
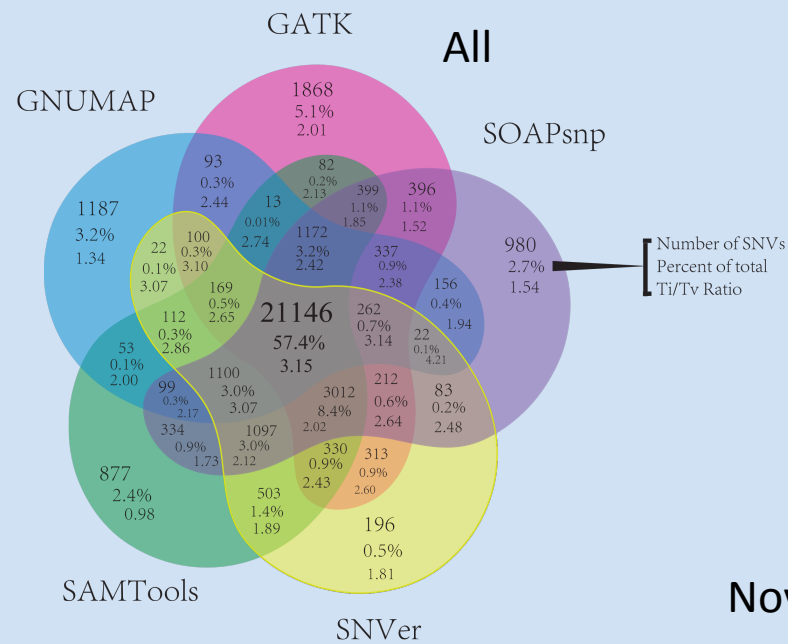


Error-Prone & Doubtful

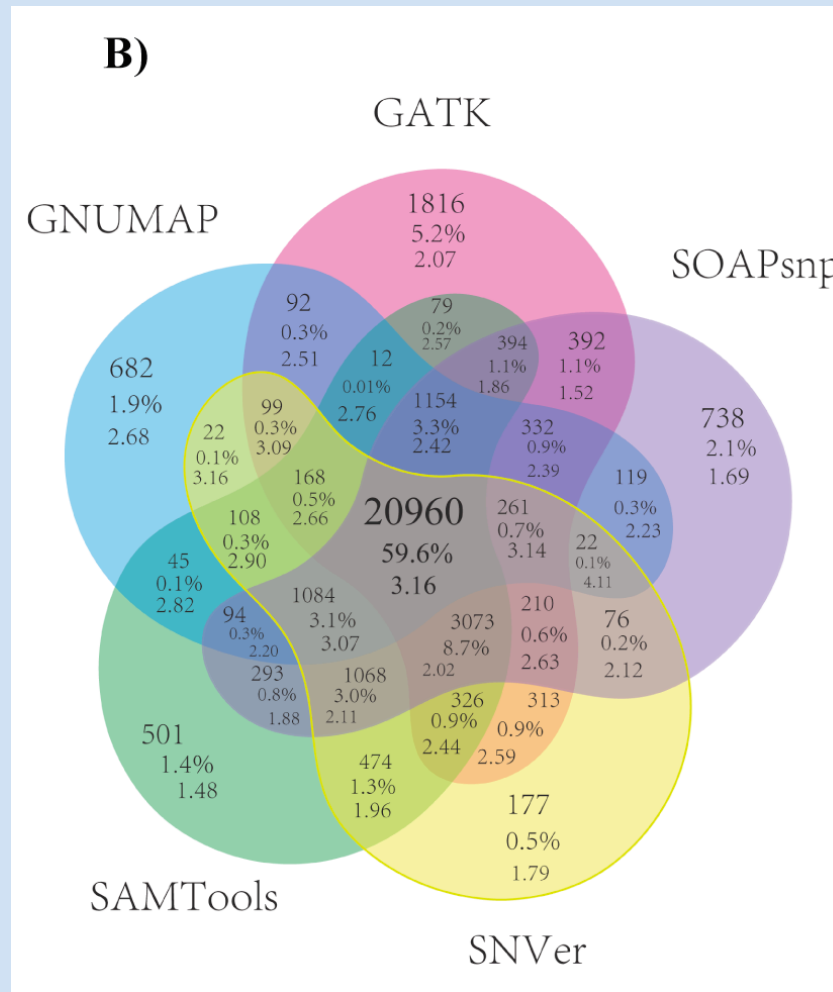


Repeatable & Reproducible

Images: Wikipedia

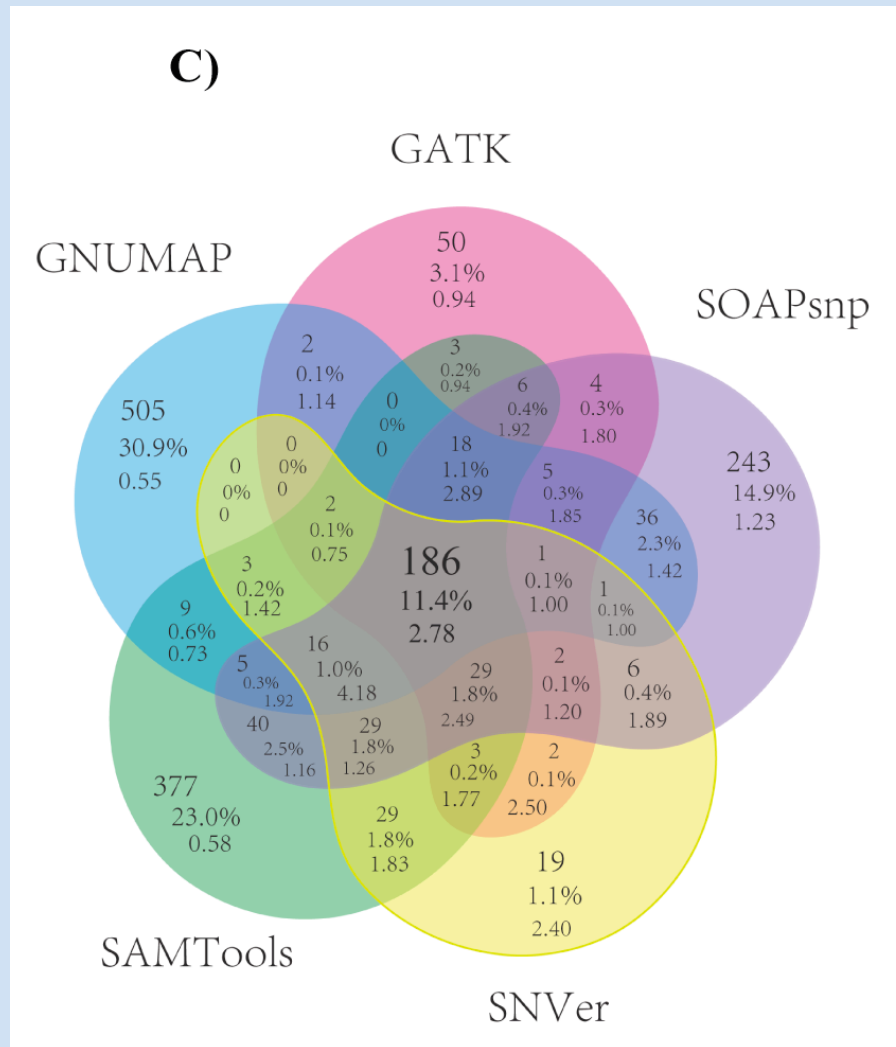


Known SNVs



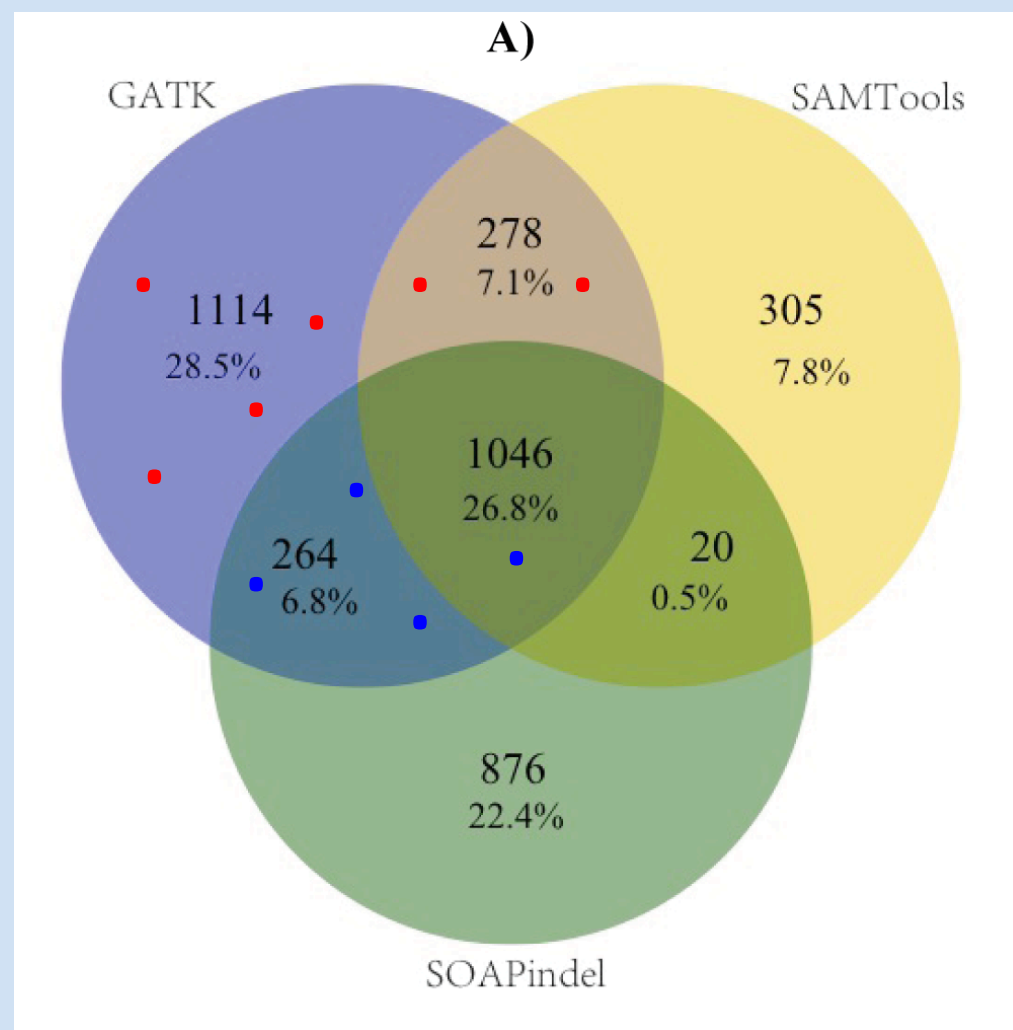
B) Mean # of known SNVs (present in dbSNP135) found by 5 pipelines across 15 exomes. The percentage in the center of the the Venn diagram is the percent of known SNVs called by all five pipelines.

Novel SNVs



C) Mean # of novel SNVs (not present in dbSNP135) found by 5 pipelines across 15 exomes. The percentage in the center of the Venn diagram is the percent of novel SNVs called by all five pipelines.

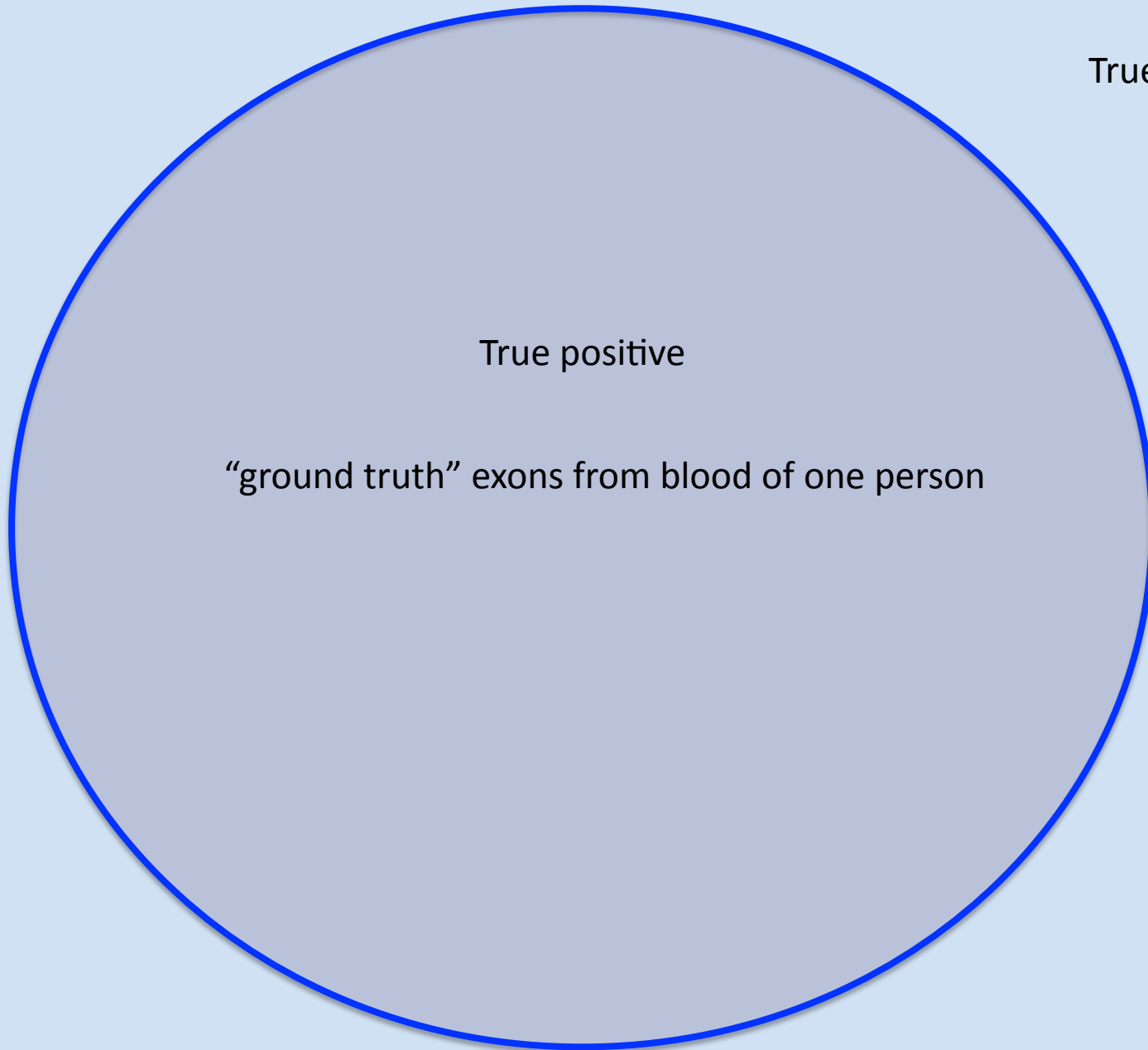
Indels called by GATK, SOAP and SAMtools

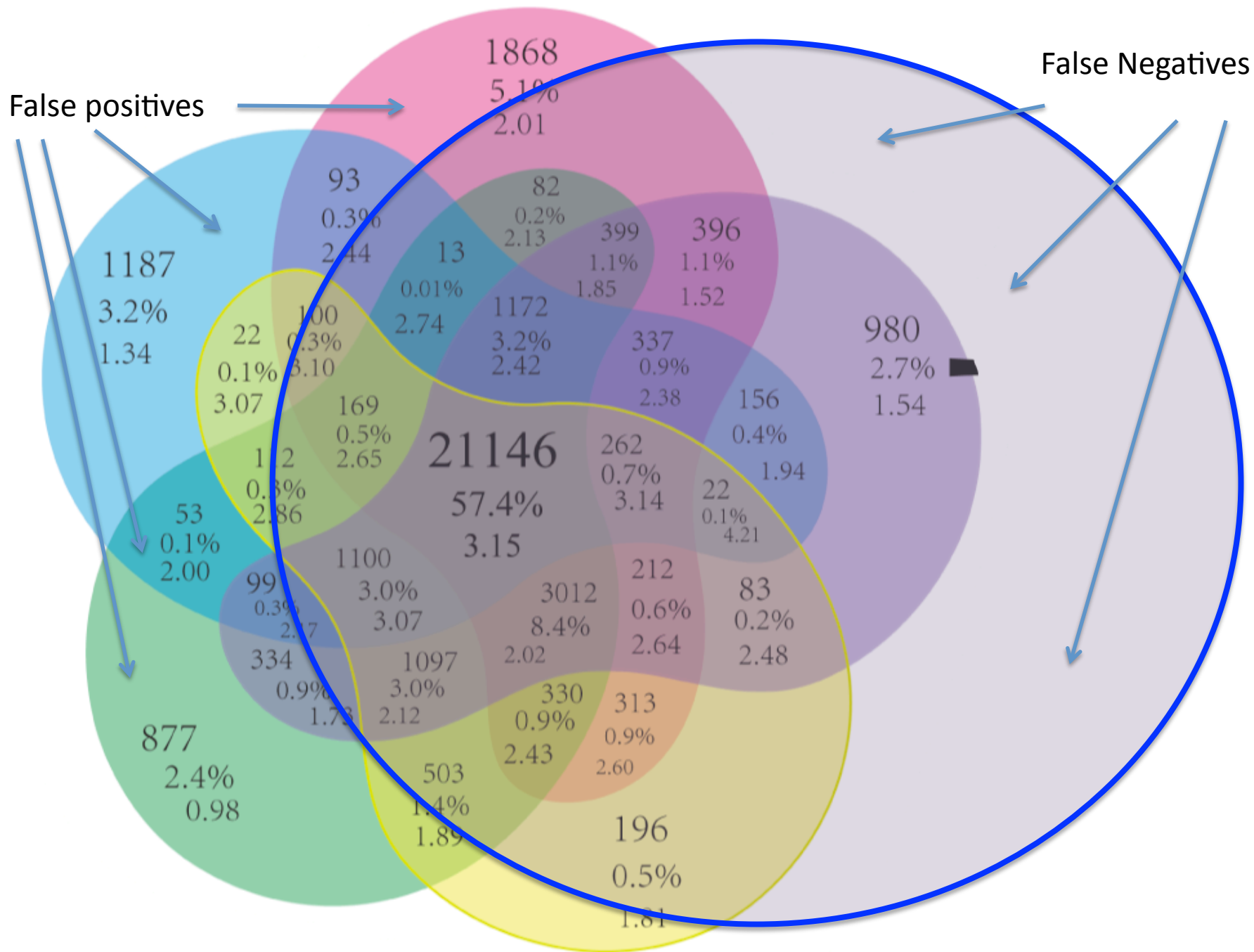


True negative

True positive

“ground truth” exons from blood of one person

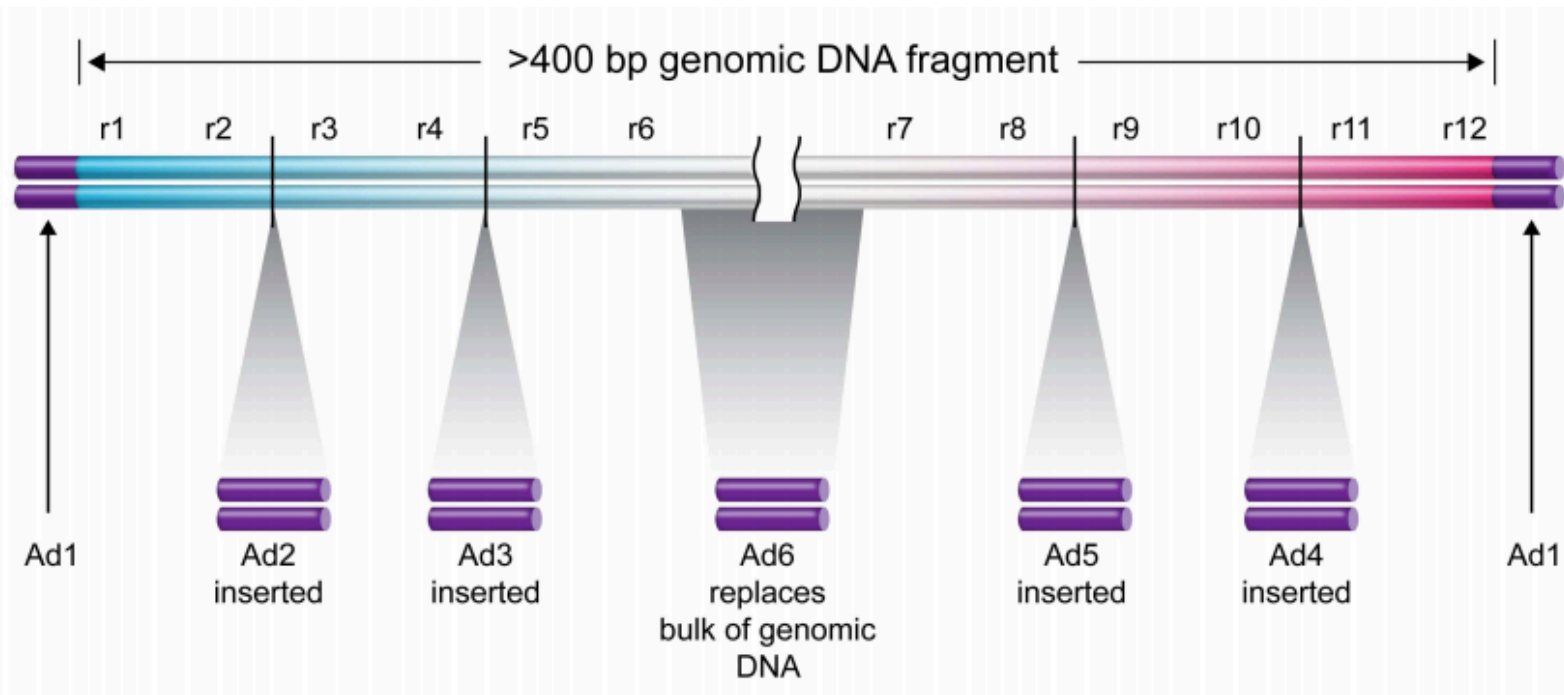
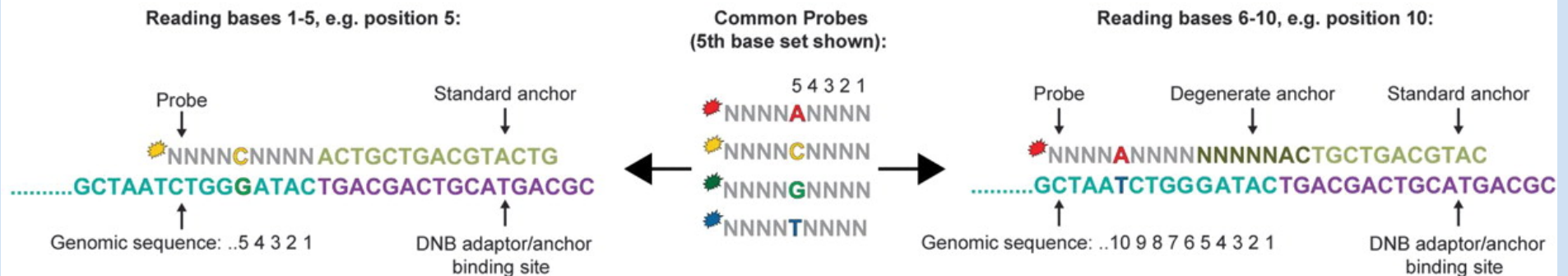




Cross validation using orthogonal
sequencing technology
(Complete Genomics)

Complete Genomics chemistry - combinatorial probe anchor ligation (cPAL)

D



Accuracy of Complete Genomics Whole Human Genome Sequencing Data

Analysis Pipeline v2.0

	FALSE POSITIVES	EST FPs	FALSE NEGATIVES	TOTAL DISCORDANCES	CONCORDANCE
Discordant SNVs per called MB	1.56×10^{-6}	4,450	1.67×10^{-6}	3.23×10^{-6}	99.9997% of bases

