

Clinical genetics of neurodevelopmental disorders

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Introduction

“our incomplete studies do not permit actual classification; but it is better to leave things by themselves rather than to force them into classes which have their foundation only on paper” - Seguin¹

“The fundamental mistake which vitiates all work based upon Mendel’s method is the neglect of ancestry, and the attempt to regard the whole effect upon offspring, produced by a particular parent, as due to the existence in the parent of particular structural characters; while the contradictory results obtained by those who have observed the offspring of parents apparently identical in certain characters show clearly enough that not only the parents themselves, but their race, that is their ancestry, must be taken into account before the result of pairing them can be predicted” - Weldon².

There are ~6 billion nucleotides in every cell of the human body, and there are ~25-100 trillion cells in each human body. Given somatic mosaicism, epigenetic changes and environmental differences, no two human beings are the same, particularly as there are only ~7 billion people on the planet. One of the next great challenges for studying human genetics will be to acknowledge and embrace complexity³⁻¹¹. Every human is unique, and the study of human disease phenotypes (and phenotypes in general) will be greatly enriched by moving from a deterministic to a more stochastic/probabilistic model¹²⁻¹⁷. The dichotomous distinction between ‘simple’ and ‘complex’ diseases is completely artificial, and we argue instead for a model that considers a spectrum of diseases that are variably manifesting in each person.

The rapid adoption of whole genome sequencing (WGS) and the Internet-mediated networking of people promise to yield more insight into this century-old debate^{2, 18-23}. Comprehensive ancestry tracking and detailed family history data, when combined with WGS or at least cascade-carrier screening²⁴, might eventually facilitate a degree of genetic prediction for some diseases in the context of their familial and ancestral etiologies. However, it is important to remain humble, as our current state of knowledge is not yet sufficient, and in principle, any number of nucleotides in the genome, if mutated or modified in a certain way and at a certain time and place, might influence some phenotype during embryogenesis or postnatal life^{9, 25-42}.

Clinical classifications and the genetic architecture of disease

“Those who have given any attention to congenital mental lesions, must have been frequently puzzled how to arrange, in any satisfactory way, the different classes of this defect which may have come under their observation. Nor will the difficulty be lessened by an appeal to what has been written on the subject. The systems of classification are generally so vague and artificial, that, not only do they assist but feebly, in any mental arrangement of the phenomena represented, but they completely fail in exerting any practical influence on the subject.” – Down⁴³

As most clinicians know from experience, it is quite difficult to characterize the range of human experience in the two-dimensional world of the printed page, as we are attempting to do here. In addition, classifications can sometimes lead people to try to force round pegs into square holes, and so

we are reluctant to further promulgate these classifications. Such classifications include terms such as: ‘Mendelian’, ‘complex disease’, ‘penetrance’, ‘expressivity’, ‘oligogenic’, and ‘polygenic’. For example, some have used the word ‘Mendelian’ to refer to a disease that appears to somehow be

‘caused’ by mutations in a single gene. As such, cystic fibrosis, Huntington’s disease, and Fragile X are all diseases that some people refer to as being ‘caused’ by mutations occurring in single genes. However, the expression of the phenotype within these diseases is extremely variable, depending in part on the exact mutations in each gene, and it is not at all clear that any mutation really and truly ‘causes’ any phenotype, at least not according to thoughtful definitions of causation that we are aware of^{44, 45}. For example, some children with certain mutations in *CFTR* may only have pancreatitis as a manifestation of cystic fibrosis, without any lung involvement^{46, 47}, and there is evidence that mutations in other genes in the genomes can have a modifying effect on the phenotype^{48, 49}. In the case of Huntington’s, there is extreme variability in the expression of the phenotype, both in time, period and scope of illness, and all of this is certainly modified substantially by the number of trinucleotide repeats⁵⁰, genetic background⁵¹ and environmental influences⁵². Even in the case of whole chromosome disorders, such as Down Syndrome, there is ample evidence of substantial phenotypic expression differences, modified again by genetic background^{53, 54}, somatic mosaicism⁵⁵ and environmental influences^{56, 57}, including synaptic and brain plasticity^{17, 58-61}. The same is true for deletion syndromes, such as velocardiofacial syndrome and other deletions⁶²⁻⁶⁷. And, of course, there is constant interaction of the environment with a person, both prenatally and postnatally. As just one example, cretinism is related to a lack of iodine in the mother’s diet, and there is incredibly variable expression of this illness based in part on the amount of iodine deficiency and how this interacts with fetal development⁶⁸.

The words ‘penetrance’ and ‘expressivity’ have been defined as:

- *Penetrance*: whether someone in a population has any symptoms of a disease, where the definition of having the disease is categorically defined as all or none, 0% or 100%.
- *Expressivity*: how much disease (or how many symptoms) someone with 100% penetrance has.

Unfortunately, these two separate terms have led to a great deal of confusion in the field. Some use the word ‘penetrance’ when they really mean

‘expressivity’ of disease in any one person, as they write things like ‘incomplete penetrance’, despite the fact that this completely destroys the original definition of the word ‘penetrance’. As such, perhaps we should get rid of the two terms altogether and just discuss the expression of each trait in the context of a phenotypic spectrum, which is of course what led Walter Frank Raphael Weldon to establish the field of biometry^{13, 69, 70}. Another way to express this point is to say that we have yet to characterize the full breadth of expression for virtually any mutation in humans, as we have not systematically sequenced or karyotyped any genetic alteration in thousands to millions of randomly selected people from a whole range of ethnic classes, i.e. clans^{71, 72}. There is an ongoing clash of world-views, with some wanting to believe that single mutations predominately drive outcome while others are explicitly acknowledging the importance of substantial phenotypic modification via genetic background and/or environmental influence(s)^{5, 25, 73-79}.

It is very likely that there will be a continuum of disease, given that the ‘effect size’ of any particular mutation will obviously vary according to genetic background and environment, as demonstrated repeatedly in model organisms^{73-76, 80-87}. Thus, while a mutation associated with hemochromatosis or breast cancer might have high expression in one particular pedigree or clan, that same mutation may have very low expression in another pedigree, clan or group of unrelated people⁸⁸. The reasons for variable expression can be myriad and are currently unknown in many instances; however, problems start to appear when scientists attempt to invoke a third allele as necessary and perhaps sufficient for the expression of any symptoms from within a typical disease. This disease model has been most clearly advocated for Bardet-Biedl Syndrome, in which the authors contend that some subjects have zero disease symptoms while possessing two autosomal recessive mutations in a known ‘disease gene’; the authors also show that some affected people have a mutation in another gene, i.e. a third allele, which they speculate is necessary and perhaps sufficient for expression of any symptoms of the disease⁸⁹⁻⁹¹. However, this model has been challenged by others⁹²⁻⁹⁶, and at least one group maintains that all people that they have studied with two autosomal recessive mutations have 100% ‘penetrance’, but with variable

expression, i.e. one person might only have retinitis pigmentosa whereas another person might have the full-blown symptoms of Bardet-Biedl syndrome⁹². One wonders whether the debate about triallelism, with this idea of 0% ‘penetrance’ in the absence of a third allele, might really just be a semantic one due to problems with the phenotyping of ‘unaffected’ people, particularly if these people were not evaluated longitudinally. Detailed online longitudinal characterizations of all such reportedly ‘unaffected’ people could aid in documenting, with some degree of certainty, that these people did indeed have zero symptoms of Bardet-Biedl syndrome, as that would then be further proof that mutations are not deterministic in any way at all. Said another way, this would be demonstration of true zero ‘penetrance’ for mutations that do contribute to a phenotype in other people with their own genetic backgrounds and environmental differences, and this observation ought to have dramatic implications for any ideas concerning prenatal diagnosis and ‘prediction’ of any genotype/phenotype relationship (discussed more below).

Surprisingly, a precise definition of the term ‘oligogenic’ is not apparent or consistent in the world literature. Some people have invoked the term ‘oligogenic’ to mean an interaction between mutations in two genes to somehow collectively ‘cause’ a disease, such as with this above case of triallelism in Bardet-Biedl syndrome⁹⁷. These authors define oligogenic inheritance as occurring “when specific alleles at more than one locus affect a genetic trait by causing and/or modifying the severity and range of a phenotype”⁹⁷. Another case in point involves the 22q11.2 locus, also known as velocardiofacial syndrome. This deletion does not involve only a single gene, but rather ~X number of genes, depending on the exact size of the deletion interval. The phenotypic manifestations can be incredibly heterogeneous, illustrated by the fact that some ~30% develop psychotic symptoms and get labeled as ‘schizophrenic’⁹⁸. Of course, heuristic diagnoses for schizophrenia are usually made based on certain semantic criteria, so it is likely that subthreshold symptoms are not counted (or perhaps not even detected). But, at least one has the advantage of knowing which people possess the deletion, allowing one to perform detailed phenotyping to determine whether subthreshold symptoms were missed within a family, and this has

indeed been done in the case of a well-known translocation involving *DISC1*^{99, 100}. Unfortunately, genome-wide studies are not yet performed routinely for people with ‘idiopathic schizophrenia’, so it has been difficult to identify and group many people by genotype(s). As we discuss below, we believe that the routine clinical use of exome and eventually whole genome sequencing might finally enable this to occur, assuming that aggregation of genotype and phenotype data is allowed on a massive scale.

The definition of ‘polygenic’ seems to have historically been associated with the combined effects of dozens (or perhaps even hundreds) of different mutations in different genes to result in a particular phenotype. Height has historically been characterized as being a polygenic phenotype, with GWAS studies implicating the possible involvement of hundreds of loci^{101, 102}. Height is an easily measured, continuous, phenotype and in human populations it has been shown to be distributed according to a Gaussian function, although the measurement of someone’s height from the top of their head to the bottom of their feet is a one-dimensional measurement in the context of something that is actually developing in four dimensions, if one includes the dimension of time. Therefore, it seems obvious that numerous variables will contribute to this one-dimensional measurement in humans (and other animals) developing over time. Psychiatric symptoms, in comparison, cannot yet even be measured with the same degree of accuracy as height, and so it seems premature to argue that height and schizophrenia (for example) are both continuous phenotypes that can be robustly characterized as being Gaussian in nature. However, this appears to be precisely what is argued by some¹⁰³. We would argue that we simply do not know enough about the phenotypic expression of the many different diseases that ‘schizophrenia’ encompasses to be able to make any conclusions regarding its genetic inheritance on a population level¹⁰⁴. Rather, one must study people within families to determine whether some people in families have disease due to mutations with variable expression, modified by genetic background and environmental influences.

There have been numerous reviews concerning the ongoing debate for common and rare variants, with arguments made for various ‘camps’ of thought, including the common disease-common

variant (CDCV) model, the infinitesimal model, the rare allele model and the broad sense heritability model ¹⁰⁵. Frankly, these models are simply semantic and reductionistic arguments that do not reflect the complexity of the human condition, and we are not sure that arguing for and against various models is useful, given that these models are basically straw men artificially constructed to be knocked down. This is very similar to the psychiatric literature in which several people decided, about 100 years ago, to introduce various names (or models) for certain diseases, such as the words ‘schizophrenia’ ¹⁰⁶ and ‘manic-depressive illness or bipolar’ ¹⁰⁷. It is quite apparent to most clinicians that the phenotypic heterogeneity of these illnesses is so tremendous so as to render these names basically moot and not particularly useful. This is akin to 50 years ago when people simply stated that someone had ‘cancer’. Now, it is not useful to say only that someone has cancer, as there are literally hundreds of molecular etiologies for cancer, divided up not only by organ expression but also by specific pathways in the cell ¹⁰⁸. We anticipate that in 50 years, these terms ‘schizophrenia’ and ‘bipolar’ will be replaced by much more precise molecularly defined terms, as is occurring now in the cancer field ^{109, 110}. Locus heterogeneity will likely play an important role in most diseases, but particularly in psychiatric disease, given the extensive phenotypic heterogeneity. Some of this complexity has been documented in reports of individual people ¹¹¹⁻¹²³, and a review of the literature related to schizophrenia ¹²⁴ rendered the distinct impression that we really hardly know anything about the mechanistic basis of these many illnesses that we currently lump together as ‘schizophrenia’. This is primarily due to overly broad descriptions and categorizations of these illnesses into these artificially named syndromes, despite the obvious heterogeneous and inconsistent nature of these categorizations. Remarkably, bipolar and schizophrenia have been artificially ‘split’ into different syndromes ^{125, 126}, in spite of the existence of a well documented literature demonstrating overlap in at least some families with symptoms from both ‘syndromes’ ¹²⁷.