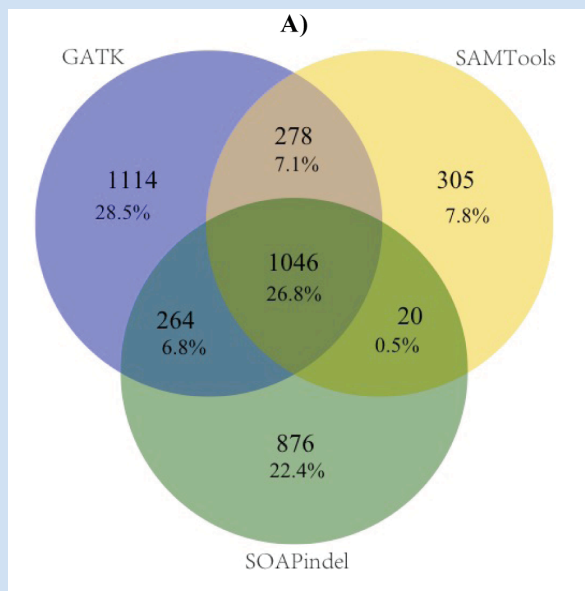
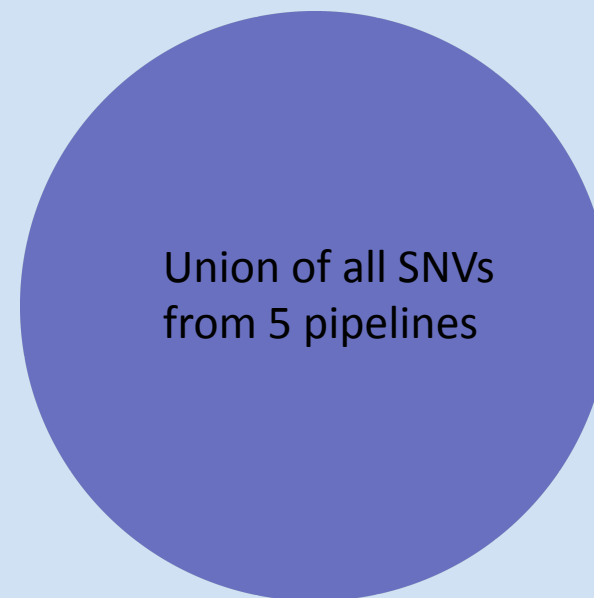
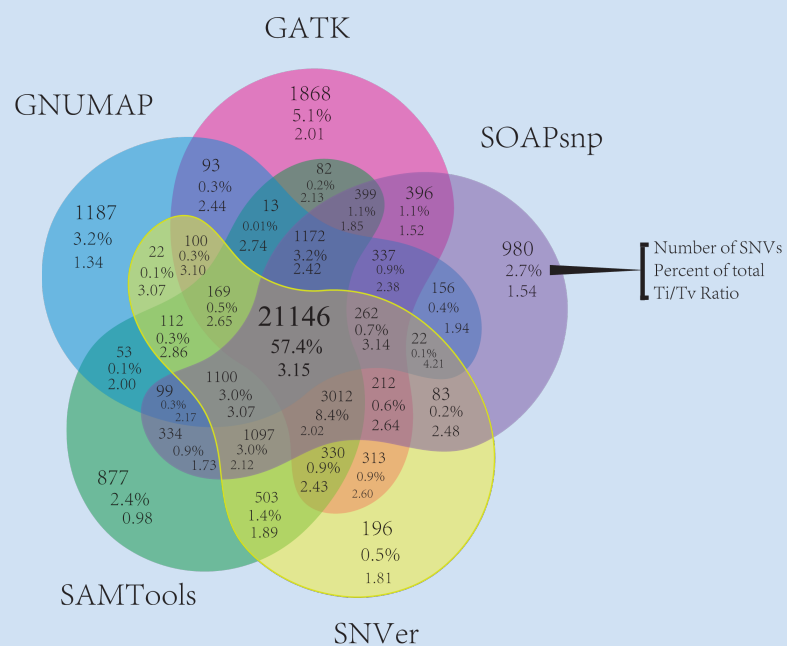
Table 2. *Concordance of Technical Replicates.*

COMPLETE GENOMICS CALL	OTHER PLATFORM	PLATFORM-SPECIFIC SNVs	VALIDATION RATE	EST FPs	FPR
Het or Hom SNV	No SNV Reported	99K	$17/18 = 94.4\%$	5,577	0.16%
No-call or Hom-Ref	SNV Reported	345K	$2/15 = 13.3\%$	299,115	8.2%

Table 3. *False Positive Rate.*

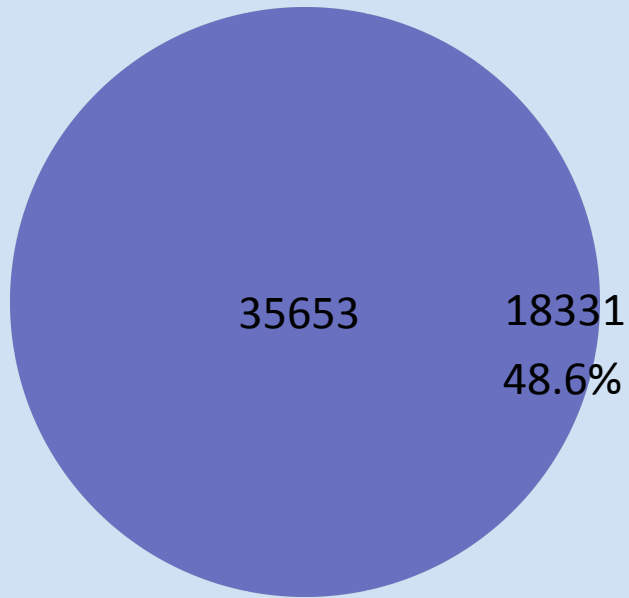
Performance comparison of whole-genome sequencing platforms

Hugo Y K Lam^{1,8}, Michael J Clark¹, Rui Chen¹, Rong Chen^{2,8}, Georges Natsoulis³, Maeve O'Huallachain¹, Frederick E Dewey⁴, Lukas Habegger⁵, Euan A Ashley⁴, Mark B Gerstein⁵⁻⁷, Atul J Butte², Hanlee P Ji³ & Michael Snyder¹

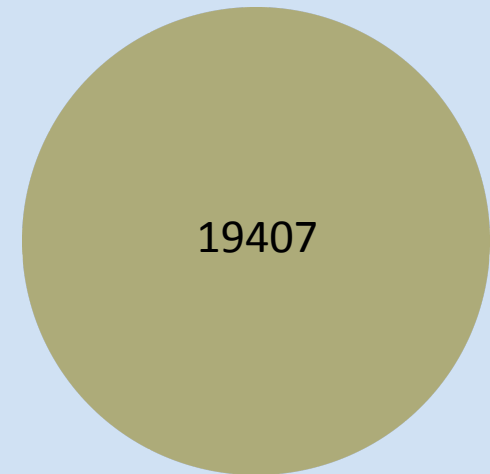


What is the "True" Personal Genome?

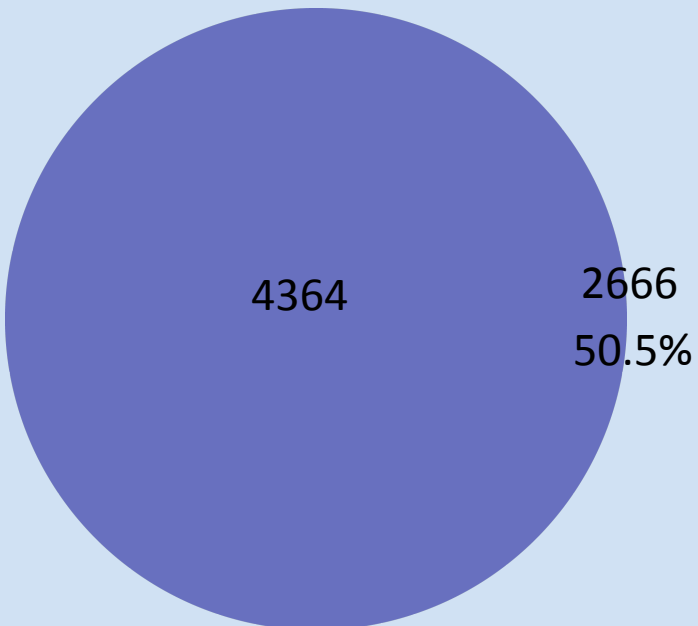
Illumina SNVs



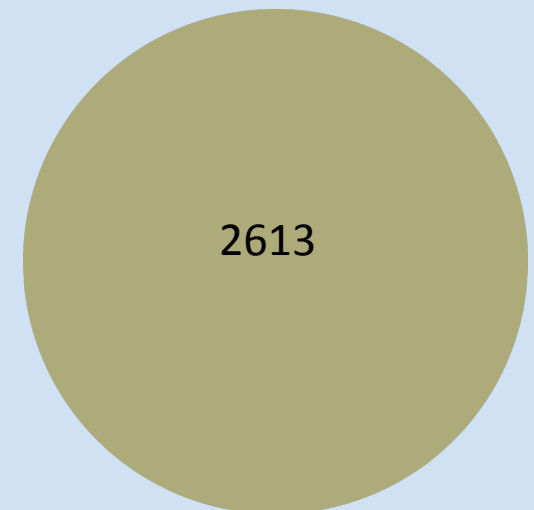
CG SNVs



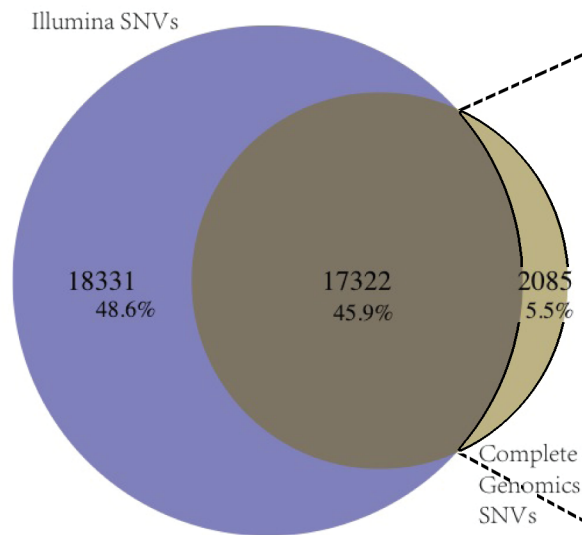
Illumina indels



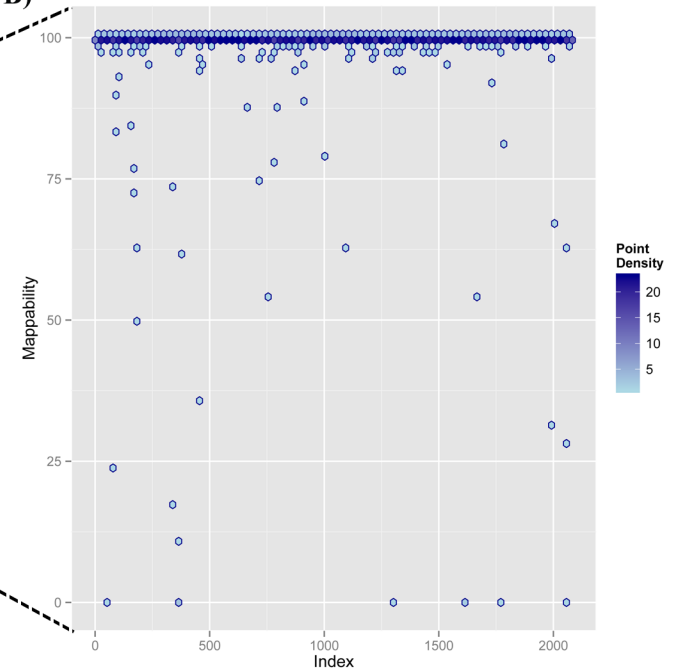
CG Indels



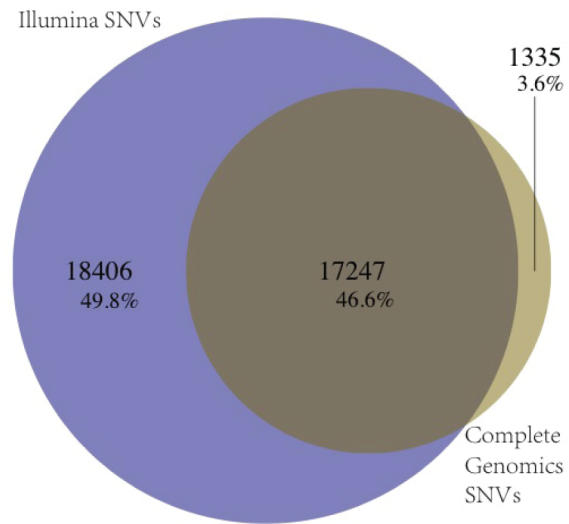
A.)



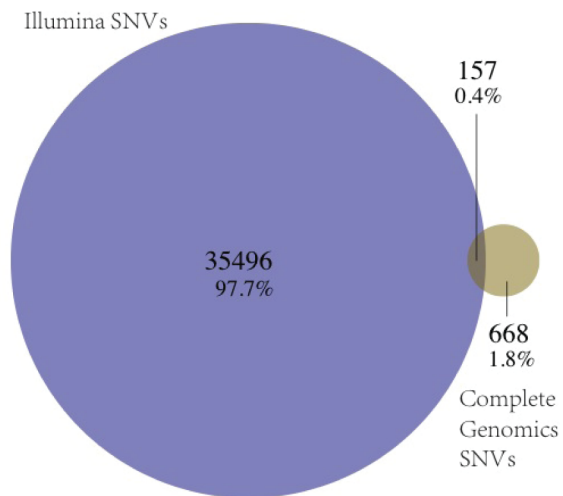
B)



C)



D.)



Genomic Dark Matter: The reliability of short read mapping illustrated by the Genome Mappability Score

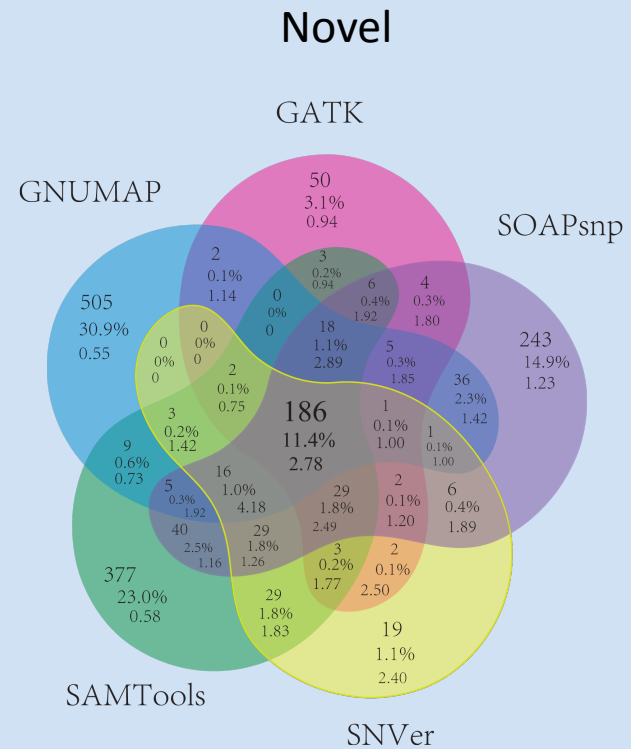
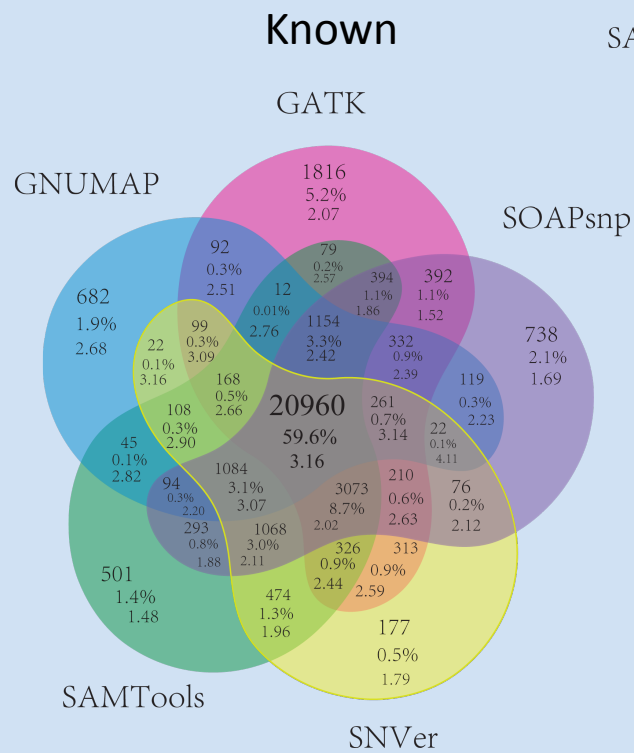
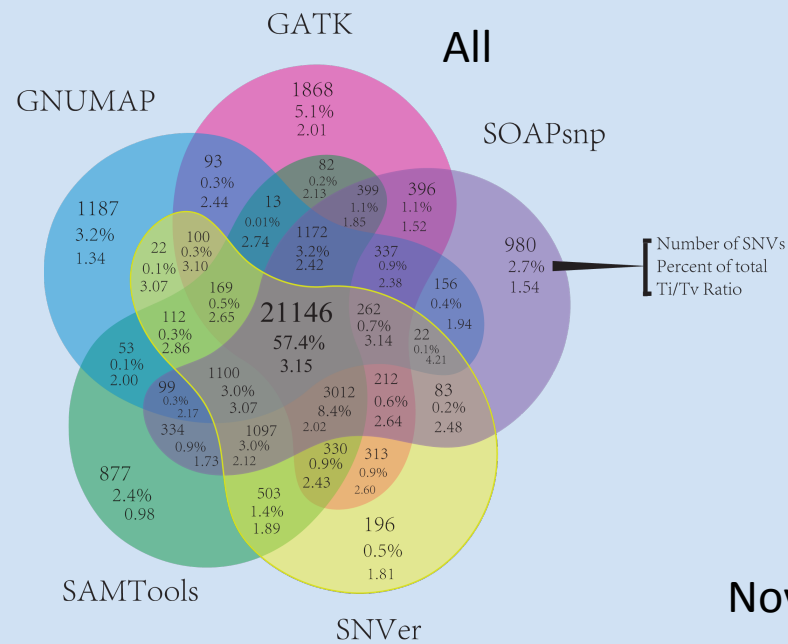
Hayan Lee^{1,2*} and Michael C. Schatz^{1,2}

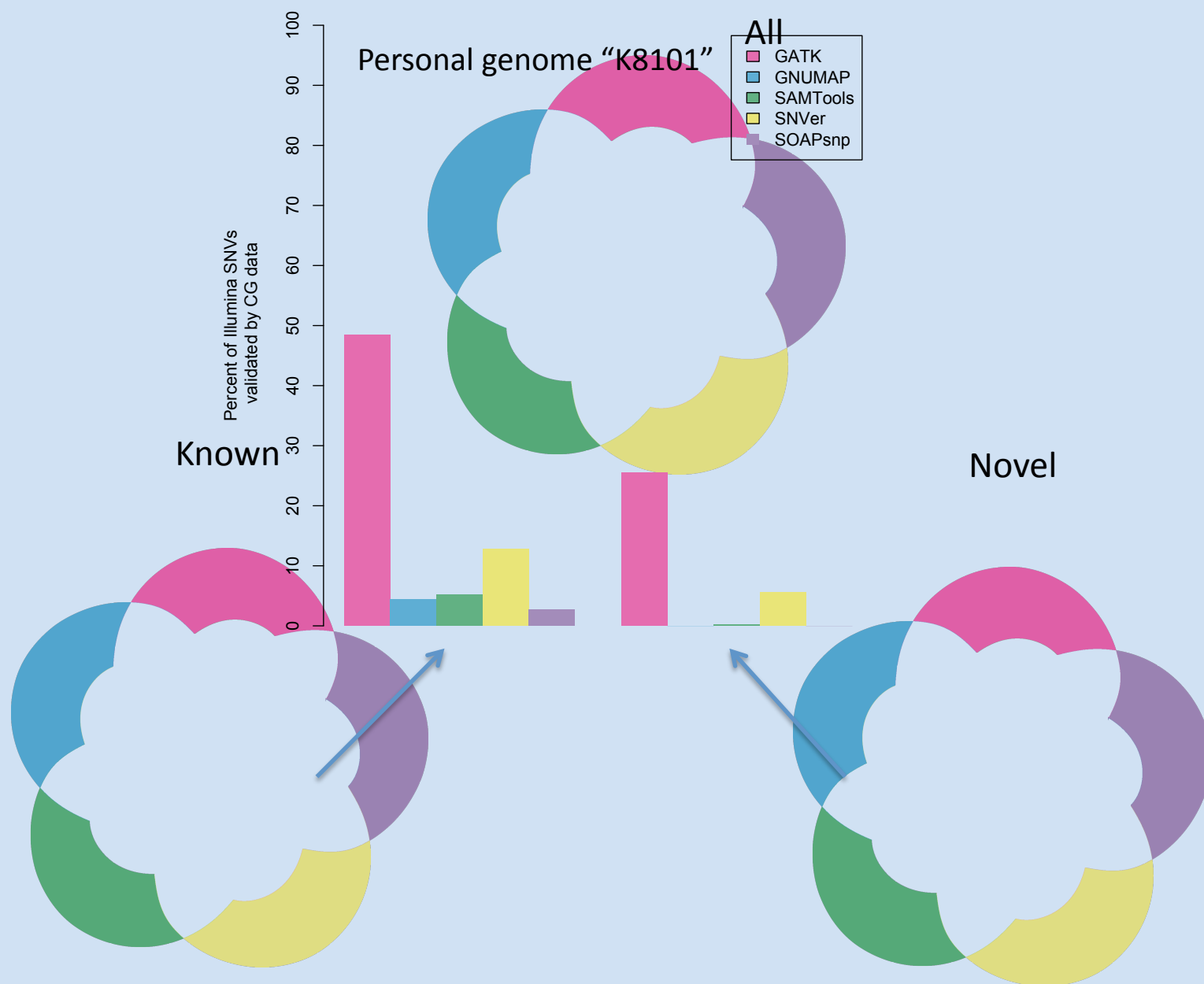
¹Department of Computer Science, Stony Brook University, Stony Brook, NY

²Simons Center for Quantitative Biology, Cold Spring Harbor Laboratory, Cold Spring Harbor, NY

Bioinformatics Advance Access published June 4, 2012

- Genome Mappability Score (GMS) -- measure of the complexity of resequencing a genome = a weighted probability that any read could be unambiguously mapped to a given position, and thus measures the overall composition of the genome itself.

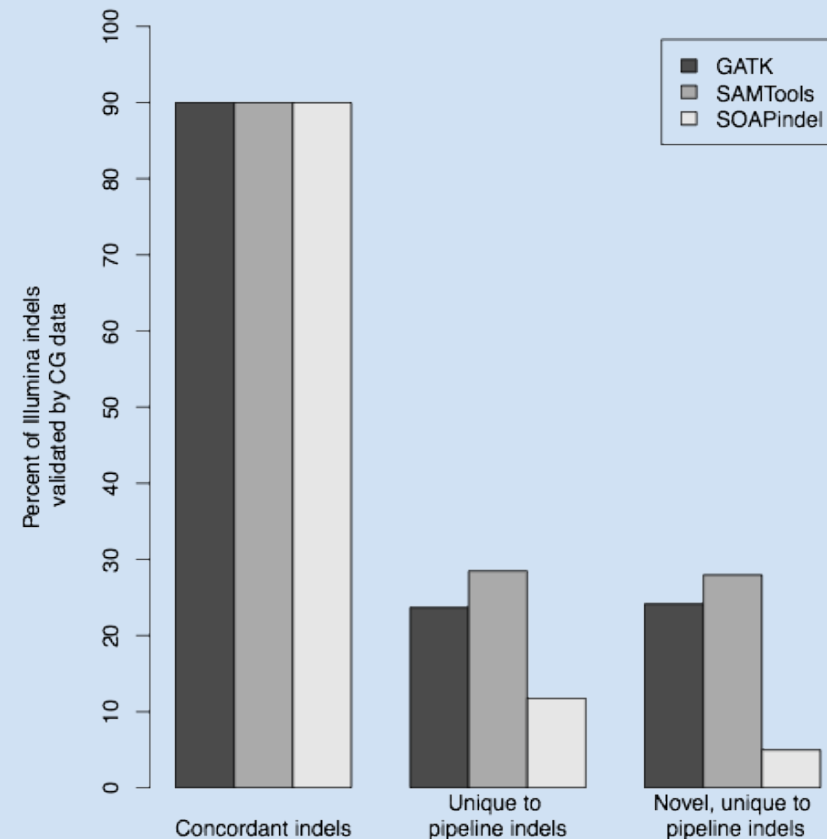
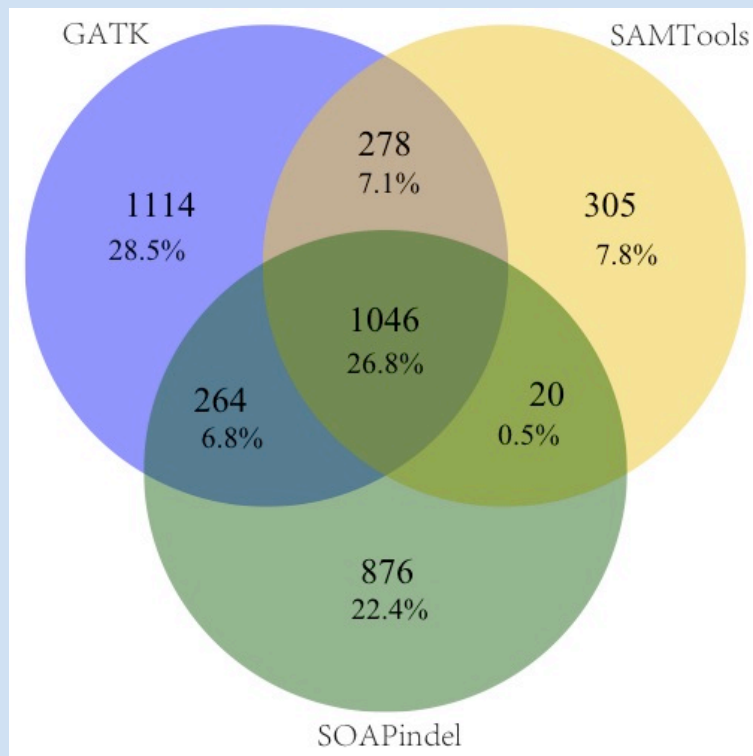




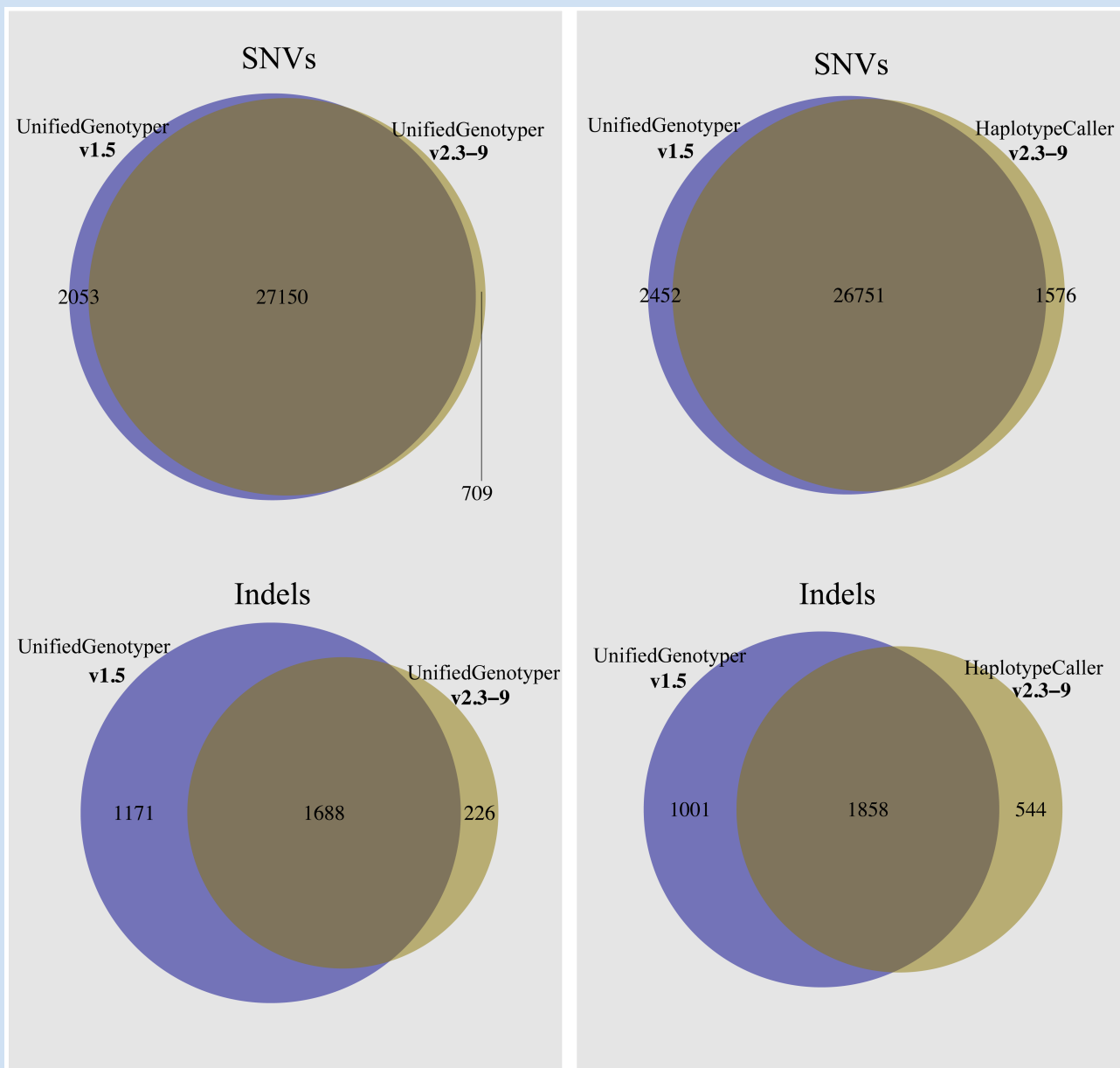
Higher Validation by CG of SNVs with the BWA-GATK(v1.5) pipeline

- Reveals higher validation rate of unique-to-pipeline variants, as well as uniquely discovered novel variants, for the variants called by BWA-GATK(v1.5), in comparison to the other 4 pipelines (including SOAP).

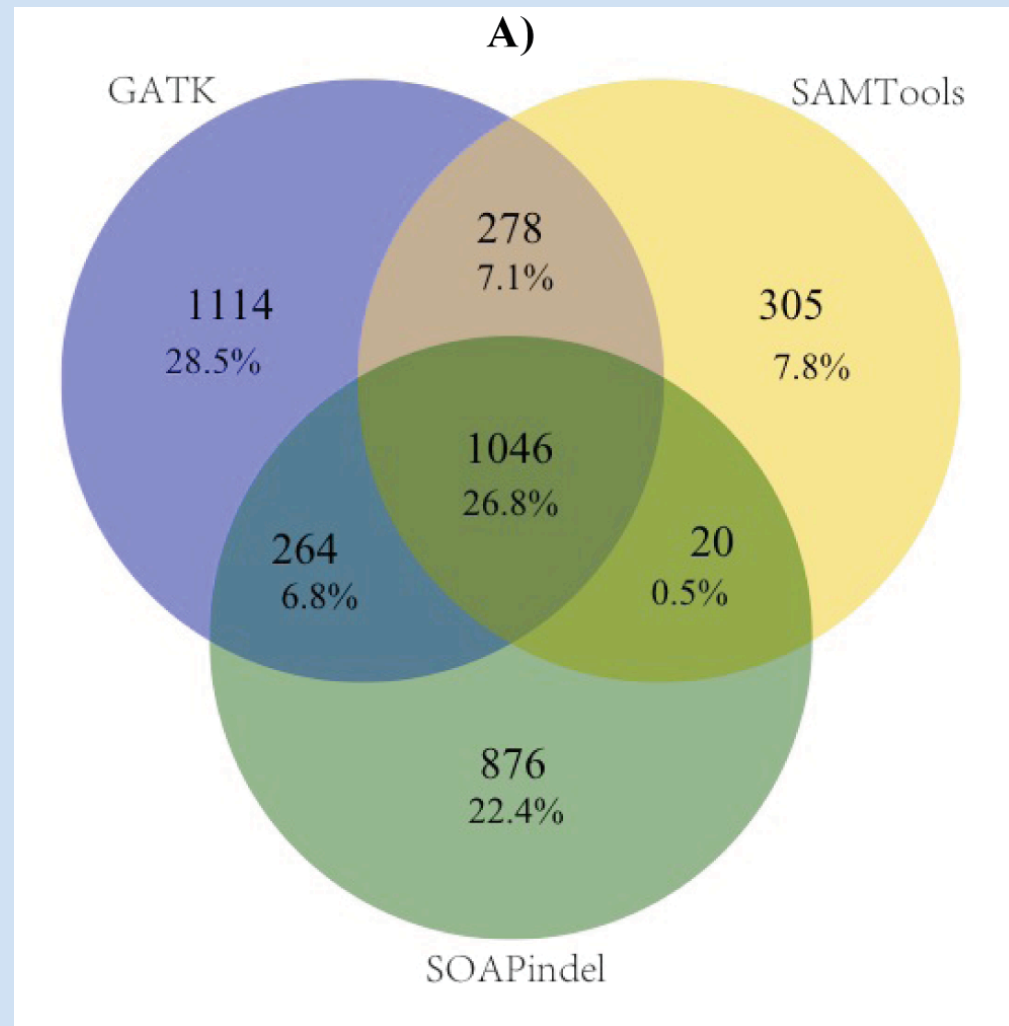
Validating Indels with Complete Genomics Data for the 3 pipelines



Comparing to New Versions of GATK



Validation of SNVs and Indels called by GATK, SOAP and both, with another platform

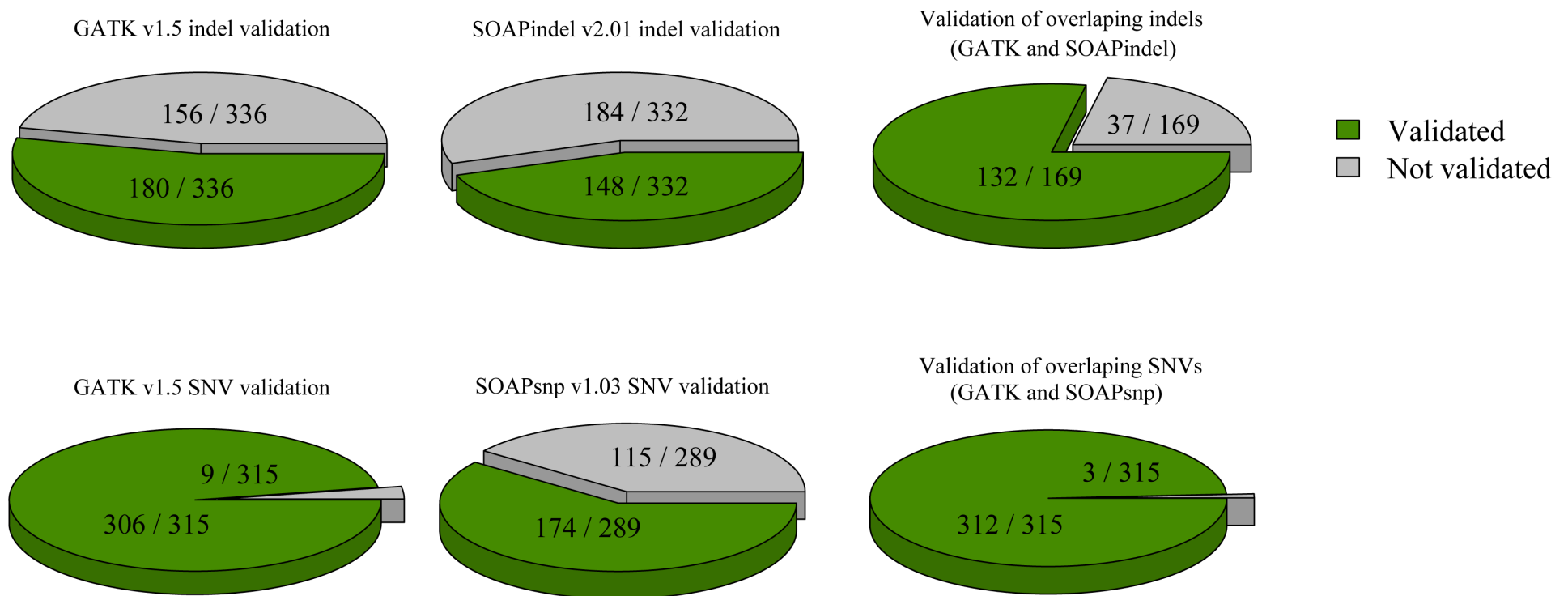


Indels

Validation with PCR amplicons and MiSeq 150 bp reads at ~5000x coverage

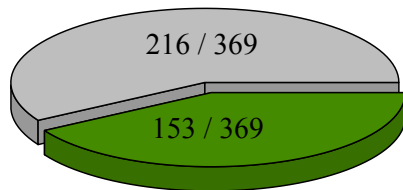
1,140 SNVs, with random sampling of 380 from the set of unique-to-GATK SNVs, 380 from the set of unique-to-SOAPsnp SNVs, and 380 from the set that were overlapping between these two pipelines.

960 indels, with random sampling of 386 from the unique-to-GATK indel set, 387 from the unique-to-SOAPindel set, and 187 from the set of indels overlapping between the two (SOAPindel and GATK).

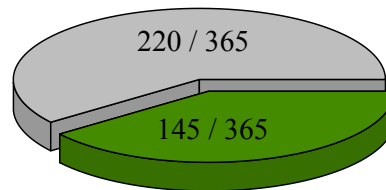


Validation of ~2000 PCR amplicons with PacBio reads from two SMRT cells (~50,000 useable reads per cell)

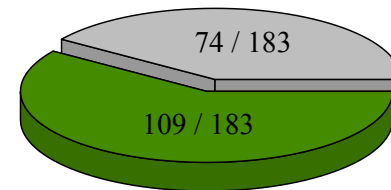
GATK v1.5 indel validation



SOAPindel v2.01 indel validation

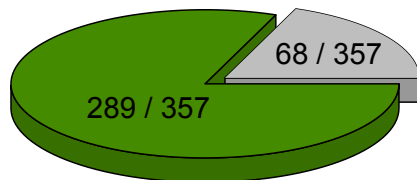


Validation of overlapping indels
(GATK and SOAPindel)

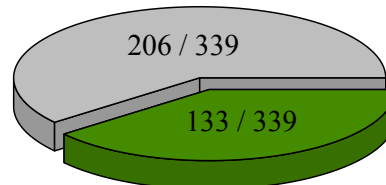


■ Validated
■ Not validated

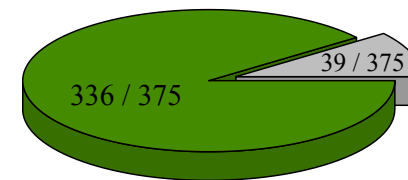
GATK v1.5 SNV validation



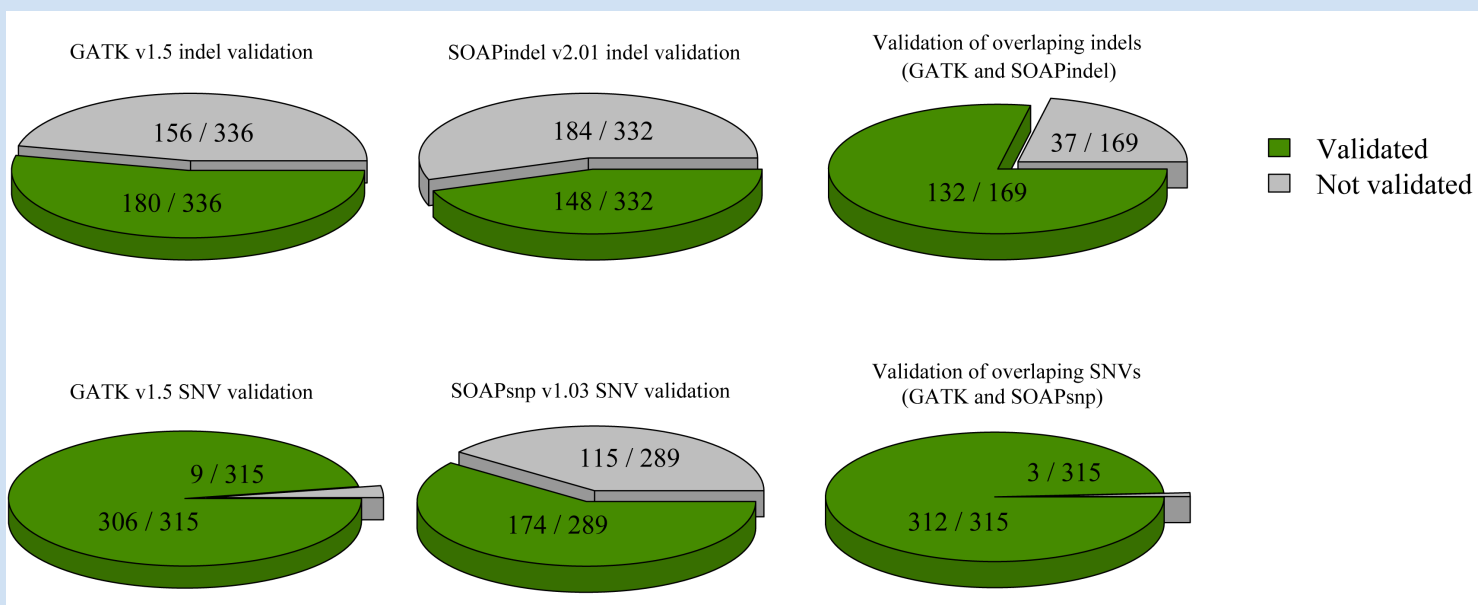
SOAPSnp v1.03 SNV validation



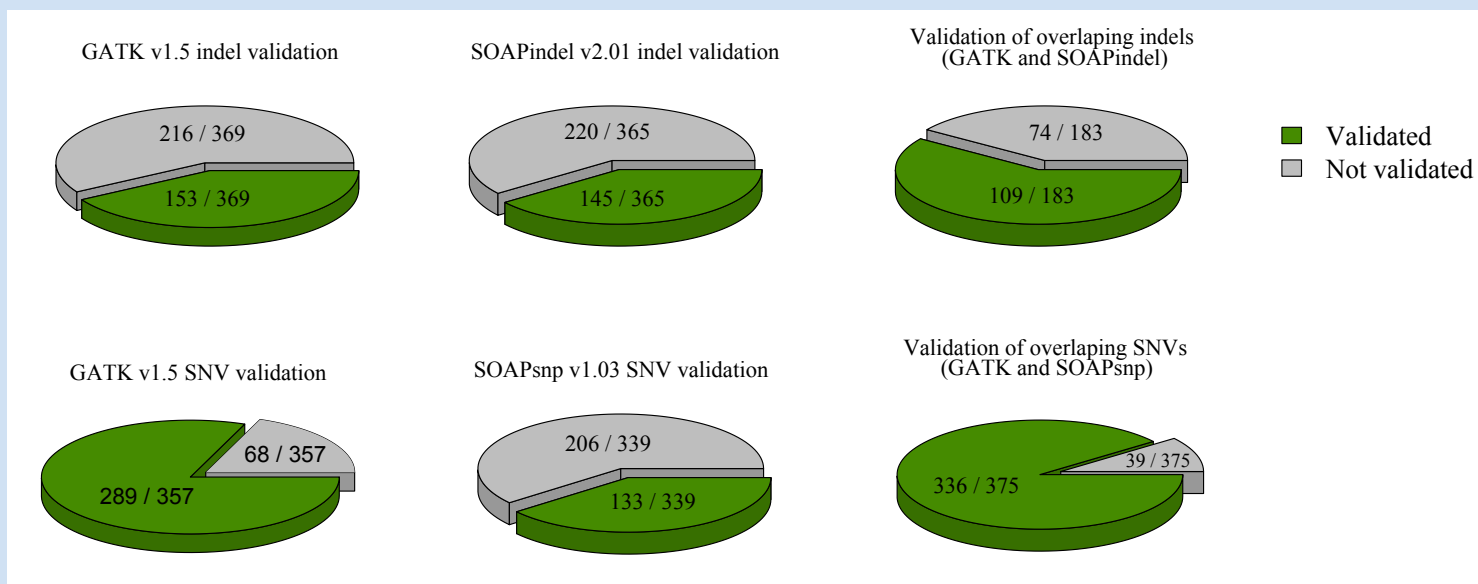
Validation of overlapping SNVs
(GATK and SOAPSnp)



MiSeq

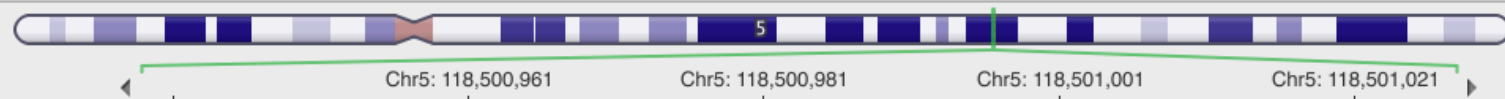


PacBio



Chr5: 118,500,938 - 118,501,026

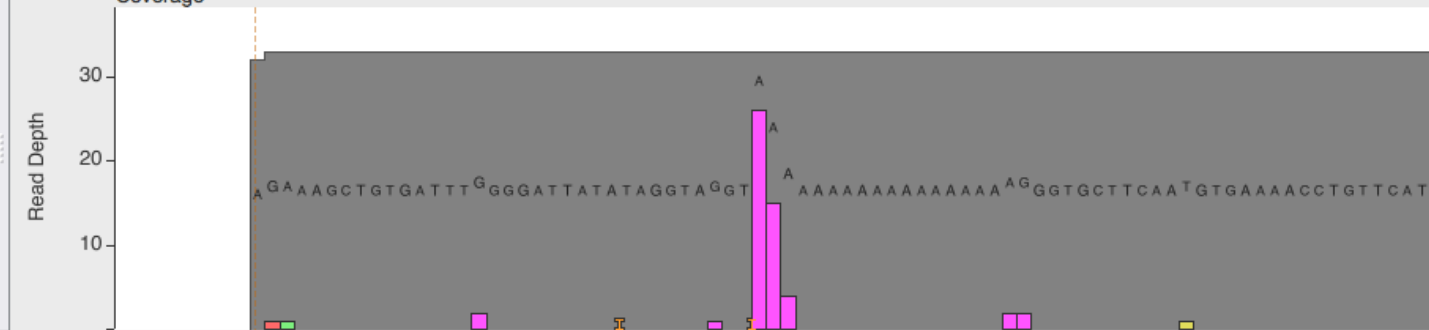
Homo sapiens (Human), GRCh37 g1k (Feb 2009)



picardbam (Homo sapiens, GRCh_37)

Coverage

bams



reads.150_trunc.sort (Homo sapiens, GRCh_37)