Oddly enough, some diseases such as Fragile X, Rett Syndrome and other now molecularly defined disorders are sometimes removed from the ‘nonsyndromic idiopathic autism’ camp, leaving the remaining disorders still eligible for a semantic debate about which ‘genetic model’ they fit into ¹²⁸. One wonders if the same thing has occurred for velocardiofacial syndrome, with its relevance to schizophrenia, given the overwhelming evidence that the single 22q11.2 deletion event predisposes its carriers to some version of ‘schizophrenia’ with an expression of 20-30% ⁹⁸. All of these disorders were at one point labeled as ‘idiopathic’ until molecular lesions associated with them were identified. It has been known by at least some researchers and clinicians for quite some time that there are likely many minor physical anomalies in people labeled as ‘nonsyndromic’ ^{129, 130}, all of which is further proof of the substantial phenotypic expression differences of all disorders. Therefore, the dichotomous use of the words ‘syndromic’ and ‘nonsyndromic’ is completely artificial and does not reflect the reality or complexity of the situation in any one person.

A very recent paper using exome sequencing to study hypertension pedigrees made the following statements: “These findings demonstrate the utility of exome sequencing in disease gene identification despite the combined complexities of locus heterogeneity, mixed models of transmission and frequent de novo mutation Gene identification was complicated by the combined effects of locus heterogeneity, two modes of transmission at one locus, and few informative meioses. Many so far unsolved Mendelian traits may have similar complexities. Use of control exomes as comparators for analysis of mutation burden may be broadly applicable to discovery of such loci ”¹³¹. This paper illustrates exactly what we are discussing above, in terms of the possible heterogeneity of many illnesses on many levels, making it impossible to predict (or even need) any particular model that may or may not fit with the disease. It is far better to allow the data to speak for themselves.

De novo mutations, germline mosaicism and other complexities

Although this concept of somatic mosaicism has been in the literature for many years¹³²⁻¹³⁶, it is really only recently that more people are beginning to realize that it might be much more extensive in humans than previously thought^{23, 137-152}. In fact, hardly anything is truly known regarding the extent of somatic mosaicism in humans and its effect on phenotype in even well studied diseases. For example, little is known regarding pathogenesis of the phenotype in people with trisomy 21 mosaicism and Down syndrome, although there is likely variation in phenotype associated with the percentage of trisomic cells and their tissue-specificity¹⁵³⁻¹⁵⁵. A more recent study looked at this issue of somatic mosaicism in Timothy syndrome type 1 (TS-1), which is a rare disorder that affects multiple organ systems and has a high incidence of sudden death due to profound QT prolongation and resultant ventricular arrhythmias. All previously described cases of TS-1 are associated with a missense mutation in exon 8A (p.G406R) of the L-type calcium channel gene (Ca(v)1.2, *CACNA1C*). Most people reported in the literature represent highly affected people who present early in life with severe cardiac and neurological manifestations, but these authors found somatic mosaicism in people with TS-1 with less severe manifestations than the typical person with TS-1¹⁵⁶. There are therefore likely large ascertainment biases, given that people with much less expressive pheno-

types are likely not coming to anyone's attention. The implications of these findings with somatic mosaicism are that one cannot currently predict phenotype from genotype, particularly in the absence of any comprehensive characterization of which tissues are mutated in any one person. Also, putative 'de novo' mutations can instead represent cases of parental mosaicism (including in the germline), which could be revealed by careful genotyping of parental tissues other than peripheral blood lymphocytes. In fact, we are increasingly becoming aware of many instances of germline mosaicism, in which a mutation is not present or is present only at a very low level in the blood sample from a parent, but clearly must be in their germline, as they have two or more children with the same mutation that must therefore have originated through the parent's germline¹⁵⁷⁻¹⁷⁷. Clearly, we are truly ignorant concerning the extent of diversity brought about by somatic mosaicism, and it is therefore far too simplistic to assume that a single blood draw truly represents the entire genome of a human being, with anywhere from 25-100 trillion cells in their body divided up among multiple organs and other tissue systems. Of course, even the words "whole genome sequencing" are misleading, as there might very well be millions to trillions of similar (but not the exact same) genomes in each person's body.

Rare and compensatory mutations

There is an increasingly rich literature regarding rare mutations with seemingly large phenotypic effects¹⁷⁸⁻¹⁸¹. An example of this is Liam Hoekstra, known as the world's strongest toddler when he was age 3, and who has an extremely rare mutation in the gene encoding myostatin, leading to myostatin-related muscle hypertrophy with increased muscle mass and reduced body fat¹⁸². However, the effects of these mutations have mainly been reported in the context of particular genetic backgrounds, and so our knowledge of the expression of these mutations in the context of any number of genetic backgrounds is lacking. It is likely that there can be, and are, many genomic elements that

act in concert to influence these traits in a phenotypic spectrum. Of course, compensatory mutations can be explored in the context of other organisms¹⁸³⁻¹⁸⁵, but human migration and breeding is certainly not something that can be experimentally manipulated!

There are many disabling psychiatric syndromes, which have been lumped under certain artificial categories, such as schizophrenia, Tourette Syndrome (TS), obsessive compulsive disorder (OCD), and attention deficit hyperactivity disorder (ADHD). A very good way forward is to study these syndromes in large families living in the same geographic region, so as to control for

ancestry differences, minimize environmental influences, and focus on specific genotypes in these families. It is possible that a low number of genetic mutations will be shared in a relatively small combination (on the order of 1-3 such variants) among affected relatives within some pedigrees, and that these variants will not be present in the same combination in unaffected relatives or in other families with very little to no neuropsychiatric disorders^{23, 104, 186-190}. An alternative is that some affected people in these families have these illnesses due to additive and/or epistatic interactions among dozens to hundreds of loci within each person¹⁹¹⁻¹⁹³. The currently classified syndromes of schizophrenia, obsessive compulsive disorder (OCD), attention deficit hyperactivity disorder (ADHD), autism and other mental illnesses are quite heterogeneous within and between families, and these symptoms have also been observed in known single locus disorders such as Fragile X and 22q11.2 velocardi-ofacial syndrome^{104, 187}.

Some of these syndromes are referred to as ‘complex’ diseases simply because the presentation is so incredibly heterogeneous that it is very likely that there will be multiple different genetic and environmental explanations. One possible genetic explanation is that some symptoms of severe mental illness may emerge in a particular family due to a genetic constellation including dozens to hundreds of loci acting in each person either additively or via epistasis (and possibly modified by environment; G X E), which some refer to as the ‘polygenic’ model¹⁹¹⁻¹⁹⁴, as previously discussed.

If true, for predictive efforts in any particular family, the solution will ultimately require whole genome sequencing to tease out the numerous mutations involved. On the other hand, some discuss this concept of “many rare variants of large effect”, which they refer to as the ‘oligogenic’ model of inheritance^{195, 196}, as previously discussed. Some families have deleterious copy number variants^{187, 197-200}, and de novo single nucleotide mutations have recently been implicated as important for spontaneous ‘singleton’ cases in at least some families²⁰¹⁻²⁰⁶. There could also be a set of families with single, pair or triplet interactions among 1-3 gene mutations of high expression that can largely, on their own, contribute to a set of symptoms currently overlapping with named syndromes, such as ‘autism’ and ‘schizophrenia’²⁰⁷. As there is no way of really distinguishing between these two artificially created models in any one particular family, it is reasonable (with current costs) to perform whole genome sequencing as a comprehensive way to ascertain most of the relevant genetic variance in any particular family.

It is becoming generally accepted that at least 5% of the ‘autisms’ appear to be associated with various large copy number variants²⁰⁸. So, it is likely that a substantial portion of the heritability will be influenced by other types of mutations, with some evidence pointing to a role for ‘de novo’ mutations in singleton, uninherited cases of autism^{202-205, 209} and other evidence suggesting that there might be multiple genetic and environmental influences in each person¹⁹¹.

Current ability / approaches

There has been an explosive growth in exome and whole genome sequencing (WGS)²³ led, in part, by dramatic cost reductions. The same is true for genotyping microarrays, which are becoming increasingly denser with various markers while maintaining a relatively stable cost²¹⁰. With rapid advancements in sequencing technologies²¹¹ and improved haplotype-phasing^{212,213}, high-throughput sequencing (HTS) data on the genomes of a diverse number of species are being generated at an unprecedented rate. The development of bioinformatics tools for handling these data has been somewhat lagged in response, creating a gap be-

tween the massive data being generated, and the ability to fully exploit the biological content of these data. Many short read alignment software tools are now available, along with several single nucleotide variants (SNVs) and copy number variant (CNVs) calling algorithms²³. However, there is a paucity of methods that can simultaneously handle a large number of genetic variants and annotate their functional impacts (particularly for a human genome, which typically hosts >3 million variants), despite the fact that this is an important task in many sequencing applications. Functional interpretation of genetic variants therefore becomes one of

the major obstacles to connect sequencing data with biomedical researchers who are willing to embrace the sequencing technology.

In the medical world, WGS has since led to the discovery of the genetic basis of Miller Syndrome²¹⁴ and in another instance, it was used to investigate the genetic basis of Charcot-Marie-Tooth neuropathy²¹⁵, alongside a discussion of the ‘return of results’²¹⁶. In 2011, the diagnosis of a pair of twins with dopa (3,4-dihydroxyphenylalanine) responsive dystonia (DRD; OMIM #128230) and the discovery that they carried compound heterozygous mutations in the SPR gene encoding sepiapterin reductase led to supplementation of l-dopa therapy with 5-hydroxytryptophan, a serotonin precursor, resulting in clinical improvements in both twins²¹⁷.

Despite current technological limitations, mutations are continually being identified in research settings^{214, 218-222}. However, the human genomics community has recognized a number of distinct challenges, including with phenotyping, sample collection, sequencing strategies, bioinformatics analysis, biological validation of variant function, clinical interpretation and validity of variant data, and delivery of genomic information to various constituents^{23, 223}. In particular, there is a need for large pedigree sample collection, high-quality sequencing data acquisition, rigorous generation of variant calls, and comprehensive functional annotation of variants^{23, 224, 225}. Empirical estimates seem to suggest that exome sequencing can identify a putative disease variant in only about 10-50% of the cases for which it is applied²³, and the genetic architecture of most neuropsychiatric illness is still largely undefined and controversial^{104, 186, 191, 192}. The sequencing of entire genomes in large families will create a dataset that can be analysed and re-analysed for years to come as new biology and new methods emerge. The cost of a whole ge-

nome will likely decrease much more rapidly in relation to the cost of exome sequencing, given the relatively fixed labor and reagent costs for capturing the exons in the genome. Also, there is emerging evidence that exon capture and sequencing only achieves high depth of sequencing coverage in about 90% of the exons, whereas WGS does not involve a capture step and thus obtains better coverage on >95% of all exons in the genome. Of course, even the definition of the exome is a moving target, as the research community is constantly annotating and finding new exons not previously discovered^{226, 227}, and therefore WGS is a much more comprehensive way to assess coding and non-coding regions of the genome.

It is obvious that in both research and clinical settings WGS can dramatically impact clinical care, and it is now a matter of economics and feasibility in terms of WGS being adopted widely in a clinical setting^{23, 225}. There are, however, still many challenges in showing how any one mutation can contribute toward a clear phenotype, particularly in the context of genetic background and possible environmental influences²²⁸. Bioinformatics confounders, such as poor data quality²²⁹, sequence inaccuracy, and variation introduced by different methodological approaches²³⁰ can further complicate biological and genetic inferences. Furthermore, one cannot exclude polygenic and epistatic modes of inheritance^{90, 231-236}. To address these issues, future work will need to focus on evaluating next generation sequencing data coming from multiple sequencing and informatics platforms, and involving multiple other family members. By using a combination of data from many family members and from different sequencing technologies evaluated by a number of bioinformatics pipelines, we can maximize accuracy and thus the biological inference stemming from these data.

Prenatal diagnosis, preimplantation genetic diagnosis/screening

“Before a new function can arise, it may be essential for a lineage to evolve a potentiating genetic background that allows the actualizing mutation to occur or the new function to be expressed. Finally, novel functions often emerge in rudimentary forms that must be refined to exploit the ecological opportunities. This three-step process—in which potentiation makes a trait possible, actualization makes the trait manifest, and refinement makes it effective—is likely typical of many new functions.” – Richard Lenski⁸⁶

A great clinical geneticist, John Opitz, has observed the following: “More fetuses die prenatally than are born alive. Many die because of genetic conditions, malformations, and syndromes. Most are not autopsied, and in such cases appropriate genetic counseling is not provided or possible. In such ‘cases’ (fetuses, infants) a huge amount of genetic pathology is yet to be discovered (our last frontier!)”²³⁷.