Coverage

bams



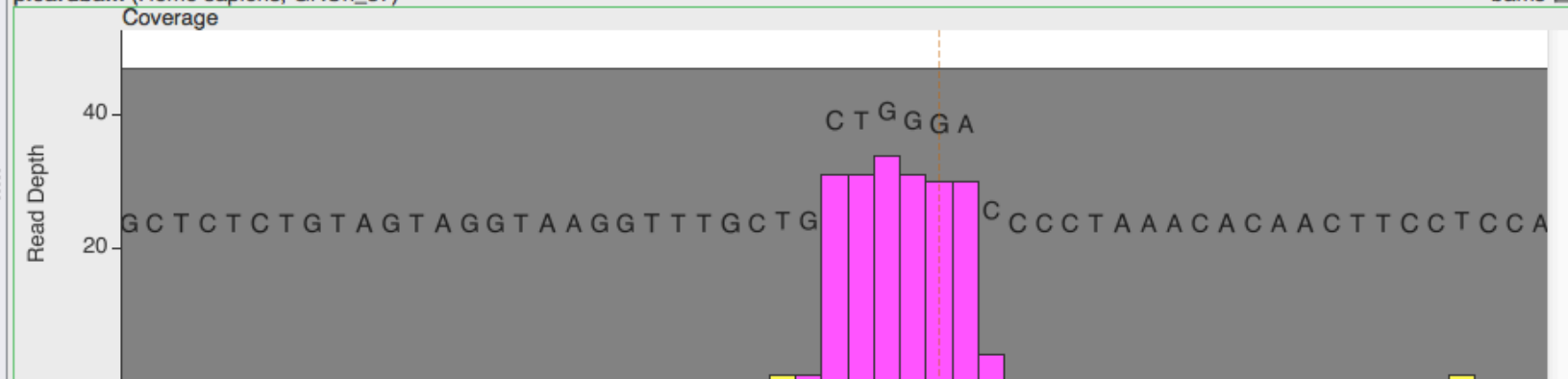
RefSeq Genes, UCSC (Homo sapiens, GRCh_37_g1k)

data.goldenhelix.com



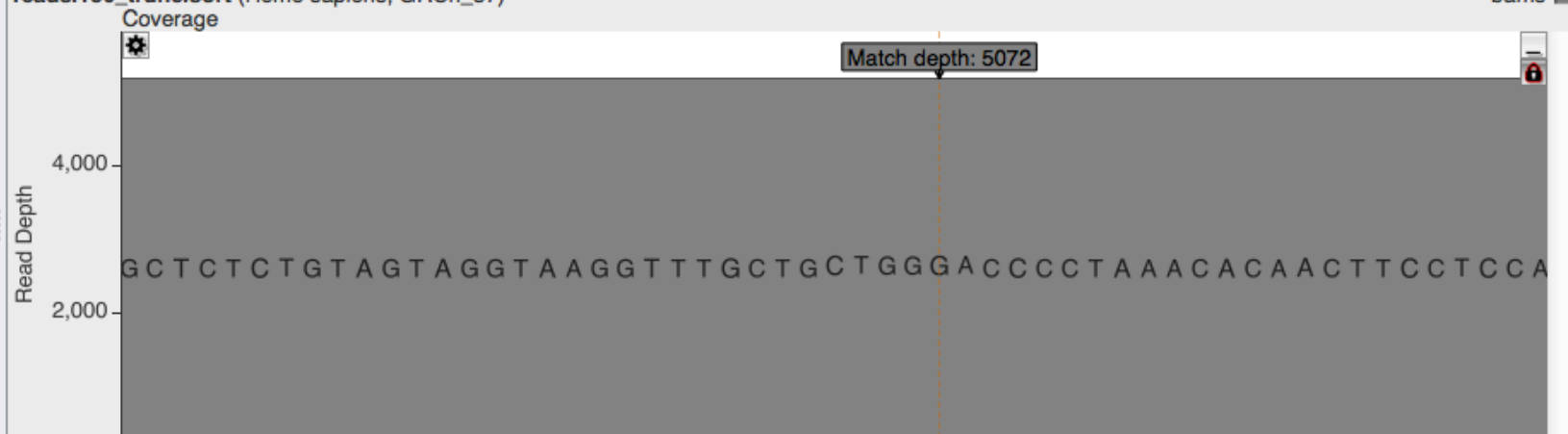
picardbam (Homo sapiens, GRCh_37)

bams

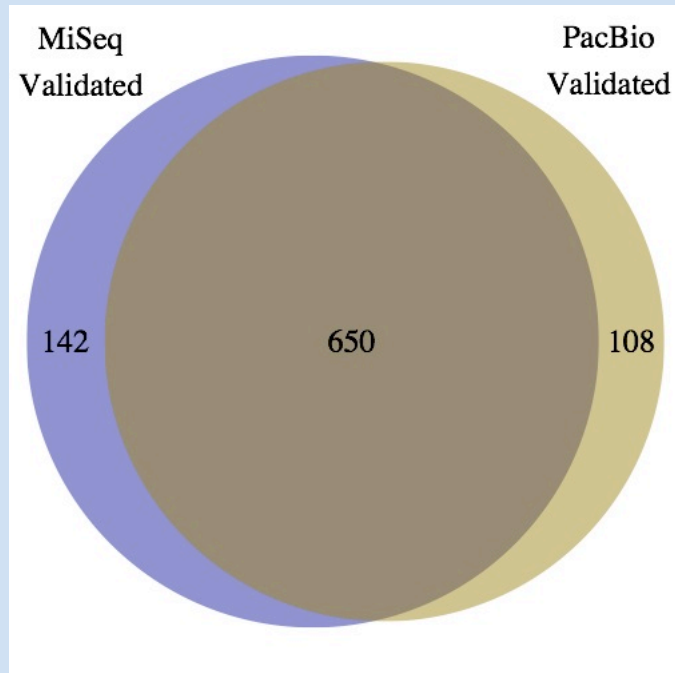
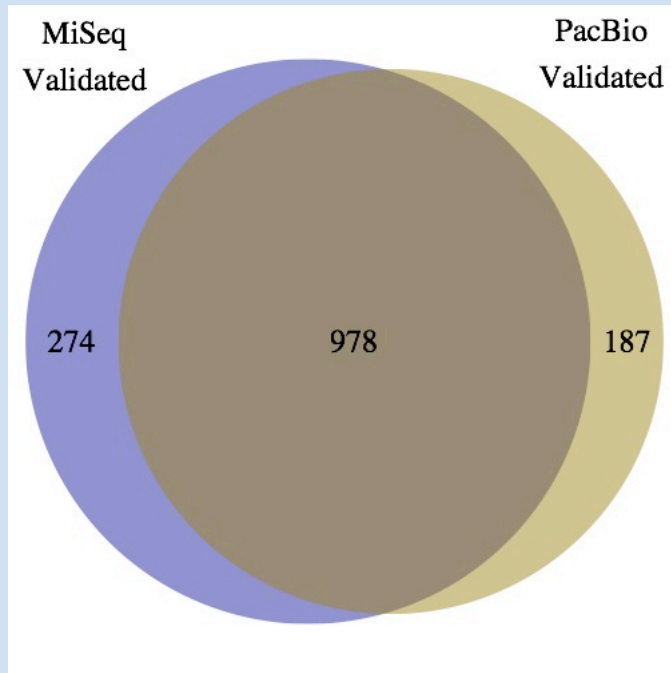


reads.150_trunc.sort (Homo sapiens, GRCh_37)

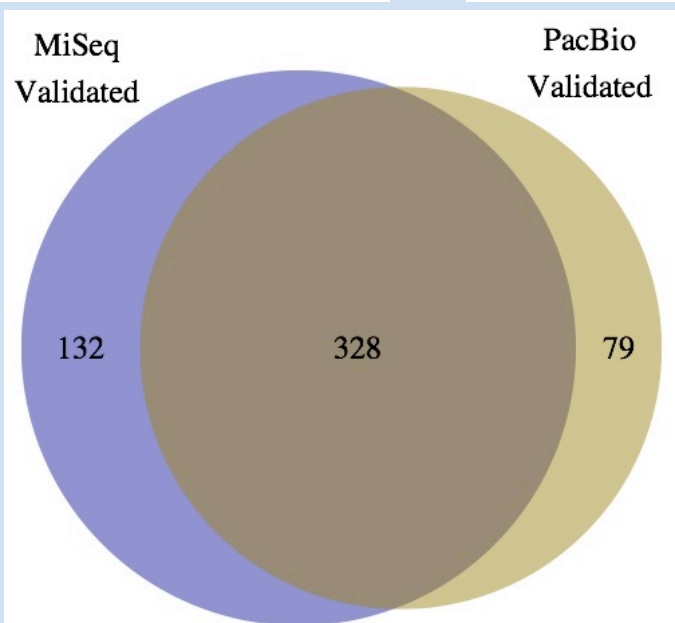
bams



SNVs
+ Indels



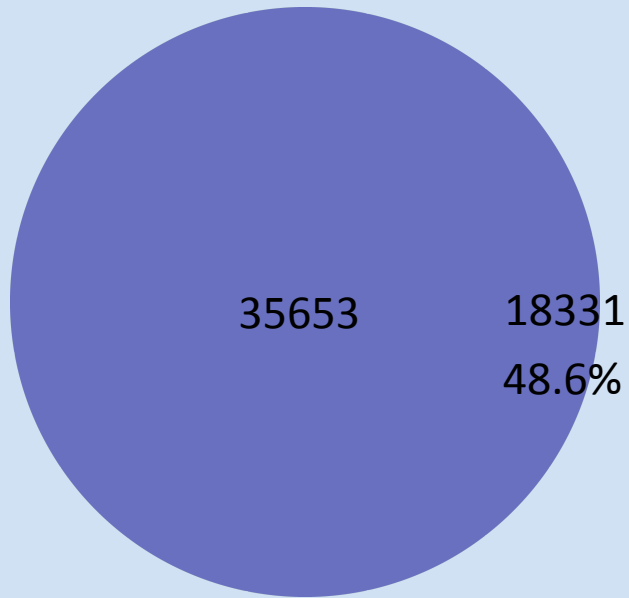
SNVs



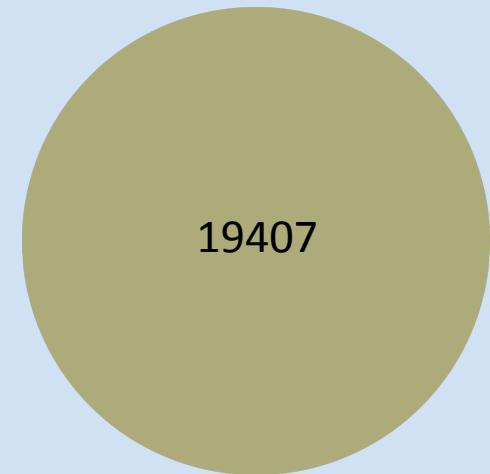
Indels

What is the "True" Personal Genome?

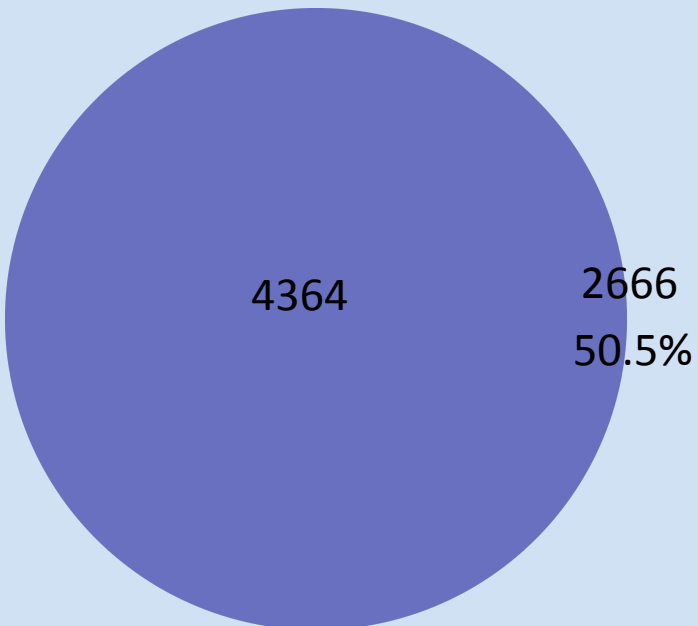
Illumina SNVs



CG SNVs



Illumina indels



CG Indels

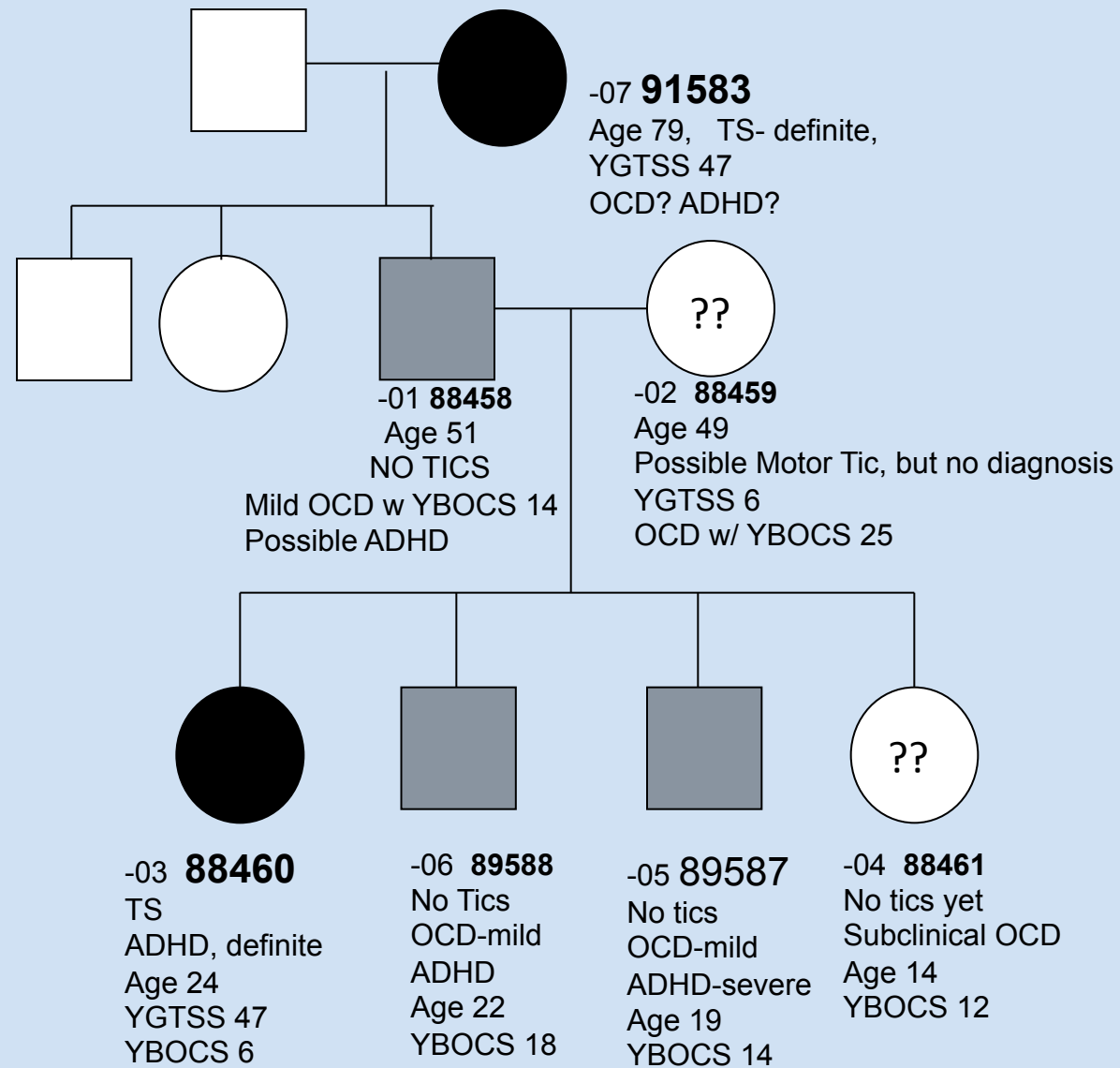


Optimizing the Variant Calling Pipeline Using Family Relationships

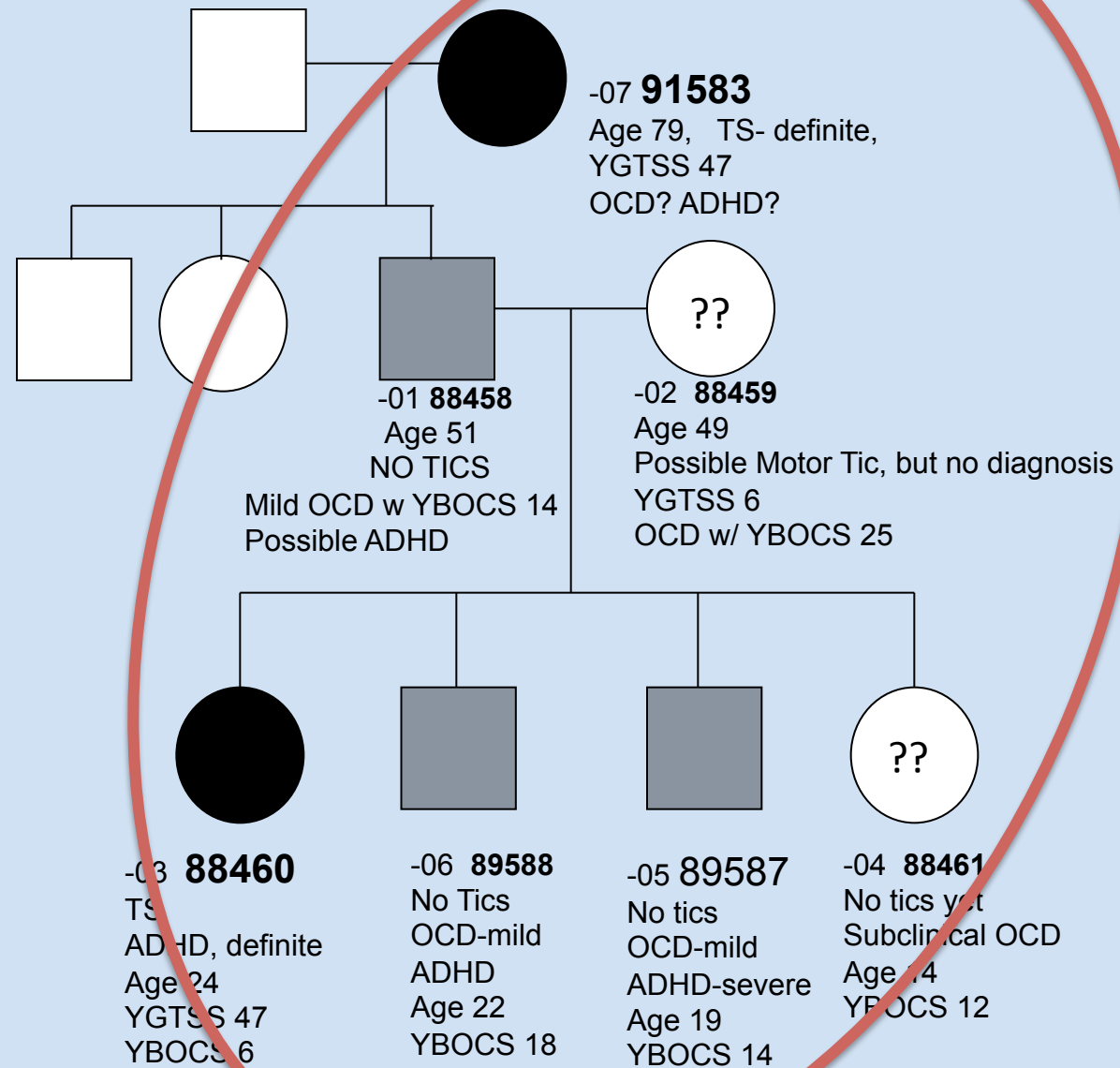
We looked for SNVs that were detected in children but not in parents using 3 different strategies:

1. We used all of the SNVs that were detected by all 5 pipelines for both parents and children
2. We used all of the detected SNVs for parents, but only the concordant SNVs between the 5 different pipelines for children.
3. We used SNVs concordant between the 5 different pipelines for children and parents.

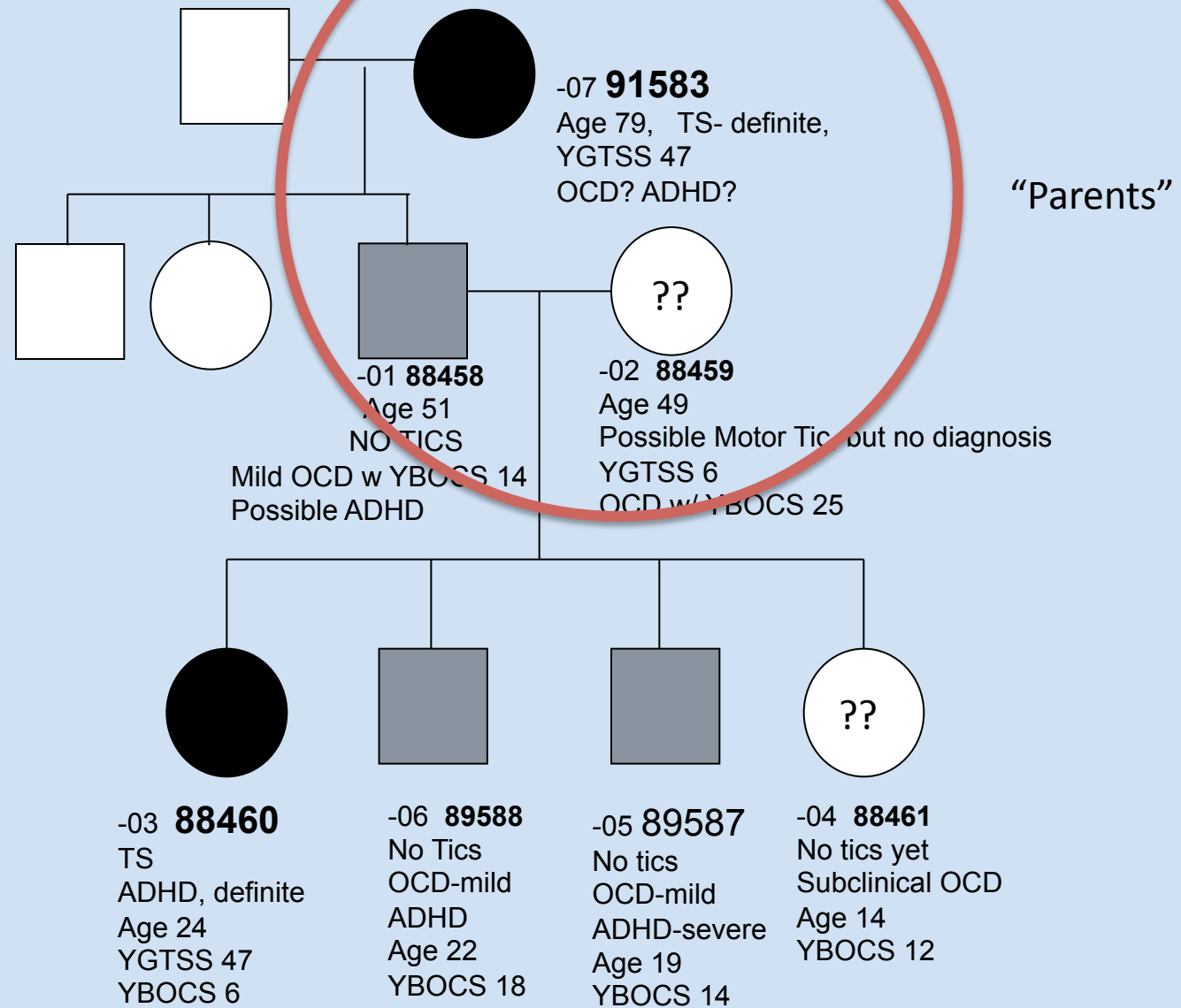
TDT- 09 -1018
K26679



TDT- 09 -1018
K26679



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K26679



Analysis based on various pipelines

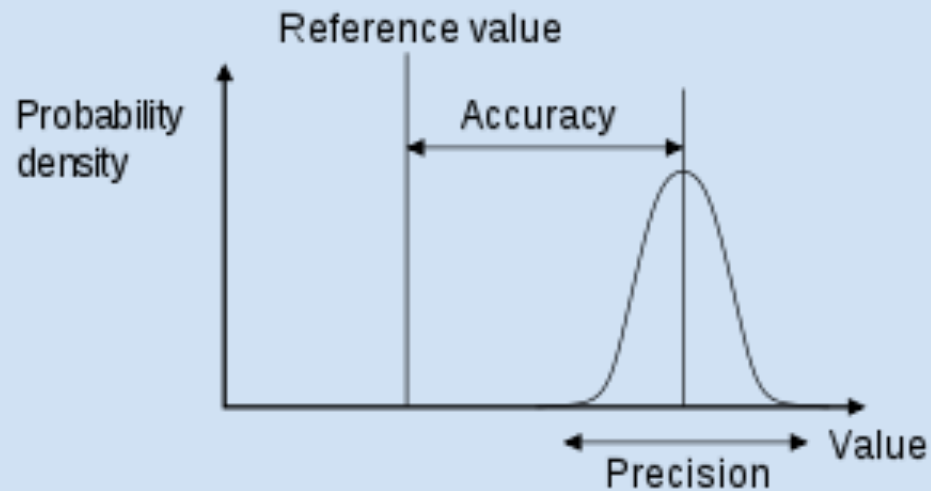
- “Parents” in this case means the mother, father AND grandmother.
- Taking the **Union** of SNVs from all 5 pipelines from “Parents”, and subtract that from the **Union** of all SNVs in each child.
- Or Subtract the **Union** of these “Parents” from the SNVs in the child **concordant** between 5 pipelines.
- Or, subtract the **concordant** variants from 5 pipelines in “Parents” from the **concordant** variants for 5 pipelines in each child.