In this regard, some have suggested a canalization model, which describes phenotypes as being robust to small perturbations, seemingly stuck within “phenotypic canals”. Phenotypes may ‘slosh’ against the sides of the canal during development, but with little effect on the final outcome of development²³⁸⁻²⁴⁰. In such a model, it is only perturbations with a magnitude exceeding a certain threshold that can direct the developmental path out of the canal (see Figure 1 for an illustrative model of canalization). Accordingly, phenotypes are robust up to a limit, with little robustness beyond this limit. This pattern may increase rates of evolution in fluctuating environments, as phenotypes are more likely to be perturbed with increased frequency and magnitude, thus leading to more rapid delineations and differentiations of canalized phenotypes.

One could argue that the birth of a child in one particular family with a clear phenotype, such as cystic fibrosis, along with previously identified associated mutations, dramatically increases the ‘prior probability’ that a future child with these same mutations being born in that same family would have a similar ‘canalized’ phenotype. It is really only in that particular situation in which one could make a somewhat informed prediction of genotype going down one particular phenotypic “canal”. And yet, a study in Australia from 2000-2004 showed that of the 82 children born with cystic fibrosis (CF) in Victoria, Australia, 5 (6%) were from families with a known history of CF.

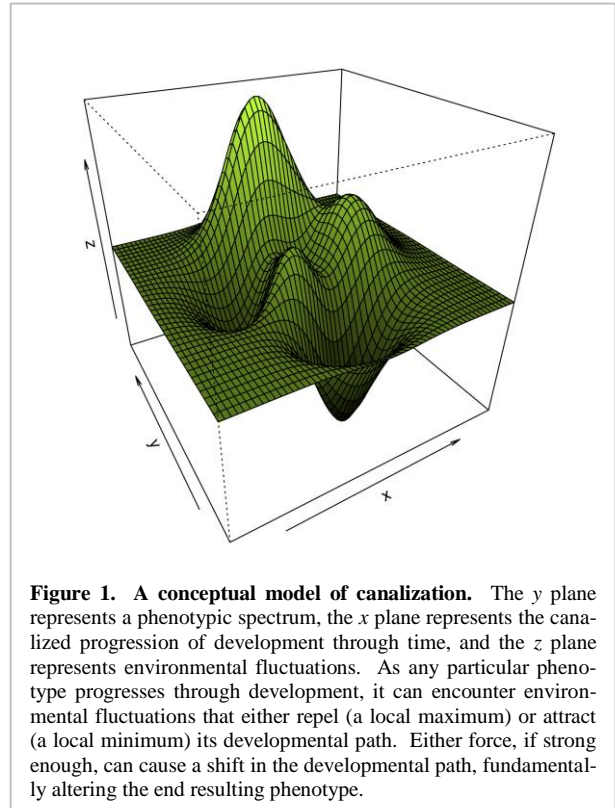


Figure 1. A conceptual model of canalization. The y plane represents a phenotypic spectrum, the x plane represents the canalized progression of development through time, and the z plane represents environmental fluctuations. As any particular phenotype progresses through development, it can encounter environmental fluctuations that either repel (a local maximum) or attract (a local minimum) its developmental path. Either force, if strong enough, can cause a shift in the developmental path, fundamentally altering the end resulting phenotype.

The authors found that “even when a family history is known, most relatives do not undertake carrier testing. In an audit of cascade carrier testing after a diagnosis of CF through newborn screening, only 11.8% of eligible (non-parent) (82/716) relatives were tested²⁴¹. These same researchers also showed that in a clinical setting, the diagnosis of a baby with CF by newborn screening “does not lead to carrier testing for the majority of the baby's non-parent relatives”²⁴. This is incredibly unfortunate, given that predictions of any reliability ought to include the prior probability of someone being born in that ‘ancestry group’ with the mutations and phenotype of interest.

Despite the above facts, non-invasive sequencing of fetal genomes is an area of intense interest in

genomic medicine, and a cynical person might argue that the rush to implement this technology is driven mainly by financial interests. This technique is based on the observation that a small proportion of the cell-free DNA in a pregnant woman's blood is derived from the fetus, so that aneuploidy or genomic sequence of a fetus may be inferred by sequencing of maternal plasma DNA and algorithmic decoupling of maternal and fetal DNA variants. A few companies are already marketing non-invasive prenatal screening (NIPS) tests for non-invasive detection of trisomy 21 associated with Down's syndrome²⁴². One can reasonably argue that detecting Down's syndrome is a conceptually and practically much simpler task than detecting individual variants within the fetal genome to assess mutations associated with disorders such as cystic fibrosis and hearing loss. However, with sufficiently high sequence depth, it is technically feasible to detect single nucleotide alterations in a fetal genome, as shown in several recent papers²⁴³⁻²⁴⁶. But, to allow accurate detection of individual vari-

ants, very high sequencing depth is required (potentially hundreds-fold higher than sequencing germline genomes); therefore, it is likely that targeted exon capture and sequencing might dominate the market until sufficiently high depth whole-genome sequencing becomes an economically feasible alternative. Given these technological developments, it is likely that some form of fetal genome testing will be available in the next few years. Others have noted that we might be reaching a point in the near-term future where it may be feasible to incorporate genetic, genomic and transcriptomic data to develop new approaches to fetal treatment^{247, 248}. One concern is that greed and financial conflicts of interest could lead to indiscriminate marketing and use of NIPS as diagnostic tests, rather than simply as screening, and that this technology will be implemented without any regard for genetic background or environmental differences, alongside a complete misunderstanding of this concept of extreme variability in phenotypic expression.

Implications for acceptance, prognosis and treatment

"When a complex system starts to dysfunction, it is generally best to fix it early. The alternative often means delaying until the system has degenerated into a disorganized, chaotic mess — at which point it may be beyond repair. Unfortunately, the general approach to cancer has ignored such common sense. The vast majority of cancer research is devoted to finding cures, rather than finding new ways to prevent disease" — Michael Sporn¹⁰⁸.

Prevention of illness through environmental modification has been, and likely always will be, the major driver for global health^{108, 110}. With this in mind, the sequencing of whole genomes on a large scale promises to enable the discovery and prediction of disease in some people. The ability to sequence an infant at birth and to be able to predict a higher probability of certain phenotypes, such as developmental delay, would allow for educational and behavioral interventions to influence the phenotype, thus altering the trajectory of that phenotype²⁴⁹⁻²⁵⁴. One recent study of chromosomal microarray (CMA) testing found that "among 1792 patients with developmental delay (DD), intellectual disability (ID), multiple congenital anomalies (MCA), and/or autism spectrum disorders (ASD), 13.1% had clinically relevant results, either ab-

normal (n = 131; 7.3%) or variants of possible significance (VPS; n = 104; 5.8%). Abnormal variants generated a higher rate of recommendation for clinical action (54%) compared with VPS (34%; Fisher exact test, P = 0.01)²⁵⁵. The authors concluded that "CMA results influenced medical management in a majority of patients with abnormal variants and a substantial proportion of those with VPS" thus supporting the use of CMA in this population²⁵⁵.

However, there are some major barriers to the widespread implementation of genomic medicine in the clinic. These include:

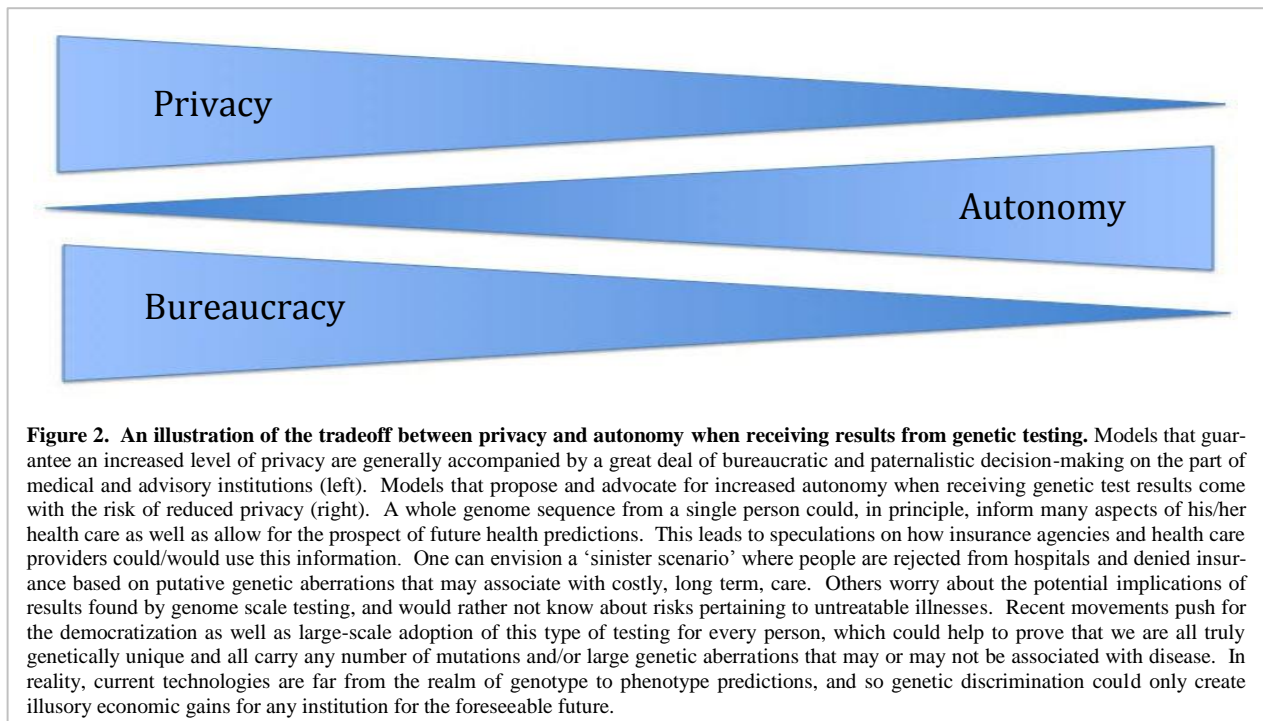
- 1) Lack of public education
- 2) Lack of physician knowledge about genetics

- 3) Apathy on the part of the populace in terms of preventive efforts
- 4) Refusal of insurance companies and governments to pay for genetic testing
- 5) Focus in our society on treatment, not on early diagnosis and prevention
- 6) Privacy concerns
- 7) Limits of our current knowledge

The emphasis should be on diagnosis and prevention, not just on treatment. During the medical training of one of the authors (GJL), two episodes helped to illustrate this. The first involved a 15-year old girl with Type I diabetes, who was hospitalized dozens of times with diabetic ketoacidosis. Literally hundreds of thousands of dollars were spent to repeatedly save her life, but very little time or money was spent on therapy or education to teach her about taking her insulin and ensuring that she did. Unfortunately, in America at least, this is due to a relative lack of reimbursement for such activities, whereas saving someone already in diabetic ketoacidosis is quite lucrative to everyone involved. A second episode involved a 14-year old boy, who had been hospitalized well over 10 times with acute pancreatitis over a ten year period, with very little thought concerning why he had recurring pancreatitis. Finally, someone obtained a genetics consult, and they recommended cystic fibrosis (CF) genetic screening, which had never been ordered before due to a prior ‘negative’ sweat test. It turns out that this boy had two rare mutations in *CFTR*, undiagnosed till then, which had been contributing to recurrent pancreatitis. He had never had any lung manifestations, and he had never had a positive sweat test for CF, mainly due to the fact that these mutations appeared to only be exerting effects in his pancreas, not in his skin or lungs. After this diagnosis, this person benefited from pancreatic enzyme supplementation, along with therapy and education. Once again, the reason it took so long to diagnose this person is because the incentive structure in America is not on early diagnosis and prevention, but rather on treatment of people only once they become severely ill^{256, 257}. This is illustrated by the fact that there are only about ~1000 medical geneticists in America and ~3000 genetic counselors, for a population of ~315 million, which certainly makes it basically impossible for these limited number of professionals to implement genomic medicine in any meaningful

way²⁵⁸. The numbers of such health care professionals are even smaller in developing regions of the world, thus making it currently very difficult to provide widespread genetic counseling^{71, 259, 260}. Stepping into this void are direct-to-consumer for-profit genetic testing companies, and this is certainly one disruptive way of trying to help people manage their genetic results online^{261, 262}, although financial motives and lack of transparency can create problems²⁶³.