Table 3. *De novo* single-nucleotide variants (SNVs) were detected in two families contained within the 15 study exomes.

	Number of putative <i>de novo</i> coding non-synonymous or nonsense SNVs detected	
	Without using the grandparents as a filter	Using the grandparents as a filter
Family 1		
Child A	241	1
Child B	211	0
Child C	102	6
Child D	242	3
Family 2		
Child A	49	NA ^a
Child B	41	NA ^a

^aN/A, no grandparent available.

Family 1 had a grandparent available for filtering purposes, whereas family 2 did not. To minimize false positives in the pool of SNVs associated with each child, only highly concordant SNVs were used (SNVs detected by all five pipelines). To construct a comprehensive set of SNVs for each parent, and hence increase filtering accuracy, false negatives for parent SNVs were reduced by taking the union of all SNV calls from all five pipelines.



High accuracy, but low precision



High precision, but low accuracy

In the fields of [science](#), [engineering](#), [industry](#), and [statistics](#), the **accuracy** of a [measurement](#) system is the degree of closeness of measurements of a [quantity](#) to that quantity's actual (true) [value](#). The **precision** of a measurement system, also called [reproducibility](#) or [repeatability](#), is the degree to which repeated measurements under unchanged conditions show the same [results](#).

http://en.wikipedia.org/wiki/Accuracy_and_precision

Conclusions

- Sequencing a grandparent seems to help eliminate errors derived from the current depth of sequencing coverage in the mother and father.
- For now, we advocate using more than one pipeline on one set of sequencing data, but we expect the field to move toward >2 sequencing platforms per sample.
- Still need substantial work on indel-calling and validation.

Acknowledgements



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Jason O'Rawe



Yiyang Wu



Kai Wang

Michael Schatz
Giuseppe Narzisi
Eric Antoniou
Dick McCombie
Sequencing core facility

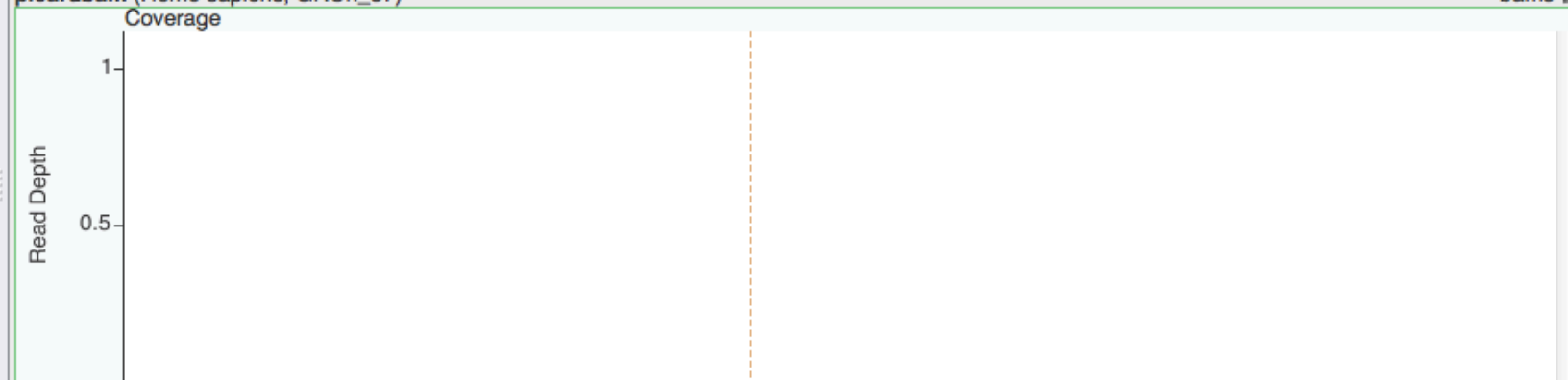


Tao Jiang
Guangqing Sun

EXTRA SLIDES – Not Shown

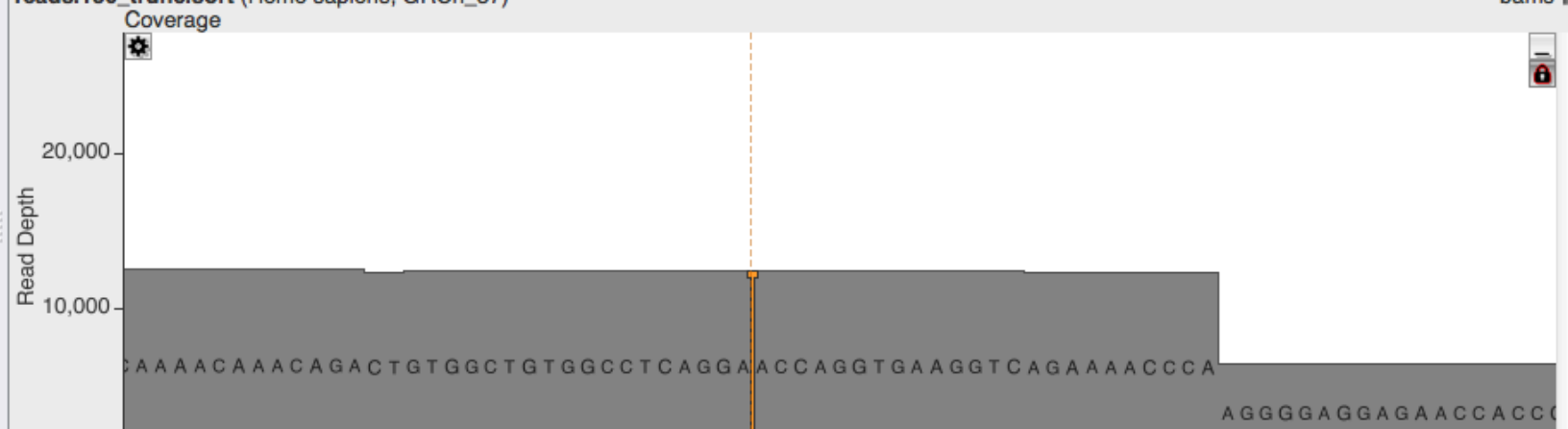
picardbam (Homo sapiens, GRCh_37)

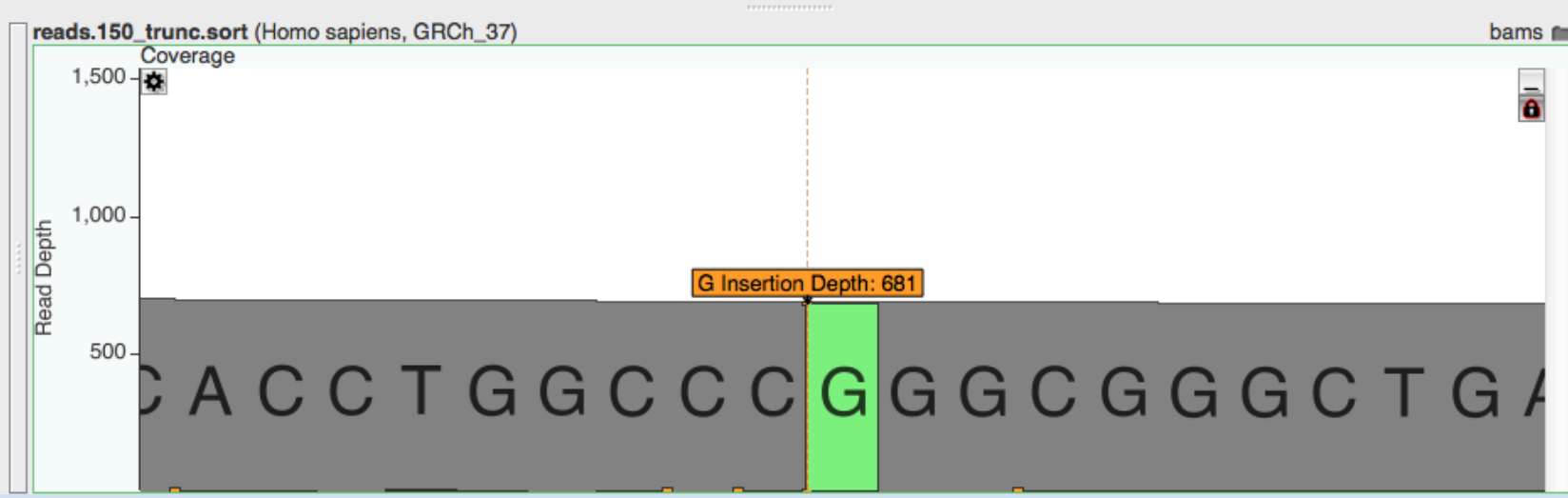
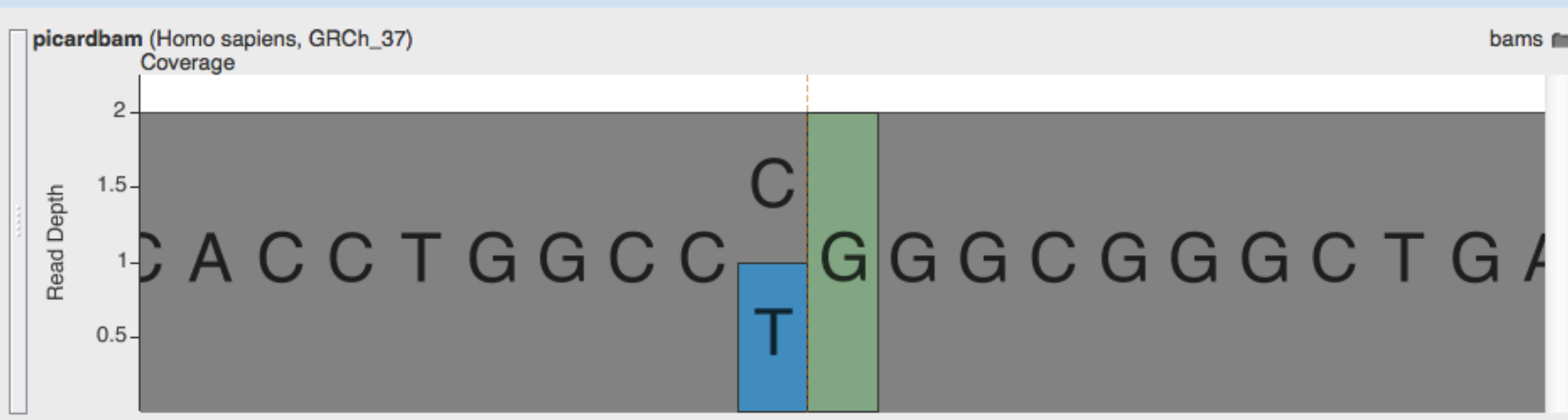
bams



reads.150_trunc.sort (Homo sapiens, GRCh_37)

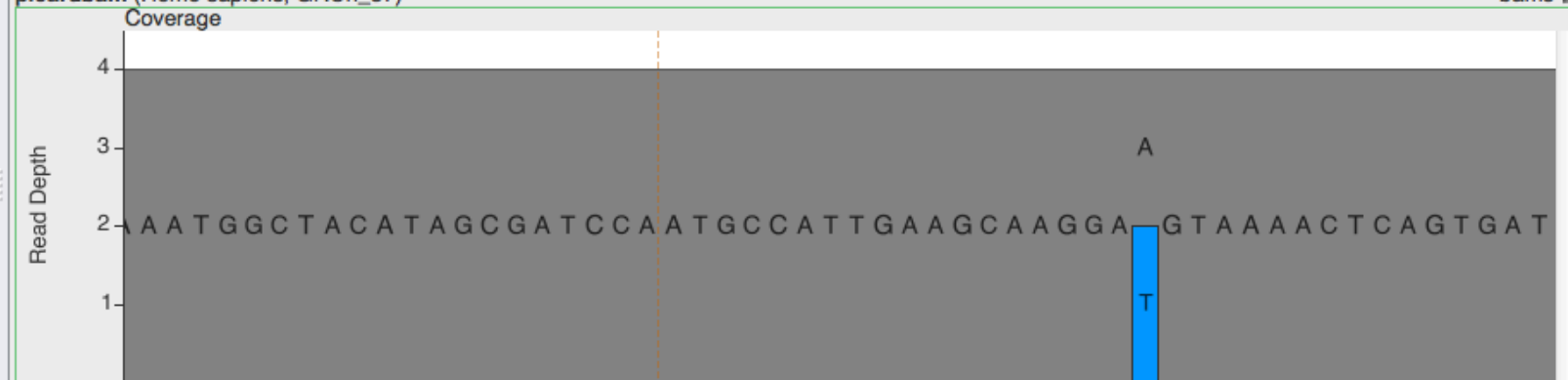
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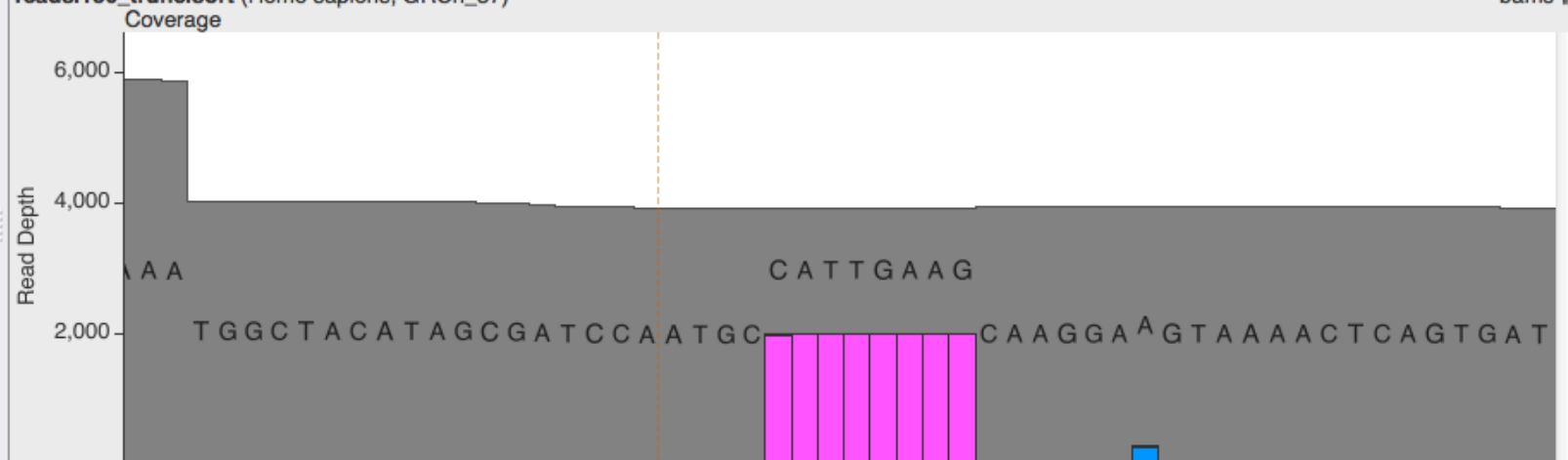
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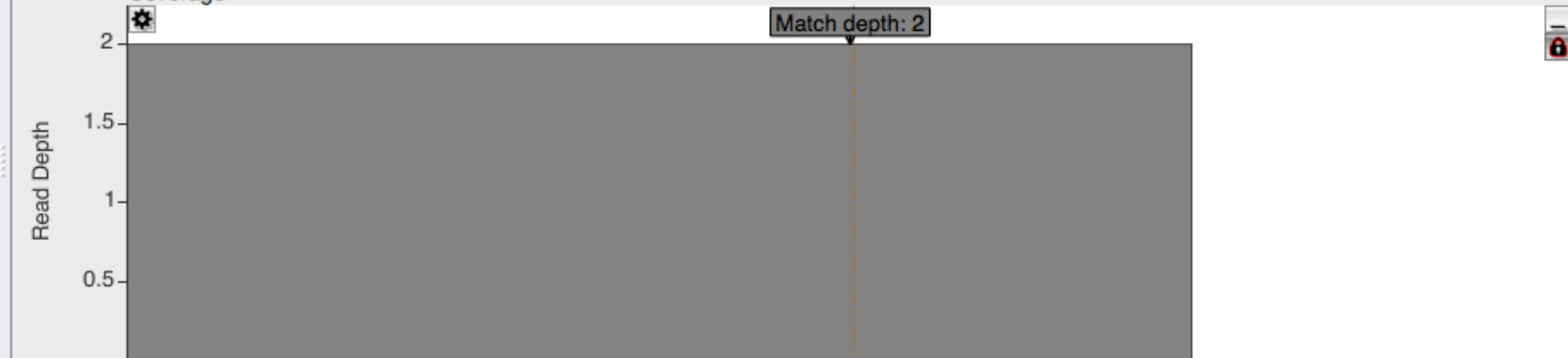
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picardbam (Homo sapiens, GRCh_37)

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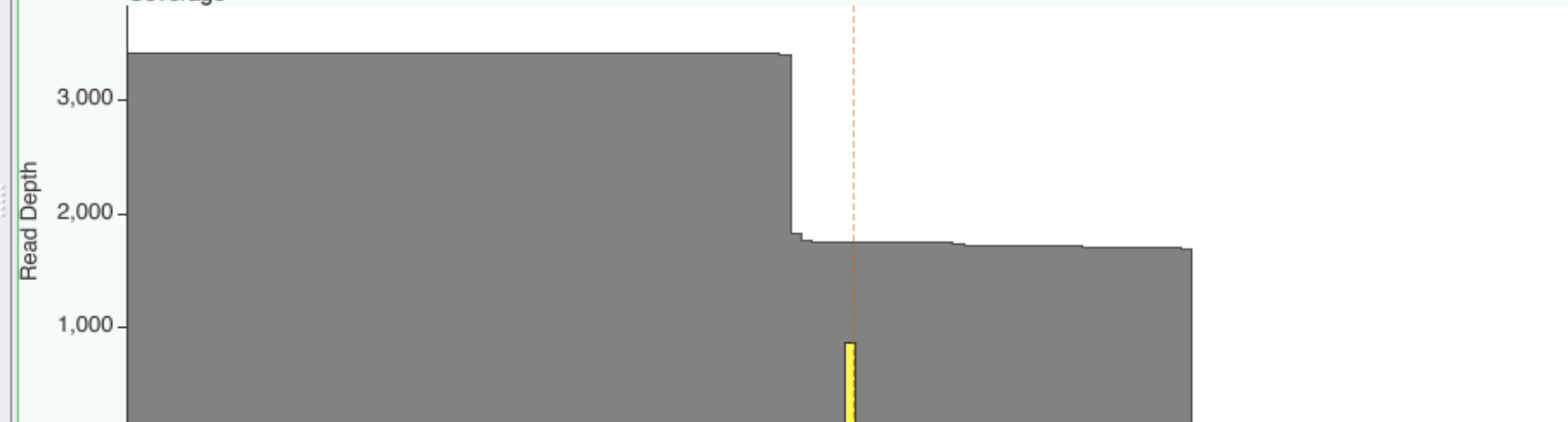
Coverage



reads.150_trunc.sort (Homo sapiens, GRCh_37)

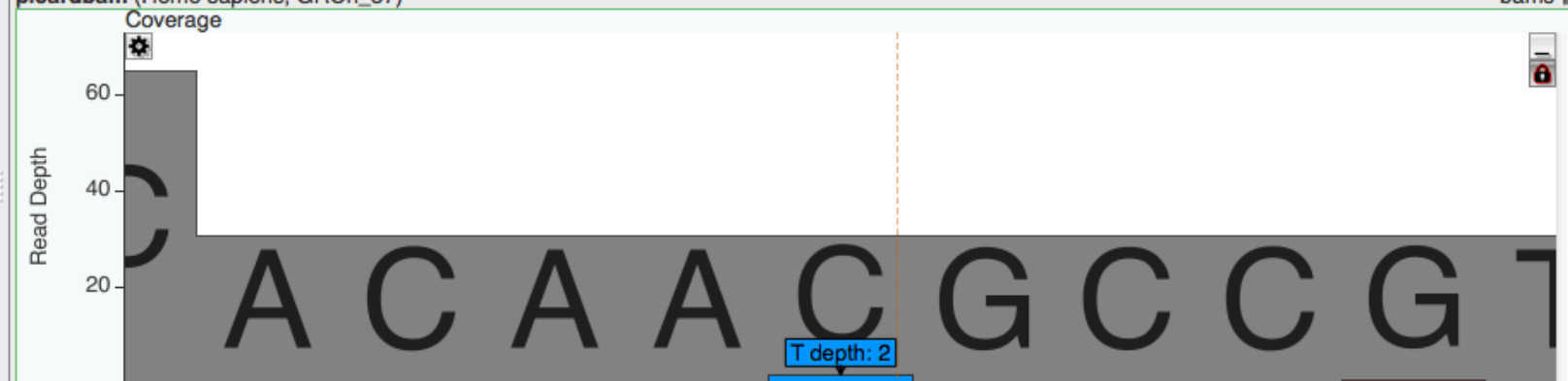
bams

Coverage



picardbam (Homo sapiens, GRCh_37)

bams



reads.150_trunc.sort (Homo sapiens, GRCh_37)

bams



picardbam (Homo sapiens, GRCh_37)

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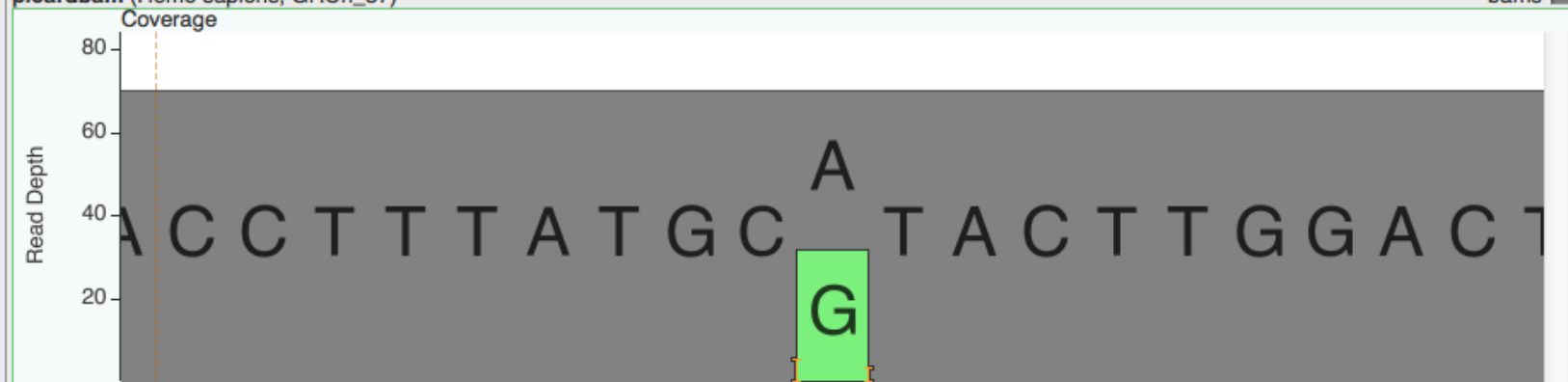
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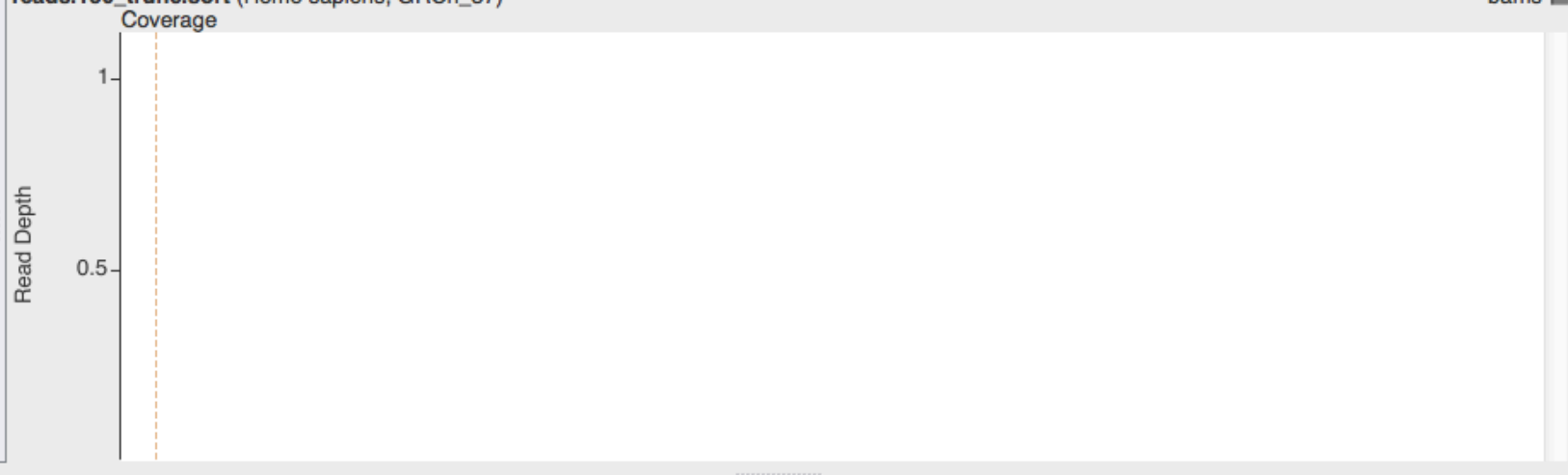
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bams



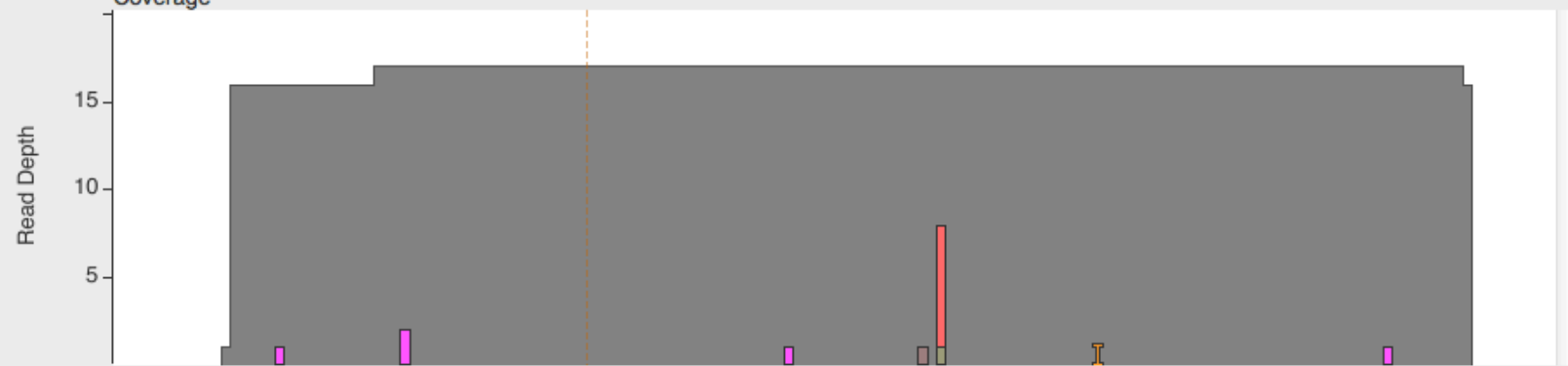
reads.150_trunc.sort (Homo sapiens, GRCh_37)

bams



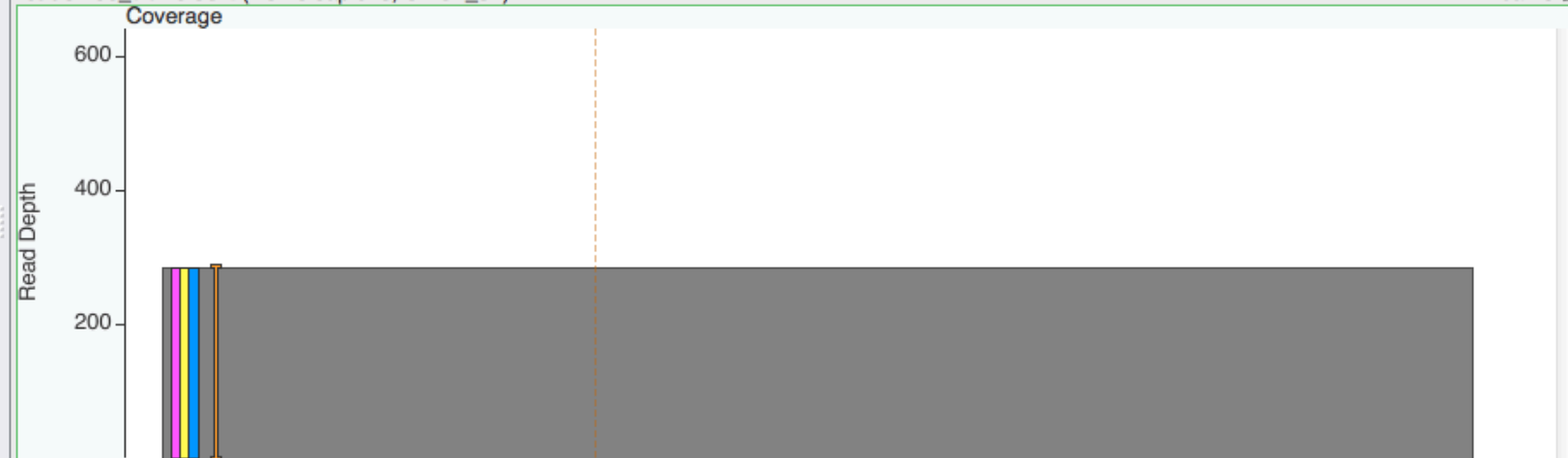
picardbam (Homo sapiens, GRCh_37)
Coverage

bams

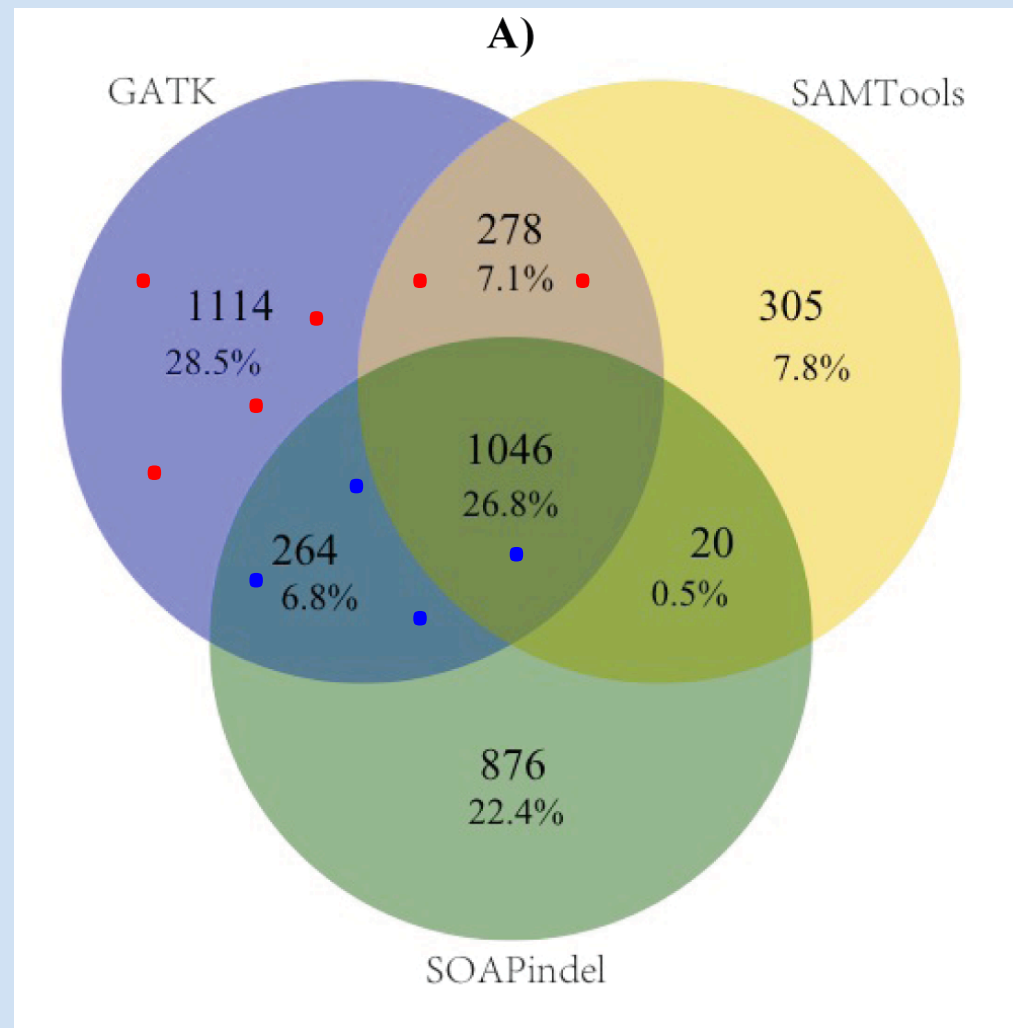


reads.150_trunc.sort (Homo sapiens, GRCh_37)
Coverage

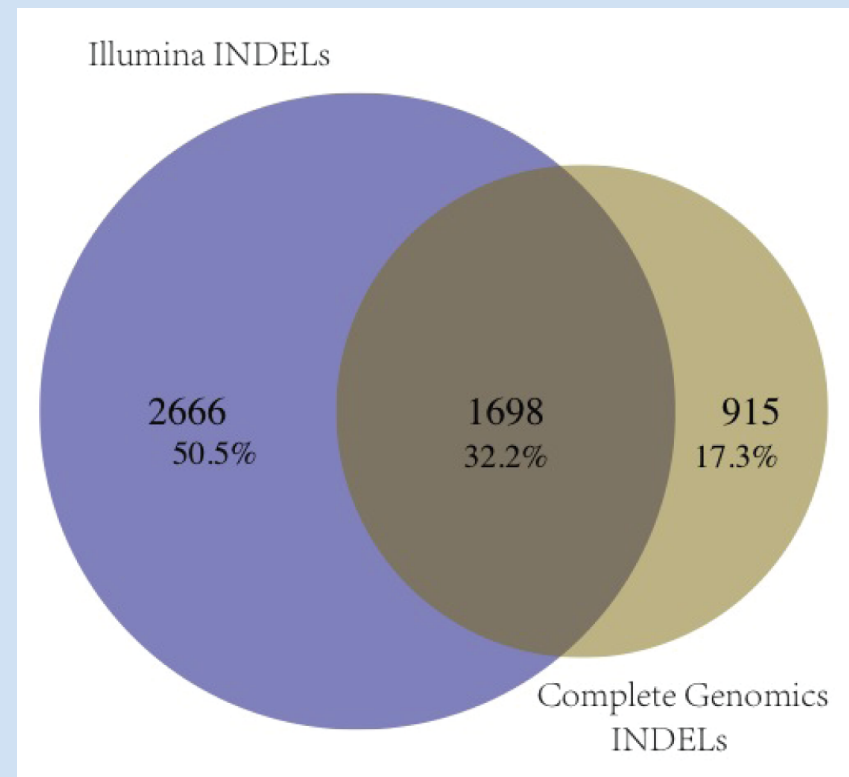
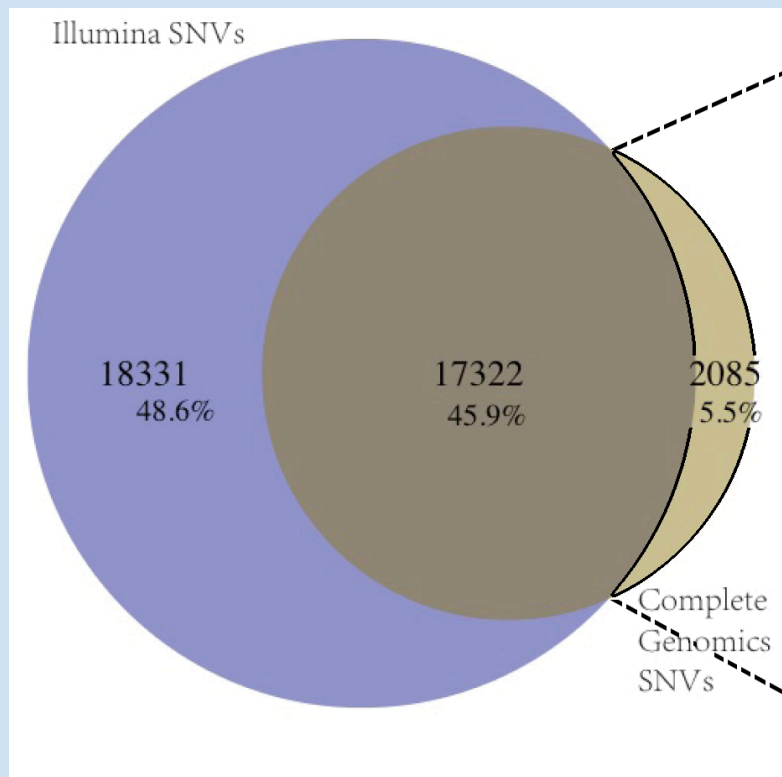
bams



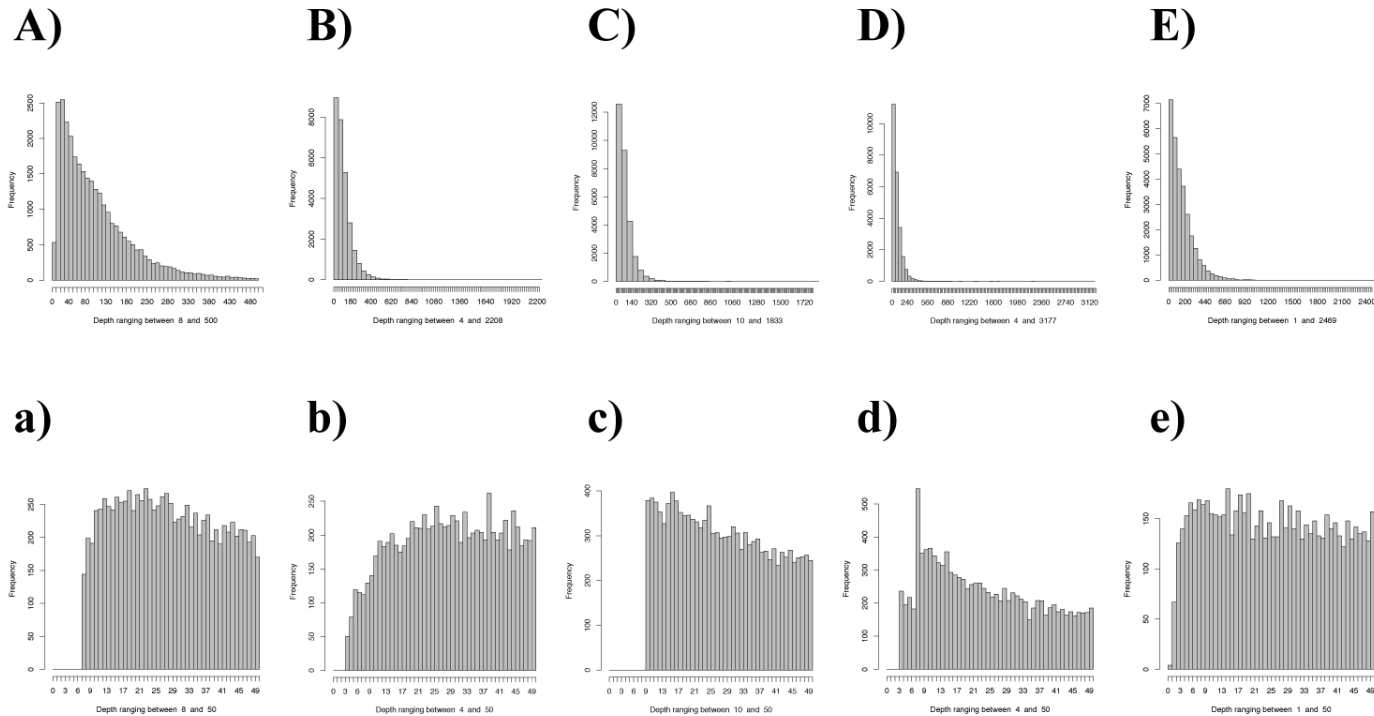
Indels called by GATK, SOAP and SAMtools



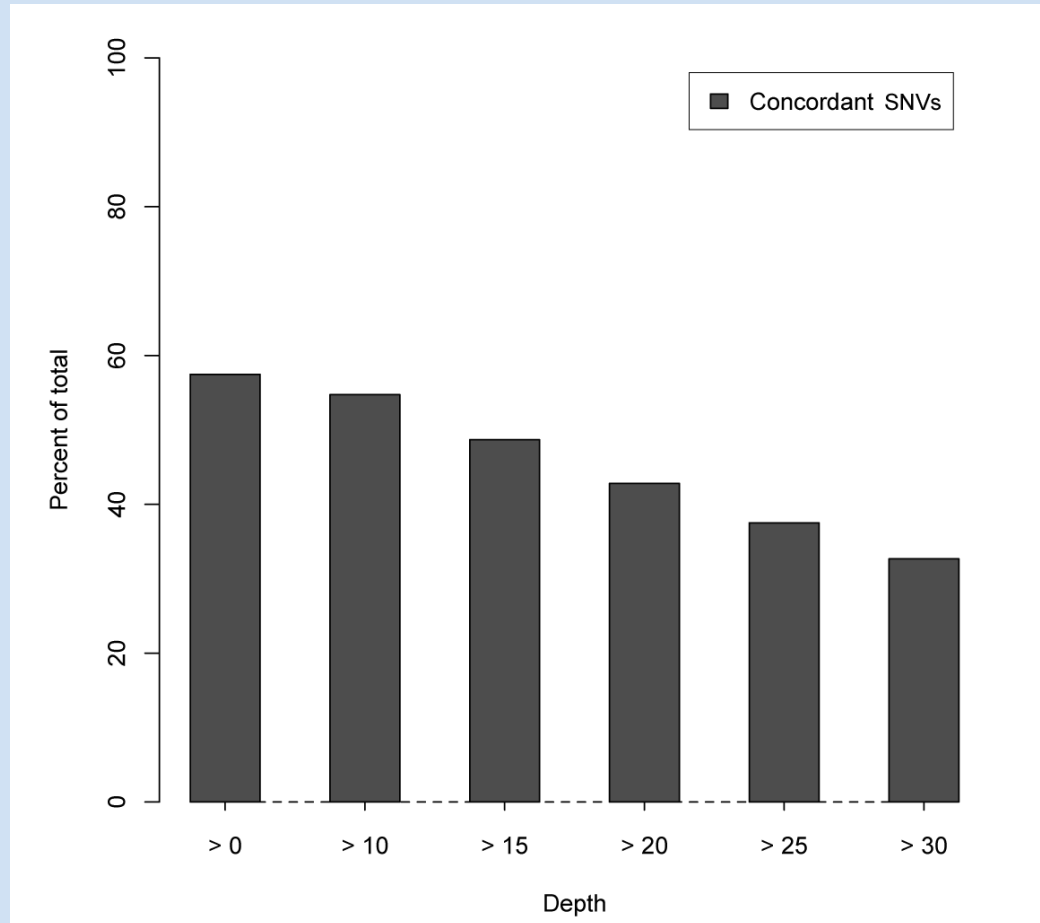
Validation of SNVs and Indels with an additional platform



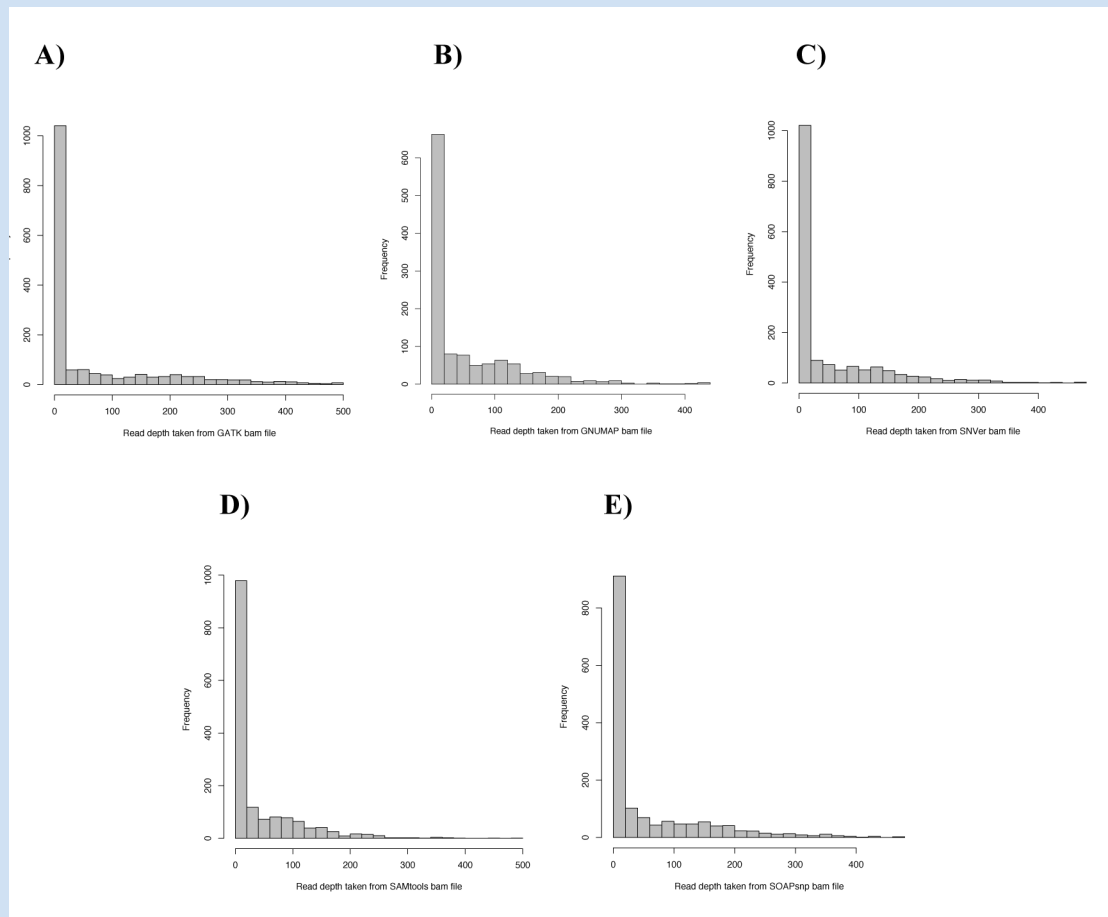
Read Depth



Additional file 2, Figure S3. Histograms of Illumina read depth taken from each pipeline's independently aligned BAM file at genomic coordinates of SNVs called by each of the 5 alignment and variant calling pipelines. **A)** SOAPsnp, **B)** SNVer, **C)** SAMTools, **D)** GNUMAP and **E)** GATK, respectively. Frequency of read depths for all SNVs (**A**, **B**, **C**, **D**, and **E**) as well as for SNVs having depths between 0 and 50 (**a**, **b**, **c**, **d**, and **e**) were plotted.

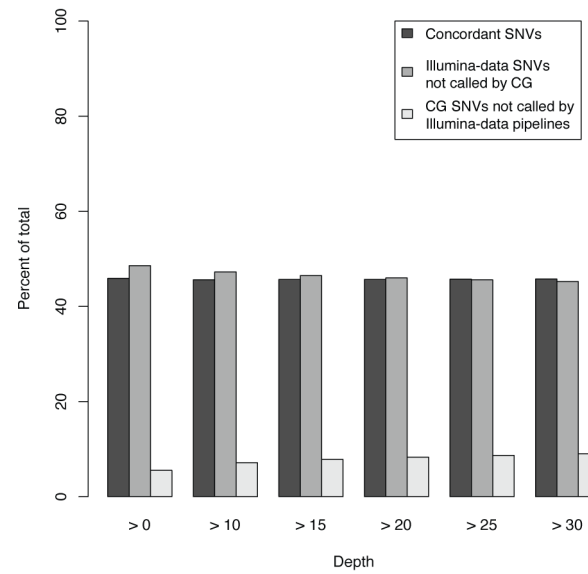


Additional file 2, Figure S4. SNV concordance for a single exome, “k8101-49685”, between five alignment and variant detection pipelines: GATK, SOAPsnp, SNVer, SAMTools, and GNUMAP. Concordance between each pipeline was determined by matching the genomic coordinate as well as the base pair change and zygosity for each detected SNV. Concordance was measured at varying Illumina read depth threshold values in each independently aligned BAM file, ranging from >0 (no threshold) to >30 reads.