Privacy concerns have added to the difficulties of implementing genomics-guided medicine. Genetic data have the potential of being informative across a wide variety of human traits and health conditions, and some worry about the potential misuse of these data by insurance agencies as well as by health care providers²⁶⁴. Genetic testing has historically been focused on targeting and examining a small number of known genetic aberrations²⁶⁵; however, since the advent of high-throughput sequencing technologies, the landscape is starting to change. With the emergence of tests that can target and examine all coding regions of the genome, or even the genome in its entirety²⁶⁶, testing can now be performed on a more global and exploratory scale. Some people worry about returning the results of such a test, whose findings can have questionable clinical significance, and in response have advocated for selectively restricting the returnable medical content. Others have proposed complicated anonymization techniques that could allow for a safe return of research results to participants whose genome is suspect to contain ‘clinically actionable’ information. One such proposition involves the cryptographic transformation of genomic data in which only by the coalescence of keys held by many different intermediate parties would the identity of the participant be revealed, and only in cases where all parties agree that there is indeed the presence of clinically actionable information²⁶⁷. These types of recommendations take a more paternalistic approach in returning test results to people, and generally involve a deciding body of people that can range in size from a single medical practitioner to a committee of experts. In contrast, there is a growing movement among the populace to learn more about their own ‘personalized’ health and health care. There has also been a renewed push for the unfiltered sharing and networking of health related data, which has been facilitated and hastened by the explosion of digitally



mediated social networking over the past decade, as well as by private institutions such as 23andMe²⁶⁸ and the Personal Genomes Project²⁶⁹ that aim to popularize and democratize genetic testing. Clearly, between these contrasting approaches, there is a tradeoff between the privacy and personal safety one can expect to retain by either freely acquiring and sharing the full breadth of one’s genetic testing data, or by allowing deciding bodies to choose what information you will receive.

Public databases containing human sequence data have grown in magnitude and in number, and relatively comprehensive sequencing data have already been generated and published on thousands of people^{270, 271}. Similar privacy concerns have since been expressed about the degree of medical and personal privacy that these and other research participants can expect²⁷², given that each person is genetically unique. As a demonstration of current vulnerabilities, researchers have shown that the identities of participants can be discovered using these publicly available data²⁷³. Although these data have been instrumental in furthering our understanding of human genetics, medicine, and biological processes in general, some advocate for caution when sharing and publishing human genetic sequence information²⁷⁴.

As the cost and difficulty of sequencing continually decreases, a wealth of data are becoming

available to researchers, privately funded institutions and individual consumers. More people are willing to share a larger portion of their personal life in the public arena, and we fully expect that, given the popularization of ‘personalized’ genomic health related data, more people will want to share these data and offer their own DNA sequence for others to explore. There is a trade off between the risks inherent in sharing vast quantities of health data, and maintaining personal privacy in the burgeoning age of personalized medicine and genomics. As the technology and science mature, our power to interpret and use these health data for practical and preventative measures will certainly improve. Conventions for privacy and autonomy will likely be driven by popular demand, and could vary from person to person, as all people differ in their desire for privacy and autonomy (see Figure 2 for a conceptual model of this tradeoff).

In addition, within the current paradigm of genetic determinism, which stretches back to the time of William Bateson^{275, 276}, some people would have us believe that variants can and should be binned into different classes based on clinical utility and validity²⁷⁷⁻²⁷⁹, without any obvious regard to genetic background or environmental differences. Environment and ancestry matter^{2, 3, 275, 276}, and yet some clinical geneticists trained in the current paradigm of genetic determinism clearly do

not wish to acknowledge this. In fact, one medical academy in America recently released guidelines in which they recommended the “return of secondary findings” for only 57 genes, without any real guidance for the rest of the genome or environmental influences²⁸⁰. This is therefore a very conservative set of recommendations, given that there are approximately 20,000 protein-coding genes in the human genome, along with the thousands of other identified, important noncoding elements of the genome^{9, 28-42}! As stated above, but worth repeating, there are 6 billion nucleotides of DNA in every cell of the human body, and there are 25-100 trillion cells in each human body. Given genetic modifiers, somatic mosaicism, epigenetic changes, and environmental differences, no two human beings are the same, and therefore the expression of any mutation will be different in each person. At best, phenotypes will follow canalized pathways in

direct relatives, such as mother and child, so the analysis of mutations over several generations in the same families is a worthwhile effort. But, how we will ever get to a world of millions of whole genomes shared and analyzed for numerous additive, epistatic interactions and gene by environment interactions, so that we can make any reliable predictions for any one human being, if we are only recommending ‘return of results’ from ~57 genes? We need to sequence and collate the raw data from thousands and then millions of exomes and genomes, so that we can actually begin to really understand the expression patterns of any mutation in the human genome in particular families. In medicine, people tend to create illusions of certainty, when in fact everything is probabilistic¹². Some humans like to be told things in a ‘yes/no’ manner, but there always exists a degree of unresolvable uncertainty.

Conclusions

“A new scientific truth does not triumph by convincing its opponents and making them see the light, but rather because its opponents eventually die, and a new generation grows up that is familiar with it.” -- Max Planck

With the advent of exome and whole genome sequencing, we need to focus again on families over several generations, so as to attempt to minimize genetic differences, locus heterogeneity and environmental influences. Forging strong ties with families will also enable access to other tissues to continue to study newly discovered loci with many emerging technologies. Some might consider it to be ‘social activism’ to advocate for a more comprehensive collection and collation of human pedigrees and whole genome sequencing data. But, in the words of one author: “Scientists, whether we like it or not, are members of society, and we are prone to the ideas and beliefs of the times in which we live²⁸¹.” We currently live within a paradigm of genetic determinism, but we should not be for-

ever condemned to this simplistic mode of thinking. One can imagine that in 100 years’ time, each person will be able to keep track of detailed longitudinal phenotyping data on themselves, and they will be able to link this to records of their relatives, both living and deceased. One can hope that in 100 to 200 years’ time, there might then be sufficient information available within many large families to begin to really calculate probabilistic outcomes¹²⁻¹⁶ and then attempt to alter the trajectory for many diseases. One can see this beginning already to occur in certain geographically isolated clans, such as in Iceland^{178, 179}, so there is some optimism that this can indeed occur on a global level, including in the currently less developed regions of the world²⁶⁰.

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References

1. Seguin E: *Idiocy : and its treatment by the physiological method*. New York: William Wood & Co.; 1866.
2. Weldon WFR: Mendel's laws of alternative inheritance in peas. *Biometrika* 1902, 1:228-254.
3. Radick G: Physics in the Galtonian sciences of heredity. *Studies in history and philosophy of biological and biomedical sciences* 2011, 42:129-138.
4. Bearn AG: *Archibald Garrod and the individuality of Man*. Oxford, New York: Clarendon Press; Oxford University Press; 1993.
5. Comfort NC: *The science of human perfection : how genes became the heart of American medicine*. New Haven: Yale University Press; 2012.
6. Tennesen JA, Bigham AW, O'Connor TD, Fu W, Kenny EE, Gravel S, McGee S, Do R, Liu X, Jun G, et al: Evolution and Functional Impact of Rare Coding Variation from Deep Sequencing of Human Exomes. *Science* 2012.
7. Grillo E, Lo Rizzo C, Bianciardi L, Bizzarri V, Baldassarri M, Spiga O, Furini S, De Felice C, Signorini C, Leoncini S, et al: Revealing the complexity of a monogenic disease: rett syndrome exome sequencing. *PLoS one* 2013, 8:e56599.
8. Misteli T: The cell biology of genomes: bringing the double helix to life. *Cell* 2013, 152:1209-1212.
9. Sabin LR, Delas MJ, Hannon GJ: Dogma derailed: the many influences of RNA on the genome. *Molecular cell* 2013, 49:783-794.
10. Sriver CR: The PAH gene, phenylketonuria, and a paradigm shift. *Hum Mutat* 2007, 28:831-845.
11. Allchin D: The dilemma of dominance. *Biology & Philosophy* 2005, 20:427-451.
12. Gigerenzer G: *Calculated risks : how to know when numbers deceive you*. New York: Simon & Schuster; 2002.
13. Sokal RR: *Biometry : the principles and practice of statistics in biological research*. [Extensively rev.] 4th edn. New York: W.H. Freeman; 2012.
14. Gigerenzer G, Galesic M: Why do single event probabilities confuse patients? *BMJ* 2012, 344:e245.
15. Gigerenzer G, Wegwarth O, Feufel M: Misleading communication of risk. *BMJ* 2010, 341:c4830.
16. Kurz-Milcke E, Gigerenzer G, Martignon L: Transparency in risk communication: graphical and analog tools. *Annals of the New York Academy of Sciences* 2008, 1128:18-28.
17. Freund J, Brandmaier AM, Lewejohann L, Kirste I, Kritzler M, Kruger A, Sachser N, Lindenberger U, Kempermann G: Emergence of individuality in genetically identical mice. *Science* 2013, 340:756-759.
18. Bateson W, Mendel G: *Mendel's principles of heredity a defense*. Cambridge Eng.: University Press; 1902.
19. Olby R: The dimensions of scientific controversy: the biometric--Mendelian debate. *British journal for the history of science* 1989, 22:299-320.
20. Provine WB: *The origins of theoretical population genetics*. 2nd edn. Chicago: University of Chicago Press; 2001.
21. Nielsen MA: *Reinventing discovery : the new era of networked science*. Princeton, N.J.: Princeton University Press; 2012.
22. Lyon GJ, Segal JP: Practical, ethical and regulatory considerations for the evolving medical and research genomics landscape. *Applied & Translational Genomics* 2013.
23. Lyon GJ, Wang K: Identifying disease mutations in genomic medicine settings: current challenges and how to accelerate progress. *Genome medicine* 2012, 4:58.
24. McClaren BJ, Metcalfe SA, Aitken M, Massie RJ, Ukoumunne OC, Amor DJ: Uptake of carrier testing in families after cystic fibrosis diagnosis through newborn screening. *European journal of human genetics : EJHG* 2010, 18:1084-1089.
25. Keller EF: *The mirage of a space between nature and nurture*. Durham N.C.: Duke University Press; 2010.
26. Pennacchio LA, Bickmore W, Dean A, Nobrega MA, Bejerano G: Enhancers: five essential questions. *Nature reviews Genetics* 2013, 14:288-295.

27. Dickel DE, Visel A, Pennacchio LA: Functional anatomy of distant-acting mammalian enhancers. *Philosophical transactions of the Royal Society of London Series B, Biological sciences* 2013, 368:20120359.
28. Cartault F, Munier P, Benko E, Desguerre I, Hanein S, Boddaert N, Bandiera S, Vellayoudom J, Krejbich-Trotot P, Bintner M, et al: Mutation in a primate-conserved retrotransposon reveals a noncoding RNA as a mediator of infantile encephalopathy. *Proceedings of the National Academy of Sciences of the United States of America* 2012, 109:4980-4985.
29. Salzman J, Gawad C, Wang PL, Lacayo N, Brown PO: Circular RNAs are the predominant transcript isoform from hundreds of human genes in diverse cell types. *PLoS one* 2012, 7:e30733.
30. Batista PJ, Chang HY: Long noncoding RNAs: cellular address codes in development and disease. *Cell* 2013, 152:1298-1307.
31. Hansen TB, Jensen TI, Clausen BH, Bramsen JB, Finsen B, Damgaard CK, Kjems J: Natural RNA circles function as efficient microRNA sponges. *Nature* 2013, 495:384-388.
32. Kapusta A, Kronenberg Z, Lynch VJ, Zhuo X, Ramsay L, Bourque G, Yandell M, Feschotte C: Transposable Elements Are Major Contributors to the Origin, Diversification, and Regulation of Vertebrate Long Noncoding RNAs. *PLoS Genet* 2013, 9:e1003470.
33. Khoddami V, Cairns BR: Identification of direct targets and modified bases of RNA cytosine methyltransferases. *Nature biotechnology* 2013.
34. Ledford H: Circular RNAs throw genetics for a loop. *Nature* 2013, 494:415.
35. Maxmen A: RNA: The genome's rising stars. *Nature* 2013, 496:127-129.
36. Memczak S, Jens M, Elefsinioti A, Torti F, Krueger J, Rybak A, Maier L, Mackowiak SD, Gregersen LH, Munschauer M, et al: Circular RNAs are a large class of animal RNAs with regulatory potency. *Nature* 2013, 495:333-338.
37. Mercer TR, Mattick JS: Structure and function of long noncoding RNAs in epigenetic regulation. *Nature structural & molecular biology* 2013, 20:300-307.
38. Miura P, Shenker S, Andreu-Agullo C, Westholm JO, Lai EC: Widespread and extensive lengthening of 3' UTRs in the mammalian brain. *Genome Res* 2013, 23:812-825.
39. Moreau MP, Bruse SE, Jornsten R, Liu Y, Brzustowicz LM: Chronological Changes in MicroRNA Expression in the Developing Human Brain. *PLoS one* 2013, 8:e60480.
40. Ning S, Wang P, Ye J, Li X, Li R, Zhao Z, Huo X, Wang L, Li F: A global map for dissecting phenotypic variants in human lincRNAs. *European journal of human genetics : EJHG* 2013.
41. Perrat PN, DasGupta S, Wang J, Theurkauf W, Weng Z, Rosbash M, Waddell S: Transposition-driven genomic heterogeneity in the Drosophila brain. *Science* 2013, 340:91-95.
42. Wilusz JE, Sharp PA: Molecular biology. A circuitous route to noncoding RNA. *Science* 2013, 340:440-441.
43. Down JL: Observations on an ethnic classification of idiots. 1866. *Mental retardation* 1995, 33:54-56.
44. Hume D, Selby-Bigge LA: *A treatise of human nature*. Oxford,: Clarendon press; 1896.
45. causality [<http://en.wikipedia.org/wiki/Causality>]
46. Corleto VD, Gambardella S, Gullotta F, D'Apice MR, Piciucchi M, Galli E, Lucidi V, Novelli G, Delle Fave G: New PRSS1 and common CFTR mutations in a child with acute recurrent pancreatitis, could be considered an "Hereditary" form of pancreatitis ? *BMC gastroenterology* 2010, 10:119.
47. Derikx MH, Drenth JP: Genetic factors in chronic pancreatitis; implications for diagnosis, management and prognosis. *Best practice & research Clinical gastroenterology* 2010, 24:251-270.
48. Rosendahl J, Landt O, Bernadova J, Kovacs P, Teich N, Bodeker H, Keim V, Ruffert C, Mossner J, Kage A, et al: CFTR, SPINK1, CTRC and PRSS1 variants in chronic pancreatitis: is the role of mutated CFTR overestimated? *Gut* 2013, 62:582-592.
49. Emond MJ, Louie T, Emerson J, Zhao W, Mathias RA, Knowles MR, Wright FA, Rieder MJ, Tabor HK, Nickerson DA, et al: Exome sequencing of extreme phenotypes identifies DCTN4 as a modifier of chronic Pseudomonas aeruginosa infection in cystic fibrosis. *Nature genetics* 2012, 44:886-889.
50. Orr HT, Zoghbi HY: Trinucleotide repeat disorders. *Annual review of neuroscience* 2007, 30:575-621.
51. Tome S, Manley K, Simard JP, Clark GW, Slean MM, Swami M, Shelbourne PF, Tillier ER, Monckton DG, Messer A, Pearson CE: MSH3 polymorphisms and protein levels affect CAG repeat instability in Huntington's disease mice. *PLoS genetics* 2013, 9:e1003280.
52. Ciancarelli I, Tozzi Ciancarelli MG, Carolei A: Effectiveness of intensive neurorehabilitation in patients with Huntington's disease. *European journal of physical and rehabilitation medicine* 2013.

53. Li H, Cherry S, Klinedinst D, DeLeon V, Redig J, Reshey B, Chin MT, Sherman SL, Maslen CL, Reeves RH: Genetic modifiers predisposing to congenital heart disease in the sensitized Down syndrome population. *Circulation Cardiovascular genetics* 2012, 5:301-308.
54. Ackerman C, Locke AE, Feingold E, Reshey B, Espana K, Thusberg J, Mooney S, Bean LJ, Dooley KJ, Cua CL, et al: An excess of deleterious variants in VEGF-A pathway genes in Down-syndrome-associated atrioventricular septal defects. *American journal of human genetics* 2012, 91:646-659.
55. Papavassiliou P, York TP, Gursoy N, Hill G, Nicely LV, Sundaram U, McClain A, Aggen SH, Eaves L, Riley B, Jackson-Cook C: The phenotype of persons having mosaicism for trisomy 21/Down syndrome reflects the percentage of trisomic cells present in different tissues. *American journal of medical genetics Part A* 2009, 149A:573-583.
56. Solomon A: *Far from the tree : parents, children and the search for identity*. 1st Scribner hardcover edn. New York: Scribner; 2012.
57. Dodd KJ, Shields N: A systematic review of the outcomes of cardiovascular exercise programs for people with Down syndrome. *Archives of physical medicine and rehabilitation* 2005, 86:2051-2058.
58. Wang L, Kloc M, Gu Y, Ge S, Maffei A: Layer-specific experience-dependent rewiring of thalamocortical circuits. *The Journal of neuroscience : the official journal of the Society for Neuroscience* 2013, 33:4181-4191.
59. Maffei A, Bucher D, Fontanini A: Homeostatic plasticity in the nervous system. *Neural Plast* 2012, 2012:913472.
60. Maffei A: Enriching the environment to disinhibit the brain and improve cognition. *Front Cell Neurosci* 2012, 6:53.
61. Maffei A, Turrigiano G: The age of plasticity: developmental regulation of synaptic plasticity in neocortical microcircuits. *Progress in brain research* 2008, 169:211-223.
62. Guris DL, Duester G, Papaioannou VE, Imamoto A: Dose-dependent interaction of Tbx1 and Crkl and locally aberrant RA signaling in a model of del22q11 syndrome. *Developmental cell* 2006, 10:81-92.
63. Iascone MR, Vittorini S, Sacchelli M, Spadoni I, Simi P, Giusti S: Molecular characterization of 22q11 deletion in a three-generation family with maternal transmission. *American journal of medical genetics* 2002, 108:319-321.
64. Liao J, Kochilas L, Nowotschin S, Arnold JS, Aggarwal VS, Epstein JA, Brown MC, Adams J, Morrow BE: Full spectrum of malformations in velo-cardio-facial syndrome/DiGeorge syndrome mouse models by altering Tbx1 dosage. *Human molecular genetics* 2004, 13:1577-1585.
65. McDonald-McGinn DM, Fahiminiya S, Revil T, Nowakowska BA, Suhl J, Bailey A, Mlynarski E, Lynch DR, Yan AC, Bilaniuk LT, et al: Hemizygous mutations in SNAP29 unmask autosomal recessive conditions and contribute to atypical findings in patients with 22q11.2DS. *Journal of medical genetics* 2013, 50:80-90.
66. Stalmans I, Lambrechts D, De Smet F, Jansen S, Wang J, Maity S, Kneer P, von der Ohe M, Swillen A, Maes C, et al: VEGF: a modifier of the del22q11 (DiGeorge) syndrome? *Nature medicine* 2003, 9:173-182.
67. Moreno-De-Luca A, Myers SM, Challman TD, Moreno-De-Luca D, Evans DW, Ledbetter DH: Developmental brain dysfunction: revival and expansion of old concepts based on new genetic evidence. *Lancet neurology* 2013, 12:406-414.
68. Zimmermann MB: The effects of iodine deficiency in pregnancy and infancy. *Paediatric and perinatal epidemiology* 2012, 26 Suppl 1:108-117.
69. Walter Frank Raphael Weldon [http://en.wikipedia.org/wiki/Walter_Frank_Raphael_Weldon]
70. Jamieson A, Radick G: Putting Mendel in his place: How curriculum reform in genetics and counterfactual history of science can work together. . In *THE PHILOSOPHY OF BIOLOGY: A COMPANION FOR EDUCATORS*. Edited by Kampourakis K. Dordrecht: Springer; 2013: 577-595
71. Bittles AH, Black ML: Evolution in health and medicine Sackler colloquium: Consanguinity, human evolution, and complex diseases. *Proceedings of the National Academy of Sciences of the United States of America* 2010, 107 Suppl 1:1779-1786.
72. Lupski JR, Belmont JW, Boerwinkle E, Gibbs RA: Clan genomics and the complex architecture of human disease. *Cell* 2011, 147:32-43.
73. Dolinoy DC, Weidman JR, Waterland RA, Jirtle RL: Maternal genistein alters coat color and protects Avy mouse offspring from obesity by modifying the fetal epigenome. *Environmental health perspectives* 2006, 114:567-572.
74. Bernal AJ, Jirtle RL: Epigenomic disruption: the effects of early developmental exposures. *Birth defects research Part A, Clinical and molecular teratology* 2010, 88:938-944.

75. Weinhouse C, Anderson OS, Jones TR, Kim J, Liberman SA, Nahar MS, Rozek LS, Jirtle RL, Dolinoy DC: An expression microarray approach for the identification of metastable epialleles in the mouse genome. *Epigenetics : official journal of the DNA Methylation Society* 2011, 6:1105-1113.
76. Casanueva MO, Burga A, Lehner B: Fitness trade-offs and environmentally induced mutation buffering in isogenic *C. elegans*. *Science* 2012, 335:82-85.
77. Burga A, Casanueva MO, Lehner B: Predicting mutation outcome from early stochastic variation in genetic interaction partners. *Nature* 2011, 480:250-253.
78. Comfort NC: *The tangled field : Barbara McClintock's search for the patterns of genetic control*. Cambridge, Mass.: Harvard University Press; 2001.
79. Beaudet AL: The utility of chromosomal microarray analysis in developmental and behavioral pediatrics. *Child development* 2013, 84:121-132.
80. Holmes FL, Summers WC: *Reconceiving the gene : Seymour Benzer's adventures in phage genetics*. New Haven: Yale University Press; 2006.
81. Greenspan RJ: Biological indeterminacy. *Science and engineering ethics* 2012, 18:447-452.
82. Greenspan RJ: Selection, gene interaction, and flexible gene networks. *Cold Spring Harbor symposia on quantitative biology* 2009, 74:131-138.
83. Greenspan RJ: Seymour Benzer (1921-2007). *Current biology : CB* 2008, 18:R106-110.
84. Kendler KS, Greenspan RJ: The nature of genetic influences on behavior: lessons from "simpler" organisms. *The American journal of psychiatry* 2006, 163:1683-1694.
85. van Swinderen B, Greenspan RJ: Flexibility in a gene network affecting a simple behavior in *Drosophila melanogaster*. *Genetics* 2005, 169:2151-2163.
86. Blount ZD, Barrick JE, Davidson CJ, Lenski RE: Genomic analysis of a key innovation in an experimental *Escherichia coli* population. *Nature* 2012, 489:513-518.
87. Meyer JR, Dobias DT, Weitz JS, Barrick JE, Quick RT, Lenski RE: Repeatability and contingency in the evolution of a key innovation in phage lambda. *Science* 2012, 335:428-432.
88. Kohane IS, Hsing M, Kong SW: Taxonomizing, sizing, and overcoming the incidentalome. *Genetics in medicine : official journal of the American College of Medical Genetics* 2012.
89. Eichers ER, Lewis RA, Katsanis N, Lupski JR: Triallelic inheritance: a bridge between Mendelian and multifactorial traits. *Annals of medicine* 2004, 36:262-272.
90. Katsanis N, Ansley SJ, Badano JL, Eichers ER, Lewis RA, Hoskins BE, Scambler PJ, Davidson WS, Beales PL, Lupski JR: Triallelic inheritance in Bardet-Biedl syndrome, a Mendelian recessive disorder. *Science* 2001, 293:2256-2259.
91. Katsanis N, Eichers ER, Ansley SJ, Lewis RA, Kayserili H, Hoskins BE, Scambler PJ, Beales PL, Lupski JR: BBS4 is a minor contributor to Bardet-Biedl syndrome and may also participate in triallelic inheritance. *American journal of human genetics* 2002, 71:22-29.
92. Abu-Safieh L, Al-Anazi S, Al-Abdi L, Hashem M, Alkuraya H, Alamr M, Sirelkhatim MO, Al-Hassnan Z, Alkuraya B, Mohamed JY, et al: In search of triallelism in Bardet-Biedl syndrome. *European journal of human genetics : EJHG* 2012.
93. Smaoui N, Chaabouni M, Sergeev YV, Kallel H, Li S, Mahfoudh N, Maazoul F, Kammoun H, Gandoura N, Bouaziz A, et al: Screening of the eight BBS genes in Tunisian families: no evidence of triallelism. *Investigative ophthalmology & visual science* 2006, 47:3487-3495.
94. Laurier V, Stoetzel C, Muller J, Thibault C, Corbani S, Jalkh N, Salem N, Chouery E, Poch O, Licaire S, et al: Pitfalls of homozygosity mapping: an extended consanguineous Bardet-Biedl syndrome family with two mutant genes (BBS2, BBS10), three mutations, but no triallelism. *European journal of human genetics : EJHG* 2006, 14:1195-1203.
95. Nakane T, Biesecker LG: No evidence for triallelic inheritance of MKKS/BBS loci in Amish Mckusick-Kaufman syndrome. *American journal of medical genetics Part A* 2005, 138:32-34.
96. Mykytyn K, Nishimura DY, Searby CC, Beck G, Bugge K, Haines HL, Cornier AS, Cox GF, Fulton AB, Carmi R, et al: Evaluation of complex inheritance involving the most common Bardet-Biedl syndrome locus (BBS1). *American journal of human genetics* 2003, 72:429-437.
97. Beales PL, Badano JL, Ross AJ, Ansley SJ, Hoskins BE, Kirsten B, Mein CA, Froguel P, Scambler PJ, Lewis RA, et al: Genetic interaction of BBS1 mutations with alleles at other BBS loci can result in non-Mendelian Bardet-Biedl syndrome. *American journal of human genetics* 2003, 72:1187-1199.
98. Philip N, Bassett A: Cognitive, behavioural and psychiatric phenotype in 22q11.2 deletion syndrome. *Behavior Genetics* 2011, 41:403-412.