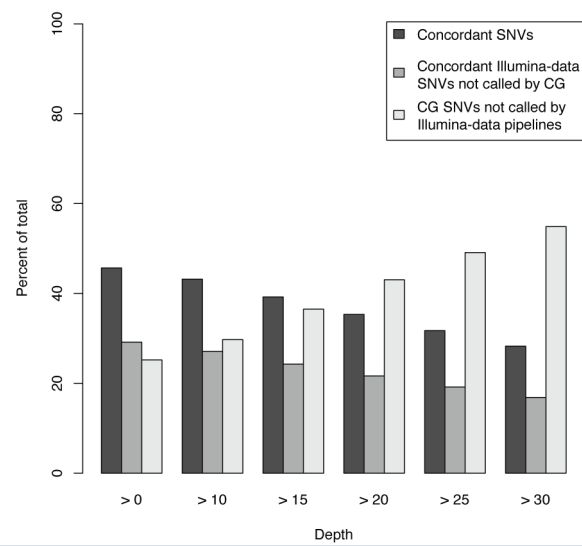


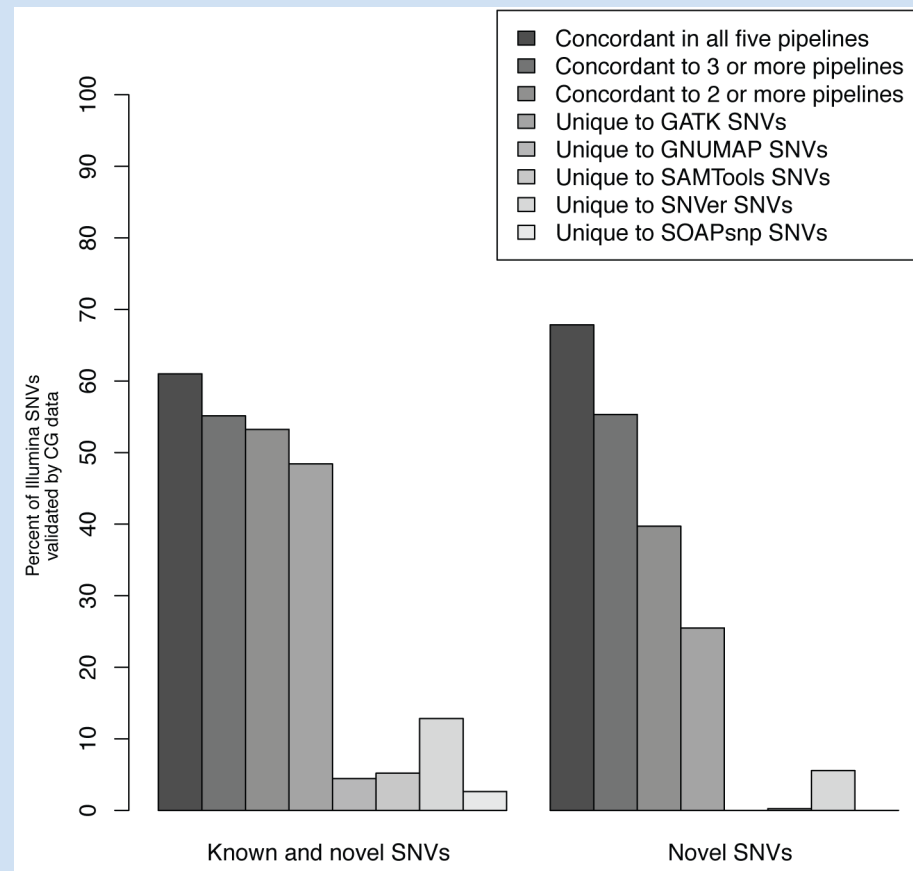
Additional file 2, Figure S5. Histograms of read depth taken from each of the five Illumina pipeline's independently aligned BAM file at genomic coordinates of SNVs that were found by Complete Genomics but not by any of the 5 Illumina pipelines: GATK, GNUMAP, SNVer, SAMTools and SOAPsnp, **A, B, C, D** and **E** respectively. All coordinates fell within the range of the Agilent SureSelect v.2 exons.

A)

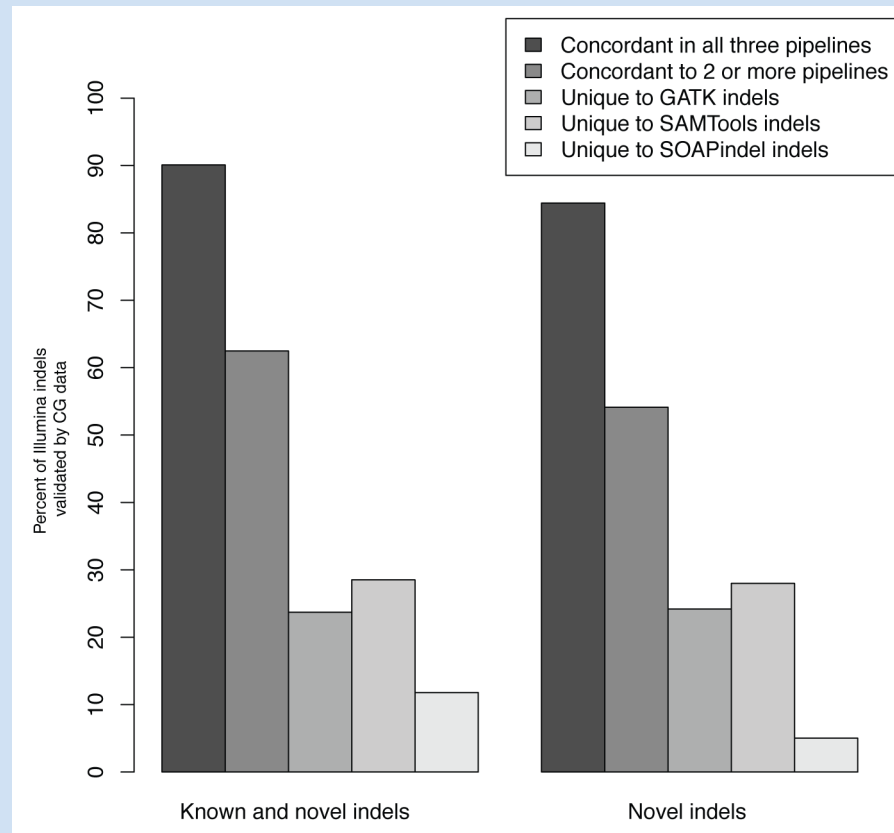


B)





Additional file 2, Figure S8. SNVs called by each Illumina-data pipeline were cross-validated using SNVs called by Complete Genomics, an orthogonal sequencing technology, in sample “k8101-49685”. The percentage of Illumina SNVs that were validated by CG sequencing was measured for variants having varying degrees of Illumina-data pipeline concordance. The same analysis was performed for variants that were considered novel (absent in dbSNP135).



Additional file 2, Figure S9. Indels called by each Illumina-data pipeline were cross-validated using indels called by Complete Genomics for sample “k8101-49685”. The percentage of Illumina indels that were validated by CG sequencing was measured across varying degrees of Illumina pipeline concordance. The same analysis was done for novel indels (indels not found in dbSNP 135).

Comparing the concordance among the 5 pipelines used to analyze Illumina data, also stratified by read depth from >0 to >30 reads.

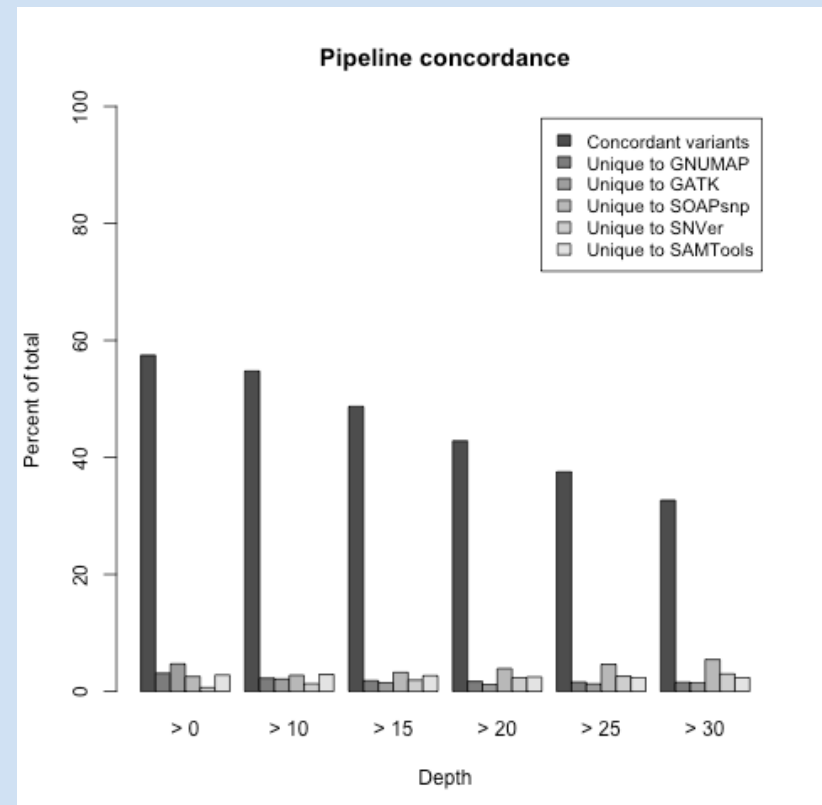
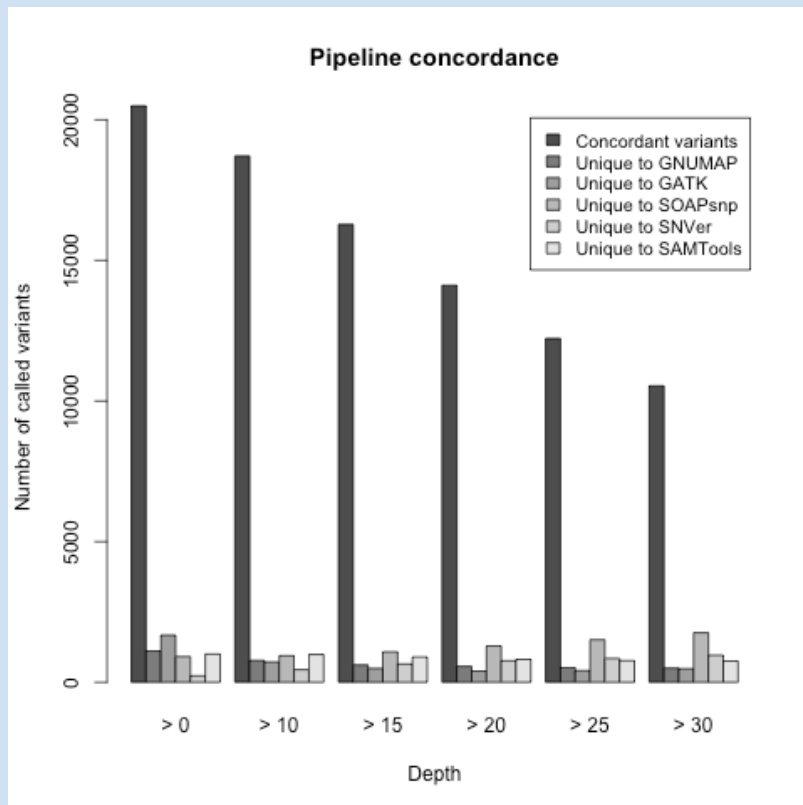


Table 2. Quality evaluation of variant detection using different variant-calling pipelines.

	Sensitivity		Specificity	
	Mean*	SD	Mean*	SD
SOAPsnp	94.68	2.26	99.79	0.03
GATK1.5	95.34	1.16	99.72	0.08
SNVer	92.33	4.40	99.78	0.04
GNUMAP	86.60	3.23	99.64	0.06
SAMtools	94.47	4.22	99.59	0.16
Any pipeline	97.67	1.20	99.62	0.11
≥2 pipelines*	96.64	2.28	99.69	0.07
≥3 pipelines*	95.62	3.13	99.73	0.05
≥4 pipelines*	92.60	3.40	99.82	0.04
5 pipelines*	80.58	5.26	99.87	0.01

*Intersection of variants contained in the number of pipelines specified.

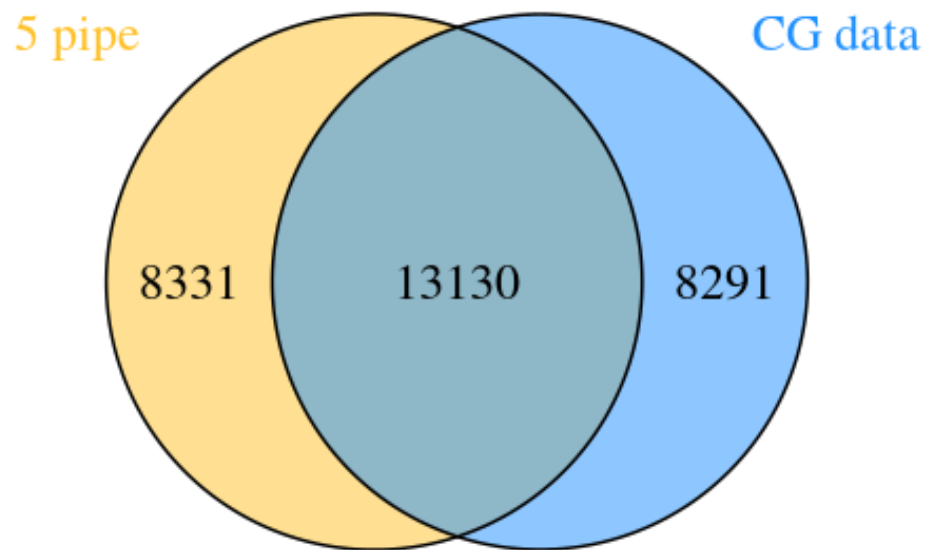
Sensitivity and specificity was calculated for each pipeline by comparing Illumina Human610-Quad version 1 SNP arrays with exome-capture sequencing results, based on the four samples whose genotyping data was available.

Table S1. Concordance rates with common SNPs genotyped on Illumina 610K genotyping chips.

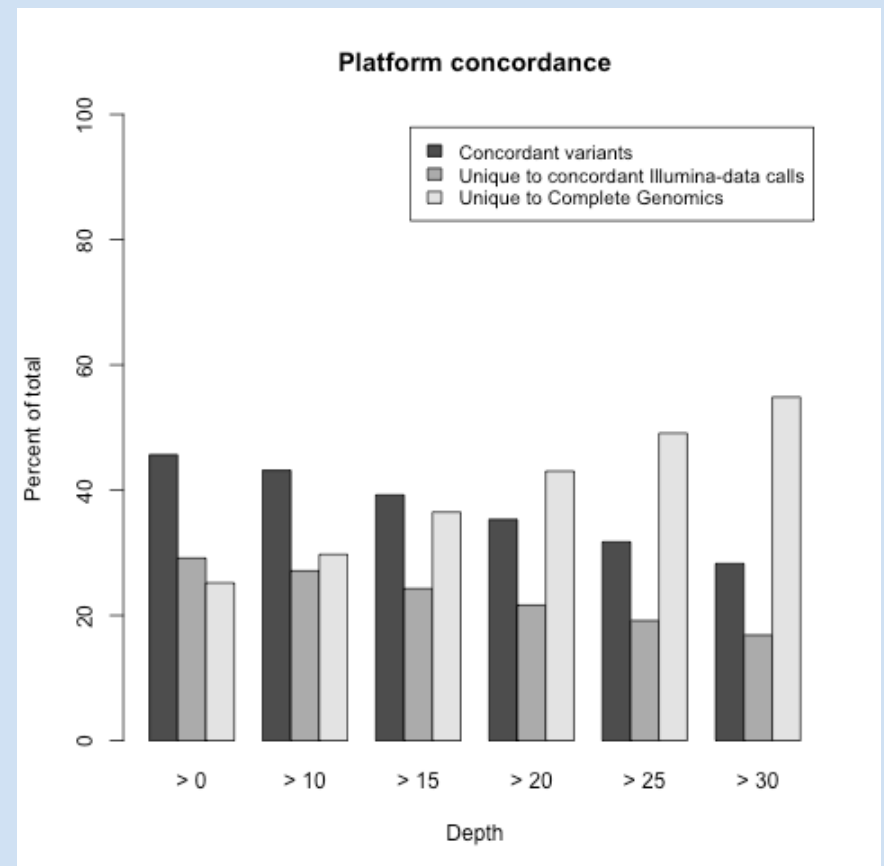
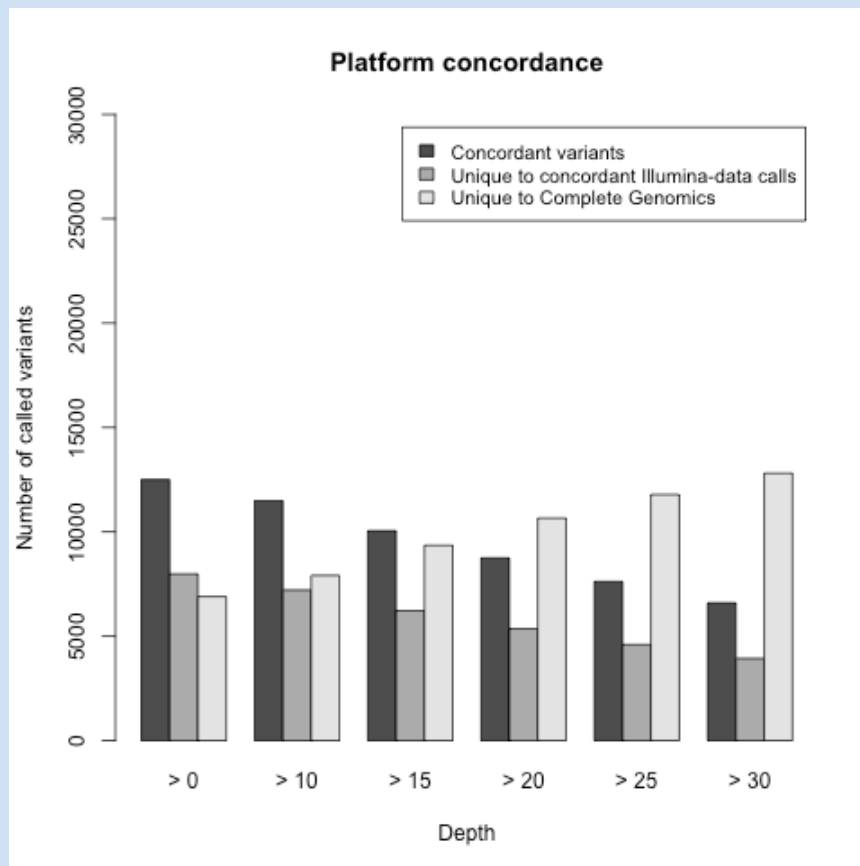
Sample	Software	Compared Sites	Concordance Sites	Concordance rate
Mother-1	SOAPsnp	6088	6074	99.77%
	GATK 1.5	6249	6224	99.60%
	SNVer	5723	5708	99.74%
	GNUMAP	5458	5434	99.56%
	SAMTools	5885	5848	99.37%
Son-1	SOAPsnp	6366	6353	99.80%
	GATK 1.5	6341	6323	99.72%
	SNVer	6255	6239	99.74%
	GNUMAP	5850	5828	99.62%
	SAMTools	6383	6362	99.67%
Son-2	SOAPsnp	6412	6401	99.83%
	GATK 1.5	6426	6413	99.80%
	SNVer	6336	6325	99.83%
	GNUMAP	5906	5889	99.71%
	SAMTools	6477	6450	99.58%
Father-1	SOAPsnp	6247	6238	99.86%
	GATK 1.5	6304	6288	99.75%
	SNVer	6205	6192	99.79%
	GNUMAP	5805	5786	99.67%
	SAMTools	6344	6327	99.73%

All pipelines are very good with identifying already known, common SNPs.

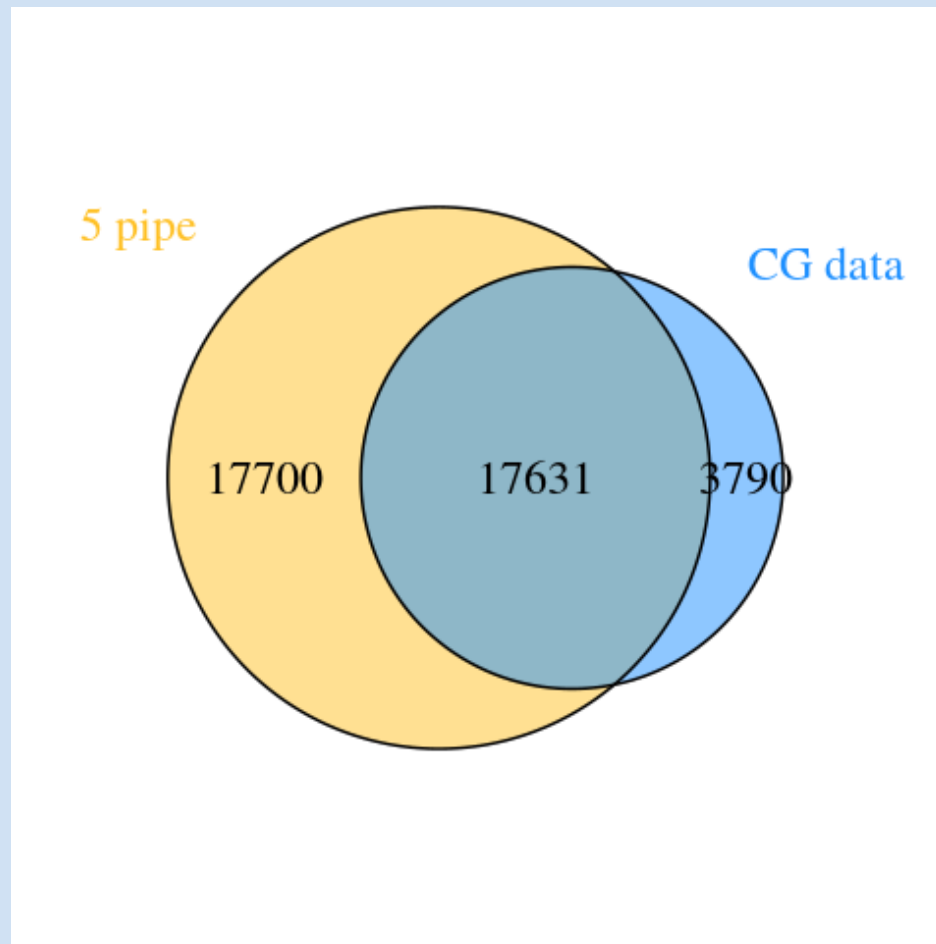
Taking SNVs concordant in 5 Illumina pipelines, and comparing to SNVs in Complete Genomics Data from same sample