99. Hamshere ML, Bennett P, Williams N, Segurado R, Cardno A, Norton N, Lambert D, Williams H, Kirov G, Corvin A, et al: Genomewide linkage scan in schizoaffective disorder: significant evidence for linkage at 1q42 close to DISC1, and suggestive evidence at 22q11 and 19p13. *Archives of general psychiatry* 2005, 62:1081-1088.
100. Blackwood DH, Fordyce A, Walker MT, St Clair DM, Porteous DJ, Muir WJ: Schizophrenia and affective disorders--cosegregation with a translocation at chromosome 1q42 that directly disrupts brain-expressed genes: clinical and P300 findings in a family. *American journal of human genetics* 2001, 69:428-433.
101. Berndt SI, Gustafsson S, Magi R, Ganna A, Wheeler E, Feitosa MF, Justice AE, Monda KL, Croteau-Chonka DC, Day FR, et al: Genome-wide meta-analysis identifies 11 new loci for anthropometric traits and provides insights into genetic architecture. *Nature genetics* 2013, 45:501-512.
102. Visscher PM, McEvoy B, Yang J: From Galton to GWAS: quantitative genetics of human height. *Genetics research* 2010, 92:371-379.
103. Visscher PM, Goddard ME, Derks EM, Wray NR: Evidence-based psychiatric genetics, AKA the false dichotomy between common and rare variant hypotheses. *Molecular psychiatry* 2012, 17:474-485.
104. Mitchell KJ: What is complex about complex disorders? *Genome biology* 2012, 13:237.
105. Gibson G: Rare and common variants: twenty arguments. *Nature reviews Genetics* 2011, 13:135-145.
106. Bleuler E: *Dementia praecox; or, The group of schizophrenias*. New York,: International Universities Press; 1958.
107. Kraepelin E: *Manic-depressive insanity and paranoia*. Edinburgh,: Livingstone; 1921.
108. Sporn MB: Perspective: The big C - for Chemoprevention. *Nature* 2011, 471:S10-11.
109. Vogelstein B, Papadopoulos N, Velculescu VE, Zhou S, Diaz LA, Jr., Kinzler KW: Cancer genome landscapes. *Science* 2013, 339:1546-1558.
110. Mukherjee S: *The emperor of all maladies : a biography of cancer*. 1st Scribner hardcover edn. New York: Scribner; 2010.
111. Lyon GJ, Coffey BJ: Complex tics and complex management in a case of severe Tourette's disorder (TD) in an adolescent. *Journal of child and adolescent psychopharmacology* 2009, 19:469-474.
112. Lyon GJ, Coffey B, Silva R: Posttraumatic stress disorder and reactive attachment disorder: outcome in an adolescent. *Journal of child and adolescent psychopharmacology* 2008, 18:641-646.
113. Lyon GJ: Possible varenicline-induced paranoia and irritability in a patient with major depressive disorder, borderline personality disorder, and methamphetamine abuse in remission. *Journal of clinical psychopharmacology* 2008, 28:720-721.
114. Sacks OW: *An anthropologist on Mars : seven paradoxical tales*. 1st edn. New York: Alfrd A. Knopf; 1995.
115. Sacks OW: *The man who mistook his wife for a hat and other clinical tales*. 1st Touchstone edn. New York, NY: Simon & Schuster; 1998.
116. Luria AR: *The man with a shattered world; the history of a brain wound*. New York,: Basic Books; 1972.
117. Luria AR: *The mind of a mnemonist : a little book about a vast memory*. Chicago: H. Regnery; 1976.
118. Van Horn JD, Irimia A, Torgerson CM, Chambers MC, Kikinis R, Toga AW: Mapping connectivity damage in the case of Phineas Gage. *PloS one* 2012, 7:e37454.
119. Ratiu P, Talos IF, Haker S, Lieberman D, Everett P: The tale of Phineas Gage, digitally remastered. *Journal of neurotrauma* 2004, 21:637-643.
120. Eichenbaum H: What H.M. taught us. *J Cogn Neurosci* 2013, 25:14-21.
121. Worthey EA, Mayer AN, Syverson GD, Helbling D, Bonacci BB, Decker B, Serpe JM, Dasu T, Tschannen MR, Veith RL, et al: Making a definitive diagnosis: successful clinical application of whole exome sequencing in a child with intractable inflammatory bowel disease. *Genet Med* 2011, 13:255-262.
122. Penrose LS: *The biology of mental defect*. [3d rev. and reset edn. London: Sidgwick and Jackson; 1963.
123. Ward OC: *John Langdon Down, 1828-1896 : a caring pioneer*. London ; New York, NY: Royal Society of Medicine Press; 1998.
124. Lyon GJ, Abi-Dargham A, Moore H, Lieberman JA, Javitch JA, Sulzer D: Presynaptic regulation of dopamine transmission in schizophrenia. *Schizophrenia bulletin* 2011, 37:108-117.
125. Craddock N, Owen MJ: The Kraepelinian dichotomy - going, going... but still not gone. *The British journal of psychiatry : the journal of mental science* 2010, 196:92-95.
126. Williams HJ, Craddock N, Russo G, Hamshere ML, Moskvina V, Dwyer S, Smith RL, Green E, Grozeva D, Holmans P, et al: Most genome-wide significant susceptibility loci for schizophrenia and bipolar disorder reported to date cross-traditional diagnostic boundaries. *Human molecular genetics* 2011, 20:387-391.

127. Lichtenstein P, Yip BH, Bjork C, Pawitan Y, Cannon TD, Sullivan PF, Hultman CM: Common genetic determinants of schizophrenia and bipolar disorder in Swedish families: a population-based study. *Lancet* 2009, 373:234-239.
128. Reiss AL: Childhood developmental disorders: an academic and clinical convergence point for psychiatry, neurology, psychology and pediatrics. *Journal of child psychology and psychiatry, and allied disciplines* 2009, 50:87-98.
129. Aldridge K, George ID, Cole KK, Austin JR, Takahashi TN, Duan Y, Miles JH: Facial phenotypes in subgroups of prepubertal boys with autism spectrum disorders are correlated with clinical phenotypes. *Molecular autism* 2011, 2:15.
130. Miles JH: Autism spectrum disorders--a genetics review. *Genetics in medicine : official journal of the American College of Medical Genetics* 2011, 13:278-294.
131. Boyden LM, Choi M, Choate KA, Nelson-Williams CJ, Farhi A, Toka HR, Tikhonova IR, Bjornson R, Mane SM, Colussi G, et al: Mutations in kelch-like 3 and cullin 3 cause hypertension and electrolyte abnormalities. *Nature* 2012, 482:98-102.
132. Vig BK: Somatic mosaicism in plants with special reference to somatic crossing over. *Environmental health perspectives* 1978, 27:27-36.
133. Bakker E, Veenema H, Den Dunnen JT, van Broeckhoven C, Grootsholten PM, Bonten EJ, van Ommen GJ, Pearson PL: Germinal mosaicism increases the recurrence risk for 'new' Duchenne muscular dystrophy mutations. *Journal of medical genetics* 1989, 26:553-559.
134. Hall JG: Review and hypotheses: somatic mosaicism: observations related to clinical genetics. *American journal of human genetics* 1988, 43:355-363.
135. Sastry GR, Cooper HB, Jr., Brink RA: Paramutation and somatic mosaicism in maize. *Genetics* 1965, 52:407-424.
136. Hollander WF: Sectorial mosaics in the domestic pigeon: 25 more years. *The Journal of heredity* 1975, 66:177-202.
137. Biesecker LG, Spinner NB: A genomic view of mosaicism and human disease. *Nature reviews Genetics* 2013, 14:307-320.
138. Jongmans MC, Verwiel ET, Heijdra Y, Vulliamy T, Kamping EJ, Hehir-Kwa JY, Bongers EM, Pfundt R, van Emst L, van Leeuwen FN, et al: Revertant somatic mosaicism by mitotic recombination in dyskeratosis congenita. *American journal of human genetics* 2012, 90:426-433.
139. Steinbusch C, van Roozendaal K, Tserpelis D, Smeets E, Kranenburg-de Koning T, de Waal K, Zweier C, Rauch A, Hennekam R, Blok M, Schrandt-Stumpel C: Somatic mosaicism in a mother of two children with Pitt-Hopkins syndrome. *Clinical genetics* 2012.
140. Yamada M, Okura Y, Suzuki Y, Fukumura S, Miyazaki T, Ikeda H, Takezaki S, Kawamura N, Kobayashi I, Ariga T: Somatic mosaicism in two unrelated patients with X-linked chronic granulomatous disease characterized by the presence of a small population of normal cells. *Gene* 2012, 497:110-115.
141. Lindhurst MJ, Sapp JC, Teer JK, Johnston JJ, Finn EM, Peters K, Turner J, Cannons JL, Bick D, Blakemore L, et al: A mosaic activating mutation in AKT1 associated with the Proteus syndrome. *The New England journal of medicine* 2011, 365:611-619.
142. Choate KA, Lu Y, Zhou J, Choi M, Elias PM, Farhi A, Nelson-Williams C, Crumrine D, Williams ML, Nopper AJ, et al: Mitotic recombination in patients with ichthyosis causes reversion of dominant mutations in KRT10. *Science* 2010, 330:94-97.
143. Coufal NG, Garcia-Perez JL, Peng GE, Marchetto MC, Muotri AR, Mu Y, Carson CT, Macia A, Moran JV, Gage FH: Ataxia telangiectasia mutated (ATM) modulates long interspersed element-1 (L1) retrotransposition in human neural stem cells. *Proceedings of the National Academy of Sciences of the United States of America* 2011, 108:20382-20387.
144. Margari L, Lamanna AL, Buttiglione M, Craig F, Petruzzelli MG, Terenzio V: Long-term follow-up of neurological manifestations in a boy with incontinentia pigmenti. *European journal of pediatrics* 2013.
145. Huisman SA, Redeker EJ, Maas SM, Mannens MM, Hennekam RC: High rate of mosaicism in individuals with Cornelia de Lange syndrome. *Journal of medical genetics* 2013, 50:339-344.
146. Lindhurst MJ, Parker VE, Payne F, Sapp JC, Rudge S, Harris J, Witkowski AM, Zhang Q, Groeneveld MP, Scott CE, et al: Mosaic overgrowth with fibroadipose hyperplasia is caused by somatic activating mutations in PIK3CA. *Nature genetics* 2012, 44:928-933.
147. Tanaka T, Takahashi K, Yamane M, Tomida S, Nakamura S, Oshima K, Niwa A, Nishikomori R, Kambe N, Hara H, et al: Induced pluripotent stem cells from CINCA syndrome patients as a model for dissecting somatic mosaicism and drug discovery. *Blood* 2012, 120:1299-1308.

148. Kurek KC, Luks VL, Ayturk UM, Alomari AI, Fishman SJ, Spencer SA, Mulliken JB, Bowen ME, Yamamoto GL, Kozakewich HP, Warman ML: Somatic mosaic activating mutations in PIK3CA cause CLOVES syndrome. *American journal of human genetics* 2012, 90:1108-1115.
149. Macosko EZ, McCarroll SA: Exploring the variation within. *Nature genetics* 2012, 44:614-616.
150. Weiss KM: Cryptic causation of human disease: reading between the (germ) lines. *Trends Genet* 2005, 21:82-88.
151. Baugher JD, Baugher BD, Shirley MD, Pevsner J: Sensitive and specific detection of mosaic chromosomal abnormalities using the Parent-of-Origin-based Detection (POD) method. *BMC genomics* 2013, 14:367.
152. Shirley MD, Tang H, Gallione CJ, Baugher JD, Frelin LP, Cohen B, North PE, Marchuk DA, Comi AM, Pevsner J: Sturge-Weber syndrome and port-wine stains caused by somatic mutation in GNAQ. *The New England journal of medicine* 2013, 368:1971-1979.
153. Hulten MA, Jonasson J, Nordgren A, Iwarsson E: Germinal and Somatic Trisomy 21 Mosaicism: How Common is it, What are the Implications for Individual Carriers and How Does it Come About? *Current genomics* 2010, 11:409-419.
154. Kovaleva NV: Germ-line transmission of trisomy 21: Data from 80 families suggest an implication of grandmaternal age and a high frequency of female-specific trisomy rescue. *Molecular cytogenetics* 2010, 3:7.
155. Iourov IY, Vorsanova SG, Yurov YB: Chromosomal mosaicism goes global. *Molecular cytogenetics* 2008, 1:26.
156. Etheridge SP, Bowles NE, Arrington CB, Pilcher T, Rope A, Wilde AA, Alders M, Saarel EV, Tavernier R, Timothy KW, Tristani-Firouzi M: Somatic mosaicism contributes to phenotypic variation in Timothy syndrome. *American journal of medical genetics Part A* 2011, 155A:2578-2583.
157. Aldred MA, Bagshaw RJ, Macdermot K, Casson D, Murch SH, Walker-Smith JA, Trembath RC: Germline mosaicism for a GNAS1 mutation and Albright hereditary osteodystrophy. *Journal of medical genetics* 2000, 37:E35.
158. Barbosa RH, Vargas FR, Aguiar FC, Ferman S, Lucena E, Bonvicino CR, Seunez HN: Hereditary retinoblastoma transmitted by maternal germline mosaicism. *Pediatric blood & cancer* 2008, 51:598-602.
159. Chaturvedi LS, Mittal RD, Srivastava S, Mukherjee M, Mittal B: Analysis of dinucleotide repeat loci of dystrophin gene for carrier detection, germline mosaicism and de novo mutations in Duchenne muscular dystrophy. *Clin Genet* 2000, 58:234-236.
160. Evans JC, Archer HL, Whatley SD, Clarke A: Germline mosaicism for a MECP2 mutation in a man with two Rett daughters. *Clin Genet* 2006, 70:336-338.
161. Frank J, Happle R: Cutaneous mosaicism: right before our eyes. *J Clin Invest* 2007, 117:1216-1219.
162. Hosoki K, Takano K, Sudo A, Tanaka S, Saitoh S: Germline mosaicism of a novel UBE3A mutation in Angelman syndrome. *American journal of medical genetics Part A* 2005, 138A:187-189.
163. Jongmans MC, Hoefsloot LH, van der Donk KP, Admiraal RJ, Magee A, van de Laar I, Hendriks Y, Verheij JB, Walpole I, Brunner HG, van Ravenswaaij CM: Familial CHARGE syndrome and the CHD7 gene: a recurrent missense mutation, intrafamilial recurrence and variability. *American journal of medical genetics Part A* 2008, 146A:43-50.
164. Mari F, Caselli R, Russo S, Cogliati F, Ariani F, Longo I, Bruttini M, Meloni I, Pescucci C, Schurfeld K, et al: Germline mosaicism in Rett syndrome identified by prenatal diagnosis. *Clin Genet* 2005, 67:258-260.
165. Meyer KJ, Axelsen MS, Sheffield VC, Patil SR, Wassink TH: Germline mosaic transmission of a novel duplication of PXDN and MYT1L to two male half-siblings with autism. *Psychiatric genetics* 2012, 22:137-140.
166. Parodi S, Bachetti T, Lantieri F, Di Duca M, Santamaria G, Ottonello G, Matera I, Ravazzolo R, Ceccherini I: Parental origin and somatic mosaicism of PHOX2B mutations in Congenital Central Hypoventilation Syndrome. *Hum Mutat* 2008, 29:206.
167. Pauli S, Pieper L, Haberle J, Grzmil P, Burfeind P, Steckel M, Lenz U, Michelmann HW: Proven germline mosaicism in a father of two children with CHARGE syndrome. *Clin Genet* 2009, 75:473-479.
168. Rand CM, Yu M, Jennings LJ, Panesar K, Berry-Kravis EM, Zhou L, Weese-Mayer DE: Germline mosaicism of PHOX2B mutation accounts for familial recurrence of congenital central hypoventilation syndrome (CCHS). *American journal of medical genetics Part A* 2012, 158A:2297-2301.
169. Sato N, Ohyama K, Fukami M, Okada M, Ogata T: Kallmann syndrome: somatic and germline mutations of the fibroblast growth factor receptor 1 gene in a mother and the son. *The Journal of clinical endocrinology and metabolism* 2006, 91:1415-1418.

170. Sbidian E, Feldmann D, Bengoa J, Fraitag S, Abadie V, de Prost Y, Bodemer C, Hadj-Rabia S: Germline mosaicism in keratitis-ichthyosis-deafness syndrome: pre-natal diagnosis in a familial lethal form. *Clin Genet* 2010, 77:587-592.
171. Shanske AL, Goodrich JT, Ala-Kokko L, Baker S, Frederick B, Levy B: Germline mosaicism in Shprintzen-Goldberg syndrome. *American journal of medical genetics Part A* 2012, 158A:1574-1578.
172. Slavin TP, Lazebnik N, Clark DM, Vengoechea J, Cohen L, Kaur M, Konczal L, Crowe CA, Corteville JE, Nowaczyk MJ, et al: Germline mosaicism in Cornelia de Lange syndrome. *American journal of medical genetics Part A* 2012, 158A:1481-1485.
173. Sol-Church K, Stably DL, Demmer LA, Agbulos A, Lin AE, Smoot L, Nicholson L, Gripp KW: Male-to-male transmission of Costello syndrome: G12S HRAS germline mutation inherited from a father with somatic mosaicism. *American journal of medical genetics Part A* 2009, 149A:315-321.
174. Tajir M, Fergelot P, Lancelot G, Elalaoui SC, Arveiler B, Lacombe D, Sefiani A: Germline mosaicism in Rubinstein-Taybi syndrome. *Gene* 2013, 518:476-478.
175. Trevisson E, Forzan M, Salviati L, Clementi M: Neurofibromatosis type 1 in two siblings due to maternal germline mosaicism. *Clin Genet* 2013.
176. Venancio M, Santos M, Pereira SA, Maciel P, Saraiva JM: An explanation for another familial case of Rett syndrome: maternal germline mosaicism. *European journal of human genetics : EJHG* 2007, 15:902-904.
177. Wuyts W, Biervliet M, Reyniers E, D'Apice MR, Novelli G, Storm K: Somatic and gonadal mosaicism in Hutchinson-Gilford progeria. *American journal of medical genetics Part A* 2005, 135:66-68.
178. Jonsson T, Atwal JK, Steinberg S, Snaedal J, Jonsson PV, Bjornsson S, Stefansson H, Sulem P, Gudbjartsson D, Maloney J, et al: A mutation in APP protects against Alzheimer's disease and age-related cognitive decline. *Nature* 2012, 488:96-99.
179. Styrkarsdottir U, Thorleifsson G, Sulem P, Gudbjartsson DF, Sigurdsson A, Jonasdottir A, Oddsson A, Helgason A, Magnusson OT, Walters GB, et al: Nonsense mutation in the LGR4 gene is associated with several human diseases and other traits. *Nature* 2013.
180. Williams MS: Myostatin mutation associated with gross muscle hypertrophy in a child. *N Engl J Med* 2004, 351:1030-1031; author reply 1030-1031.
181. Boyden LM, Mao J, Belsky J, Mitzner L, Farhi A, Mitnick MA, Wu D, Insogna K, Lifton RP: High bone density due to a mutation in LDL-receptor-related protein 5. *N Engl J Med* 2002, 346:1513-1521.
182. Liam Hoekstra Strongest Boy in the World [<http://videos.disabled-world.com/video/159/liam-hoekstra-strongest-boy-in>]
183. Fu M, Zhang X, Lai X, Wu X, Feng F, Peng J, Zhong H, Zhang Y, Wang Y, Zhou Q, et al: Generation of Sequence Variants via Accelerated Molecular Evolution Methods. *Recent patents on DNA & gene sequences* 2013.
184. Leconte AM, Dickinson BC, Yang DD, Chen IA, Allen B, Liu DR: A population-based experimental model for protein evolution: effects of mutation rate and selection stringency on evolutionary outcomes. *Biochemistry* 2013, 52:1490-1499.
185. Esvelt KM, Carlson JC, Liu DR: A system for the continuous directed evolution of biomolecules. *Nature* 2011, 472:499-503.
186. Mitchell KJ, Porteous DJ: Rethinking the genetic architecture of schizophrenia. *Psychological medicine* 2011, 41:19-32.
187. Girirajan S, Rosenfeld JA, Coe BP, Parikh S, Friedman N, Goldstein A, Filipink RA, McConnell JS, Angle B, Meschino WS, et al: Phenotypic Heterogeneity of Genomic Disorders and Rare Copy-Number Variants. *The New England journal of medicine* 2012.
188. Shi L, Zhang X, Golhar R, Otieno FG, He M, Hou C, Kim C, Keating B, Lyon GJ, Wang K, Hakonarson H: Whole-genome sequencing in an autism multiplex family. *Molecular autism* 2013, 4:8.
189. Fullston T, Gabb B, Callen D, Ullmann R, Woollatt E, Bain S, Ropers HH, Cooper M, Chandler D, Carter K, et al: Inherited balanced translocation t(9;17)(q33.2;q25.3) concomitant with a 16p13.1 duplication in a patient with schizophrenia. *Am J Med Genet B Neuropsychiatr Genet* 2011, 156:204-214.
190. Crepel A, Breckpot J, Fryns JP, De la Marche W, Steyaert J, Devriendt K, Peeters H: DISC1 duplication in two brothers with autism and mild mental retardation. *Clin Genet* 2010, 77:389-394.
191. Klei L, Sanders SJ, Murtha MT, Hus V, Lowe JK, Willsey AJ, Moreno-De-Luca D, Yu TW, Fombonne E, Geschwind D, et al: Common genetic variants, acting additively, are a major source of risk for autism. *Molecular autism* 2012, 3:9.

192. Visscher PM, Goddard ME, Derks EM, Wray NR: Evidence-based psychiatric genetics, AKA the false dichotomy between common and rare variant hypotheses. *Molecular psychiatry* 2011.
193. Zuk O, Hechter E, Sunyaev SR, Lander ES: The mystery of missing heritability: Genetic interactions create phantom heritability. *Proceedings of the National Academy of Sciences of the United States of America* 2012, 109:1193-1198.
194. Anney R, Klei L, Pinto D, Almeida J, Bacchelli E, Baird G, Bolshakova N, Bolte S, Bolton PF, Bourgeron T, et al: Individual common variants exert weak effects on the risk for autism spectrum disorders. *Human molecular genetics* 2012, 21:4781-4792.
195. Schaaf CP, Sabo A, Sakai Y, Crosby J, Muzny D, Hawes A, Lewis L, Akbar H, Varghese R, Boerwinkle E, et al: Oligogenic heterozygosity in individuals with high-functioning autism spectrum disorders. *Human molecular genetics* 2011, 20:3366-3375.
196. Gagnon F, Roslin NM, Lemire M: Successful identification of rare variants using oligogenic segregation analysis as a prioritizing tool for whole-exome sequencing studies. *BMC proceedings* 2011, 5 Suppl 9:S11.
197. Malhotra D, Sebat J: CNVs: harbingers of a rare variant revolution in psychiatric genetics. *Cell* 2012, 148:1223-1241.
198. Gai X, Xie HM, Perin JC, Takahashi N, Murphy K, Wenocur AS, D'Arcy M, O'Hara RJ, Goldmuntz E, Grice DE, et al: Rare structural variation of synapse and neurotransmission genes in autism. *Molecular psychiatry* 2012, 17:402-411.
199. Shaikh TH, Haldeman-Englert C, Geiger EA, Ponting CP, Webber C: Genes and biological processes commonly disrupted in rare and heterogeneous developmental delay syndromes. *Human molecular genetics* 2011, 20:880-893.
200. Elia J, Gai X, Xie HM, Perin JC, Geiger E, Glessner JT, D'Arcy M, deBerardinis R, Frackelton E, Kim C, et al: Rare structural variants found in attention-deficit hyperactivity disorder are preferentially associated with neurodevelopmental genes. *Molecular psychiatry* 2010, 15:637-646.
201. Novarino G, El-Fishawy P, Kayserili H, Meguid NA, Scott EM, Schroth J, Silhavy JL, Kara M, Khalil RO, Ben-Omran T, et al: Mutations in BCKD-kinase Lead to a Potentially Treatable Form of Autism with Epilepsy. *Science* 2012.
202. O'Roak BJ, Vives L, Girirajan S, Karakoc E, Krumm N, Coe BP, Levy R, Ko A, Lee C, Smith JD, et al: Sporadic autism exomes reveal a highly interconnected protein network of de novo mutations. *Nature* 2012, 485:246-250.
203. Iossifov I, Ronemus M, Levy D, Wang Z, Hakker I, Rosenbaum J, Yamrom B, Lee YH, Narzisi G, Leotta A, et al: De novo gene disruptions in children on the autistic spectrum. *Neuron* 2012, 74:285-299.
204. Neale BM, Kou Y, Liu L, Ma'ayan A, Samocha KE, Sabo A, Lin CF, Stevens C, Wang LS, Makarov V, et al: Patterns and rates of exonic de novo mutations in autism spectrum disorders. *Nature* 2012, 485:242-245.
205. Sanders SJ, Murtha MT, Gupta AR, Murdoch JD, Raubeson MJ, Willsey AJ, Ercan-Sencicek AG, DiLullo NM, Parikshak NN, Stein JL, et al: De novo mutations revealed by whole-exome sequencing are strongly associated with autism. *Nature* 2012, 485:237-241.
206. Xu B, Ionita-Laza I, Roos JL, Boone B, Woodrick S, Sun Y, Levy S, Gogos JA, Karayiorgou M: De novo gene mutations highlight patterns of genetic and neural complexity in schizophrenia. *Nature genetics* 2012.
207. Girirajan S, Rosenfeld JA, Cooper GM, Antonacci F, Siswara P, Itsara A, Vives L, Walsh T, McCarthy SE, Baker C, et al: A recurrent 16p12.1 microdeletion supports a two-hit model for severe developmental delay. *Nature genetics* 2010, 42:203-209.
208. Sanders SJ, Ercan-Sencicek AG, Hus V, Luo R, Murtha MT, Moreno-De-Luca D, Chu SH, Moreau MP, Gupta AR, Thomson SA, et al: Multiple recurrent de novo CNVs, including duplications of the 7q11.23 Williams syndrome region, are strongly associated with autism. *Neuron* 2011, 70:863-885.
209. O'Roak BJ, Vives L, Fu W, Egertson JD, Stanaway IB, Phelps IG, Carvill G, Kumar A, Lee C, Ankenman K, et al: Multiplex targeted sequencing identifies recurrently mutated genes in autism spectrum disorders. *Science* 2012, 338:1619-1622.
210. Genotyping Microarray [<http://www.illumina.com/applications/genotyping.ilmn>]
211. Schneider GF, Dekker C: DNA sequencing with nanopores. *Nature biotechnology* 2012, 30:326-328.
212. Williams AL, Patterson N, Glessner J, Hakonarson H, Reich D: Phasing of many thousands of genotyped samples. *American journal of human genetics* 2012, 91:238-251.
213. Peters BA, Kermani BG, Sparks AB, Alferov O, Hong P, Alexeev A, Jiang Y, Dahl F, Tang YT, Haas J, et al: Accurate whole-genome sequencing and haplotyping from 10 to 20 human cells. *Nature* 2012, 487:190-195.