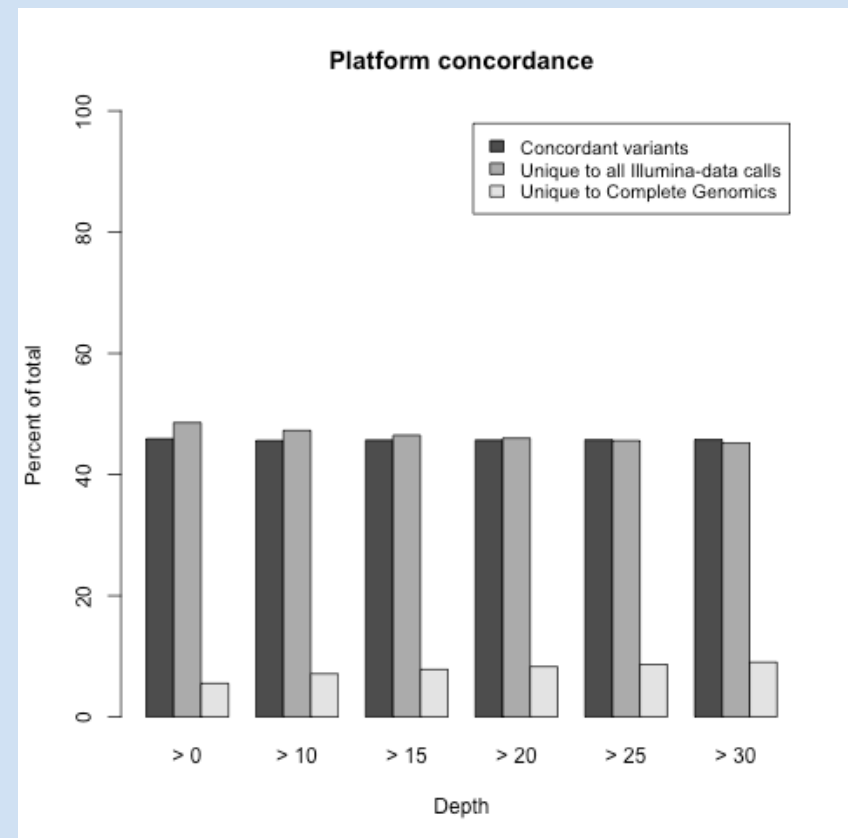
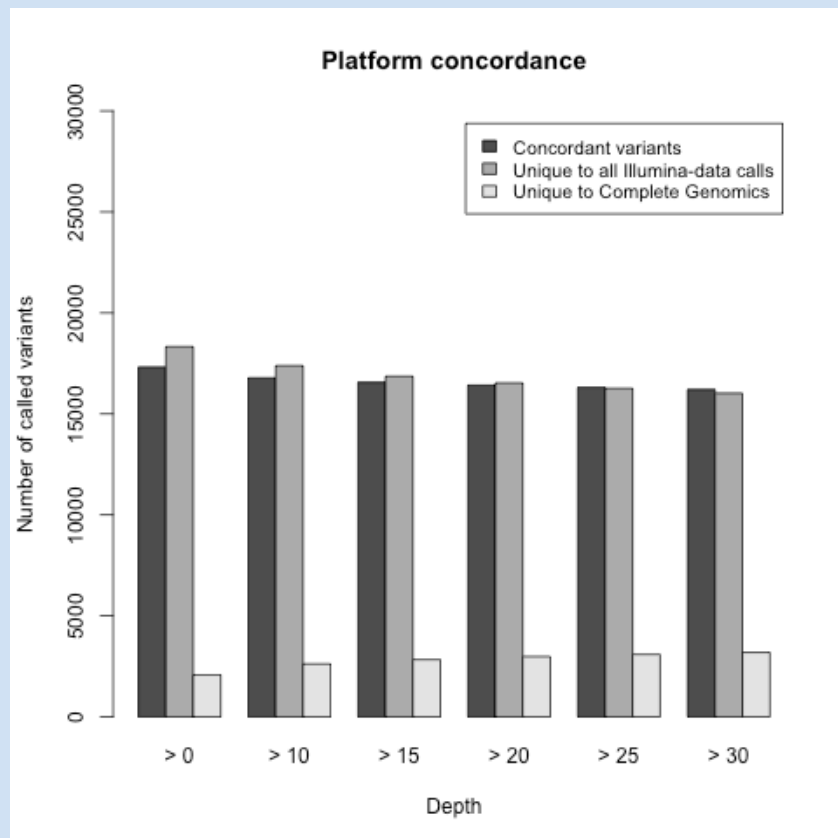
Taking SNVs concordant in 5 Illumina pipelines as per READ DEPTH, and comparing to SNVs in Complete Genomics Data from same sample



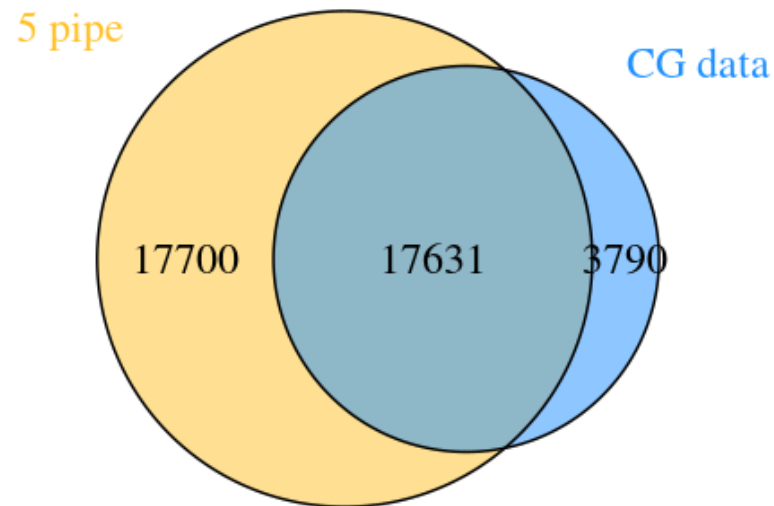
Taking SNVs found by ALL 5 Illumina pipelines (Union), and comparing to SNVs in Complete Genomics Data from same sample



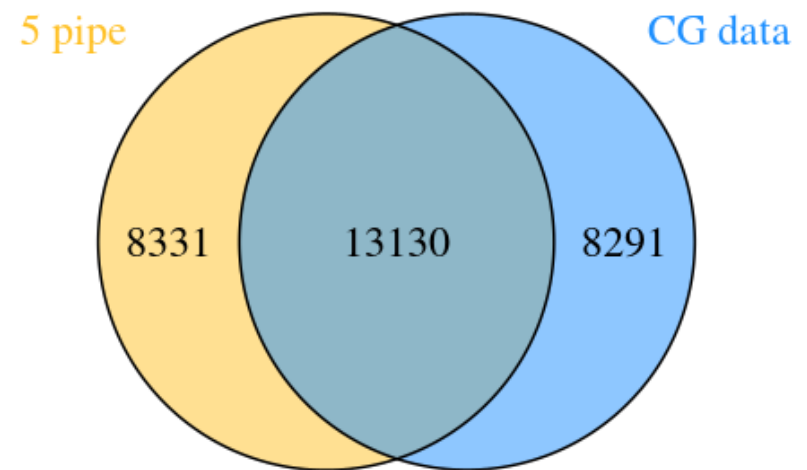
Taking the UNION of all SNVs called by Illumina pipelines, as per READ DEPTH, and comparing to SNVs in Complete Genomics Data from same sample



Comparing the UNION versus the CONCORDANCE of 5 pipelines to the Complete Genomics Data

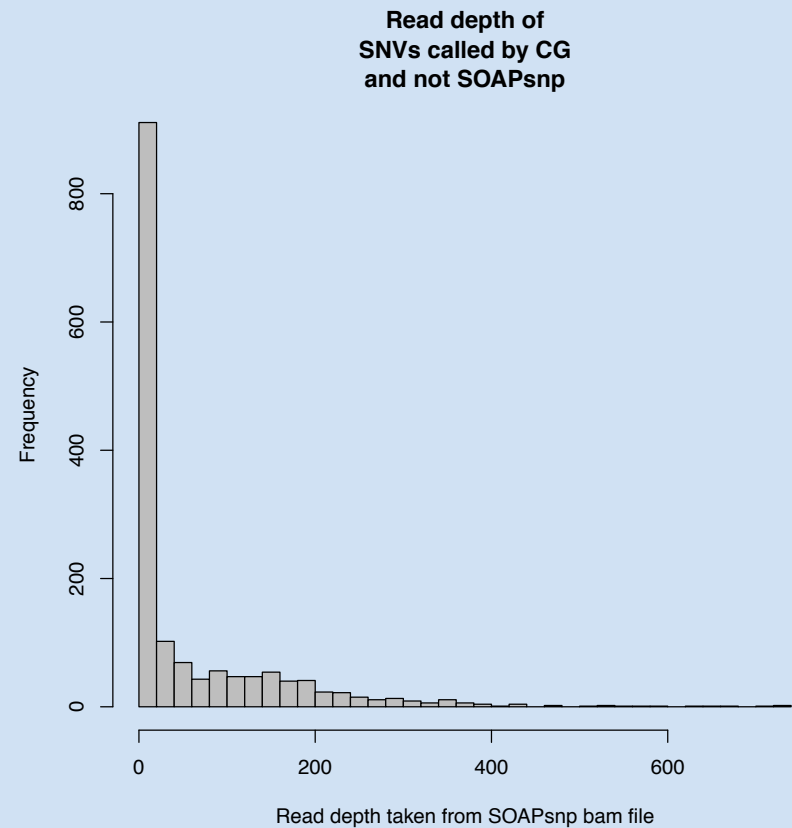
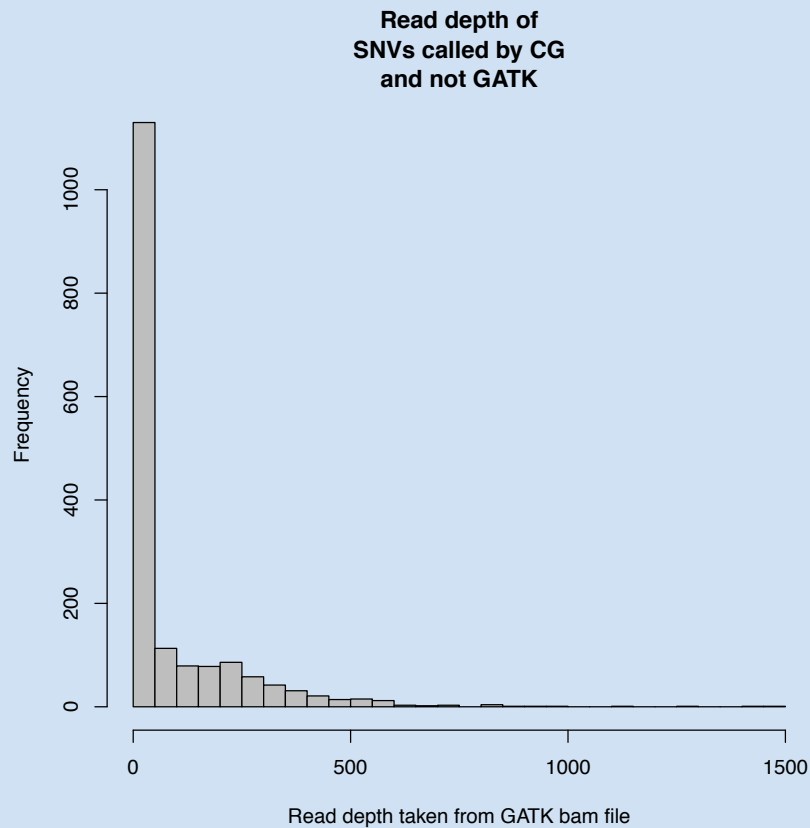


Union of Illumina variants

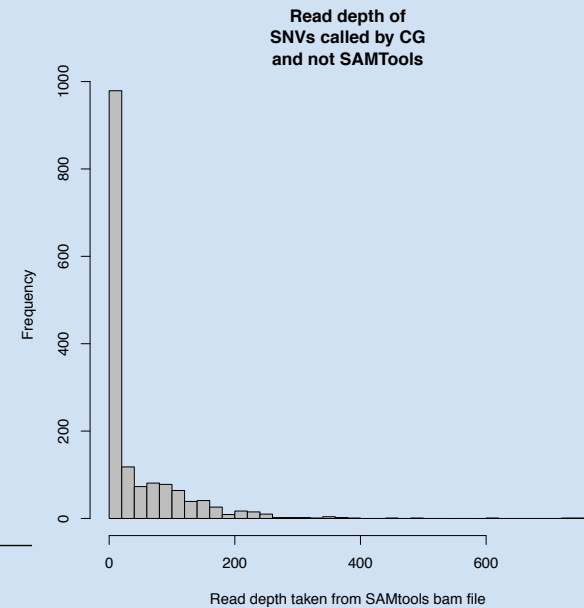
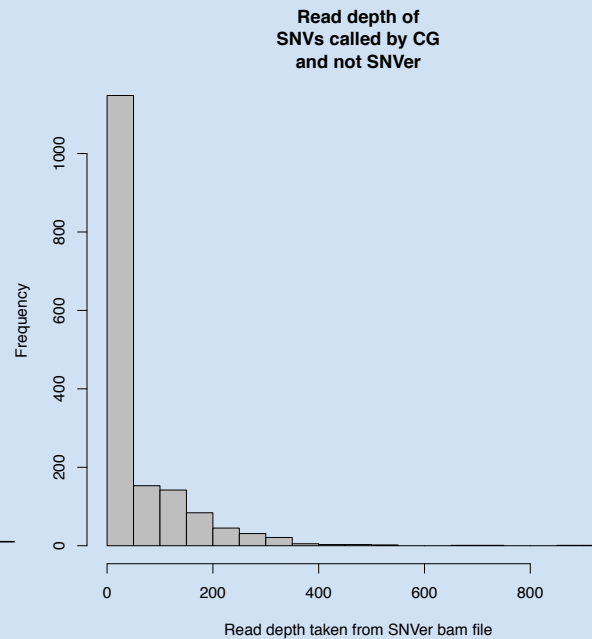
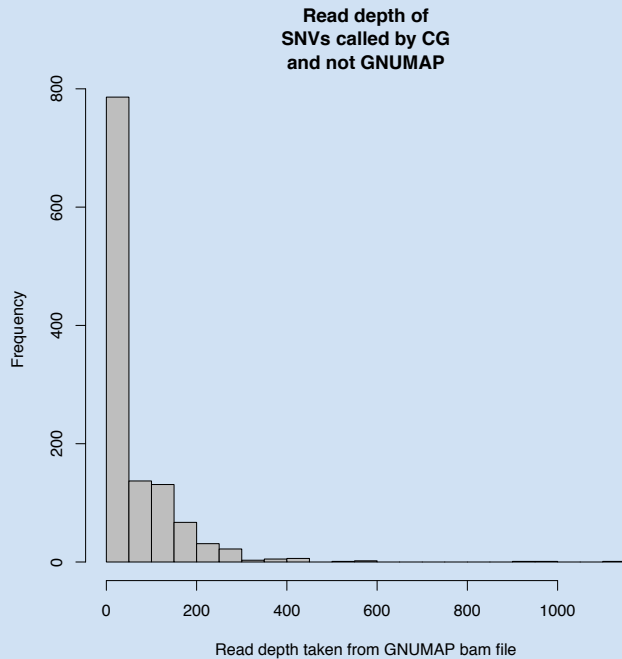


Concordant Illumina variants

Read Depth of Illumina Reads for variants called by Complete Genomics but NOT by GATK or SOAP pipelines



Read Depth of Illumina Reads for variants called by Complete Genomics but NOT by GNUMAP, SNVer or SamTools pipelines



Genomic Dark Matter, cont....

- That means that unlike typical false negatives, increasing coverage will not help identify mutations in low GMS regions, even with 0% sequencing error.
- Instead this is because the SNP-calling algorithms use the mapping quality scores to filter out unreliable mapping assignments, and low GMS regions have low mapping quality score (by definition). Thus even though many reads may sample these variations, the mapping algorithms cannot ever reliably map to them.
- Since about 14% of the genome has low GMS value with typical sequencing parameters, it is expected that about 14% of all variations of all resequencing studies will not be detected.
- To demonstrate this effect, we characterised the SNP variants identified by the 1000 genomes pilot project, and found that 99.99% of the SNPs reported were in high GMS regions of the genome, and in fact 99.95% had GMS over 90.

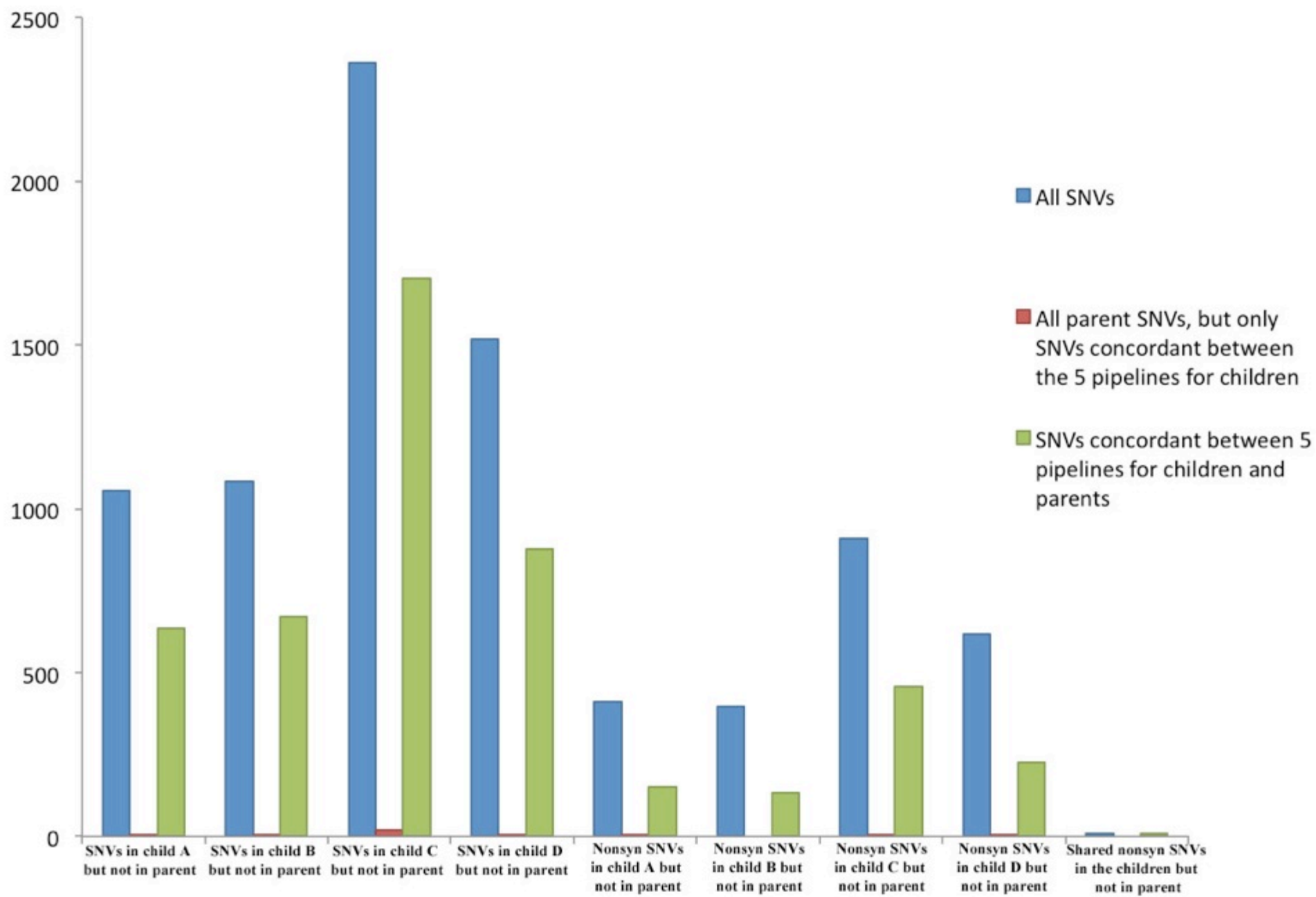
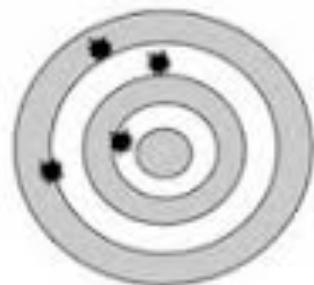


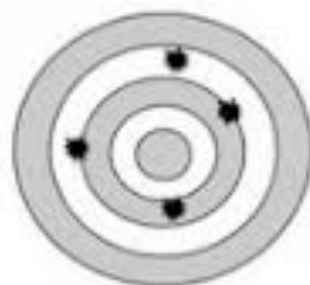
Figure 1. Mean single-nucleotide variants (SNV) concordance over 15 exomes between five alignment and variant-calling pipelines. The alignment method used, followed by the SNV variant calling algorithm is annotated here in shorthand: BWA-GATK, SOAP-Align-SOAPsnp, BWA-SNVer, BWA-SAMtools, and GNUMAP-GNUMAP. **(A)** Mean SNV concordance between each pipeline was determined by matching the genomic coordinate as well as the base-pair change and zygosity for each detected SNV. **(B)** The same analysis as in (A) but filtered to include only SNVs already found in dbSNP135. **(C)** The same analysis as in (A), but filtered to include novel SNVs (that is, SNVs not found in dbSNP135).

Pipelines Used on Same Set of Seq Data by Different Analysts, using Hg19 Reference Genome

- 1) BWA - Sam format to Bam format - Picard to remove duplicates - **GATK** (version 1.5) with recommended parameters (GATK IndelRealigner, base quality scores were re-calibrated by GATK Table Recalibration tool. Genotypes called by GATK UnifiedGenotyper.
- 2) BWA - Sam format to Bam format-Picard to remove duplicates - **SamTools** version 0.1.18 to generate genotype calls -- The “mpileup” command in SamTools were used for identify SNPs and indels.
- 3) **SOAP**-Align – SOAPsnp – then BWA-SOAPindel (adopts local assembly based on an extended de Bruijn graph)
- 4) **GNUMAP-SNP** (probabilistic Pair-Hidden Markov which effectively accounts for uncertainty in the read calls as well as read mapping in an unbiased fashion)
- 5) BWA - Sam format to Bam format - Picard to remove duplicates - **SNVer**
- 6) BWA - Sam format to Bam format - Picard to remove duplicates - **SCALPEL**



Not Accurate
Not Precise



Accurate
Not Precise

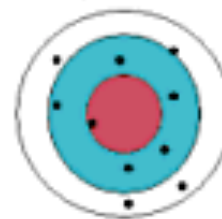


Not Accurate
Precise



Accurate
Precise

A Both accuracy and precision



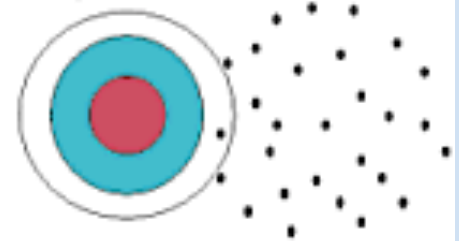
B Accuracy only



C Precision only



D Neither accuracy nor precision



Pipelines Used on Same Set of Seq Data by Different Analysts, using Hg19 Reference Genome

- 1) BWA - **GATK** (version 1.5) with recommended parameters (GATK IndelRealigner, base quality scores were re-calibrated by GATK Table Recalibration tool. Genotypes called by GATK UnifiedGenotyper. For SNVs and indels.
- 2) BWA - **SamTools** version 0.1.18 to generate genotype calls -- The “mpileup” command in SamTools was used for identify SNVs and indels.
- 3) **SOAP**-Align – SOAPsnp for SNVs– and BWA-SOAPindel (adopts local assembly based on an extended de Bruijn graph) for indels.
- 4) **GNUMAP-SNP** (probabilistic Pair-Hidden Markov which effectively accounts for uncertainty in the read calls as well as read mapping in an unbiased fashion), for SNVs only.
- 5) BWA - Sam format to Bam format - Picard to remove duplicates – **SNVer** , for SNVs only

	All SNVs, both for parents and children, were considered	All parental SNVs that were detected were considered. Only SNVs concordant between the 5 pipelines were considered for children	SNVs concordant between 5 pipelines for children and parents
Number of SNVs found in child A but not in parents	1057	2	637
Number of SNVs found in child B but not in parents	1084	1	672
Number of SNVs found in child C but not in parents	2363	20	1703
Number of SNVs found in child D but not in parents	1518	5	876
Number of nonsyn SNVs in child A but not in parents	411	1	150
Number of nonsyn SNVs in child B but not in parents	396	0	135
Number of nonsyn SNVs in child C but not in parents	911	6	459
Number of nonsyn SNVs in child D but not in parents	619	3	225
Number of shared nonsyn SNVs in the children, but not in parents	8	0	9

Optimizing pipeline based on literature value of ~1 true de novo protein-altering mutation per exome

	All SNVs, both for parents and children, were considered	All parental SNVs that were detected were considered. Only SNVs concordant between the 5 pipelines were considered for children	SNVs concordant between 5 pipelines for children and parents
Number of SNVs found in child A but not in parents	1308	186	1795
Number of SNVs found in child B but not in parents	1332	161	1762
Number of nonsyn SNVs in child A but not in parents	381	52	420
Number of nonsyn SNVs in child B but not in parents	392	42	394
Number of shared nonsyn SNVs in the children, but not in parents	98	14	171

The result is that using all of the detected SNVs for both parents and children should minimize the false negative rate but similarly show a relatively high false positive rate. Using all of the SNVs detected for parents but only the SNVs concordant among the five pipelines shows mutation rates similar to those reported by the literature and is expected to have moderate false positive rates and moderate false negative rates. Using only the SNVs concordant among the 5 different pipelines for both parents and children should minimize the false positive rate but similarly show a relatively high false negative rate.

Much Higher Validation of the Concordantly Called SNVs (by the CG data)