214. Roach JC, Glusman G, Smit AF, Huff CD, Hubley R, Shannon PT, Rowen L, Pant KP, Goodman N, Bamshad M, et al: Analysis of genetic inheritance in a family quartet by whole-genome sequencing. *Science* 2010, 328:636-639.
215. Lupski JR, Reid JG, Gonzaga-Jauregui C, Rio Deiros D, Chen DC, Nazareth L, Bainbridge M, Dinh H, Jing C, Wheeler DA, et al: Whole-genome sequencing in a patient with Charcot-Marie-Tooth neuropathy. *The New England journal of medicine* 2010, 362:1181-1191.
216. McGuire AL, Lupski JR: Personal genome research : what should the participant be told? *Trends in genetics : TIG* 2010, 26:199-201.
217. Bainbridge MN, Wiszniewski W, Murdock DR, Friedman J, Gonzaga-Jauregui C, Newsham I, Reid JG, Fink JK, Morgan MB, Gingras MC, et al: Whole-genome sequencing for optimized patient management. *Science translational medicine* 2011, 3:87re83.
218. Ng SB, Bigham AW, Buckingham KJ, Hannibal MC, McMillin MJ, Gildersleeve HI, Beck AE, Tabor HK, Cooper GM, Mefford HC, et al: Exome sequencing identifies MLL2 mutations as a cause of Kabuki syndrome. *Nat Genet* 2010, 42:790-793.
219. Ng SB, Buckingham KJ, Lee C, Bigham AW, Tabor HK, Dent KM, Huff CD, Shannon PT, Jabs EW, Nickerson DA, et al: Exome sequencing identifies the cause of a mendelian disorder. *Nature genetics* 2010, 42:30-35.
220. Bamshad MJ, Ng SB, Bigham AW, Tabor HK, Emond MJ, Nickerson DA, Shendure J: Exome sequencing as a tool for Mendelian disease gene discovery. *Nature reviews Genetics* 2011, 12:745-755.
221. Hedges DJ, Burges D, Powell E, Almonte C, Huang J, Young S, Boese B, Schmidt M, Pericak-Vance MA, Martin E, et al: Exome sequencing of a multigenerational human pedigree. *PloS one* 2009, 4:e8232.
222. Lyon GJ: Personal account of the discovery of a new disease using next-generation sequencing. Interview by Natalie Harrison. *Pharmacogenomics* 2011, 12:1519-1523.
223. Katsanis SH, Katsanis N: Molecular genetic testing and the future of clinical genomics. *Nature reviews Genetics* 2013, 14:415-426.
224. Lyon GJ: Personalized medicine: Bring clinical standards to human-genetics research. *Nature* 2012, 482:300-301.
225. Guest post: Time to bring human genome sequencing into the clinic [<http://www.genomesunzipped.org/2012/02/guest-post-time-to-bring-human-genome-sequencing-into-the-clinic.php>]
226. Zumbo P, Mason CE: Molecular methods for profiling the RNA world. . In *Genome Analysis: Current Procedures and Applications* Horizon Press. ; 2014
227. The AceView genes [<http://www.ncbi.nlm.nih.gov/IEB/Research/Acembly/>]
228. Moreno-De-Luca A, Myers SM, Challman TD, Moreno-De-Luca D, Evans DW, Ledbetter DH: Developmental brain dysfunction: revival and expansion of old concepts based on new genetic evidence. *The Lancet Neurology* 2013, 12:406-414.
229. Nielsen R, Paul JS, Albrechtsen A, Song YS: Genotype and SNP calling from next-generation sequencing data. *Nature reviews Genetics* 2011, 12:443-451.
230. O'Rawe J, Guangqing S, Wang W, Hu J, Bodily P, Tian L, Hakonarson H, Johnson E, Wei Z, Jiang T, et al: Low concordance of multiple variant-calling pipelines: practical implications for exome and genome sequencing. *Genome Medicine* 2013, 5:28.
231. Kajiwaru K, Berson E, Dryja T: Digenic retinitis pigmentosa due to mutations at the unlinked peripherin/RDS and ROM1 loci. *Science* 1994, 264:1604-1608.
232. El-Hattab AW, Zhang F, Maxim R, Christensen KM, Ward JC, Hines-Dowell S, Scaglia F, Lupski JR, Cheung SW: Deletion and duplication of 15q24: Molecular mechanisms and potential modification by additional copy number variants. *Genet Med* 2010, 12:573-586.
233. Davis EE, Zhang Q, Liu Q, Diplas BH, Davey LM, Hartley J, Stoetzel C, Szymanska K, Ramaswami G, Logan CV, et al: TTC21B contributes both causal and modifying alleles across the ciliopathy spectrum. *Nat Genet* 2011.
234. Lemmers RJLF, Tawil R, Petek LM, Balog J, Block GJ, Santen GWE, Amell AM, van der Vliet PJ, Almomani R, Straasheijm KR, et al: Digenic inheritance of an SMCHD1 mutation and an FSHD-permissive D4Z4 allele causes facioscapulohumeral muscular dystrophy type 2. *Nat Genet* 2012, 44:1370-1374.
235. Lai J, Li R, Xu X, Jin W, Xu M, Zhao H, Xiang Z, Song W, Ying K, Zhang M, et al: Genome-wide patterns of genetic variation among elite maize inbred lines. *Nat Genet* 2010, 42:1027-1030.

236. Bloom JS, Ehrenreich IM, Loo WT, Lite TL, Kruglyak L: Finding the sources of missing heritability in a yeast cross. *Nature* 2013, 494:234-237.
237. Opitz JM: 2011 William Allan Award: development and evolution. *American journal of human genetics* 2012, 90:392-404.
238. [http://en.wikipedia.org/wiki/Conrad_Hal_Waddington]
239. Waddington CH: Canalization of development and genetic assimilation of acquired characters. *Nature* 1959, 183:1654-1655.
240. Waddington CH: The epigenotype. 1942. *International journal of epidemiology* 2012, 41:10-13.
241. McClaren BJ, Metcalfe SA, Amor DJ, Aitken M, Massie J: A case for cystic fibrosis carrier testing in the general population. *The Medical journal of Australia* 2011, 194:208-209.
242. ACMG Releases Policy Statement on Noninvasive Prenatal Screening (NIPS) [http://www.acmg.net/docs/NIPS_Release.pdf]
243. Fan HC, Gu W, Wang J, Blumenfeld YJ, El-Sayed YY, Quake SR: Non-invasive prenatal measurement of the fetal genome. *Nature* 2012, 487:320-324.
244. Kitzman JO, Snyder MW, Ventura M, Lewis AP, Qiu R, Simmons LE, Gammill HS, Rubens CE, Santillan DA, Murray JC, et al: Noninvasive whole-genome sequencing of a human fetus. *Science translational medicine* 2012, 4:137ra176.
245. Papageorgiou EA, Patsalis PC: Maternal plasma sequencing: a powerful tool towards fetal whole genome recovery. *BMC medicine* 2013, 11:56.
246. Cheng P, M DC, Chen CF, Hoffmann RF, Armitage R, Deldin PJ: Sleep-disordered breathing in major depressive disorder. *Journal of sleep research* 2013.
247. Bianchi DW: From prenatal genomic diagnosis to fetal personalized medicine: progress and challenges. *Nature medicine* 2012, 18:1041-1051.
248. Guedj F, Bianchi DW: Noninvasive prenatal testing creates an opportunity for antenatal treatment of down syndrome. *Prenat Diagn* 2013.
249. Bates BR, Graham D, Striley K, Patterson S, Arora A, Hamel-Lambert J: Examining Antecedents of Caregivers' Access to Early Childhood Developmental Screening: Implications for Campaigns Promoting Use of Services in Appalachian Ohio. *Health Promot Pract* 2013.
250. McIntyre LL: Parent training for young children with developmental disabilities: randomized controlled trial. *Am J Ment Retard* 2008, 113:356-368.
251. Rickards AL, Walstab JE, Wright-Rossi RA, Simpson J, Reddihough DS: A randomized, controlled trial of a home-based intervention program for children with autism and developmental delay. *J Dev Behav Pediatr* 2007, 28:308-316.
252. Rickards AL, Walstab JE, Wright-Rossi RA, Simpson J, Reddihough DS: One-year follow-up of the outcome of a randomized controlled trial of a home-based intervention programme for children with autism and developmental delay and their families. *Child Care Health Dev* 2009, 35:593-602.
253. Salem Y, Gropack SJ, Coffin D, Godwin EM: Effectiveness of a low-cost virtual reality system for children with developmental delay: a preliminary randomised single-blind controlled trial. *Physiotherapy* 2012, 98:189-195.
254. Velleman SL, Mervis CB: Children with 7q11.23 Duplication Syndrome: Speech, Language, Cognitive, and Behavioral Characteristics and their Implications for Intervention. *Perspect Lang Learn Educ* 2011, 18:108-116.
255. Coulter ME, Miller DT, Harris DJ, Hawley P, Picker J, Roberts AE, Sobeih MM, Irons M: Chromosomal microarray testing influences medical management. *Genetics in medicine : official journal of the American College of Medical Genetics* 2011, 13:770-776.
256. Brawley OW, Goldberg P: *How we do harm : a doctor breaks ranks about being sick in America*. 1st edn. New York: St. Martin's Press; 2012.
257. Makary M: *Unaccountable : what hospitals won't tell you and how transparency can revolutionize health care*. 1st U.S. edn. New York: Bloomsbury Press; 2012.
258. Brandt DS, Shinkunas L, Hillis SL, Daack-Hirsch SE, Driessnack M, Downing NR, Liu MF, Shah LL, Williams JK, Simon CM: A Closer Look at the Recommended Criteria for Disclosing Genetic Results: Perspectives of Medical Genetic Specialists, Genomic Researchers, and Institutional Review Board Chairs. *J Genet Couns* 2013.
259. Hamamy H: Consanguineous marriages : Preconception consultation in primary health care settings. *J Community Genet* 2012, 3:185-192.
260. Bittles AH: Genetics and global healthcare. *J R Coll Physicians Edinb* 2013, 43:7-10.

261. Francke U, Dijamco C, Kiefer AK, Eriksson N, Moiseff B, Tung JY, Mountain JL: Dealing with the unexpected: consumer responses to direct-access BRCA mutation testing. *PeerJ* 2013, 1:e8.
262. Chua EW, Kennedy MA: Current State and Future Prospects of Direct-to-Consumer Pharmacogenetics. *Frontiers in pharmacology* 2012, 3:152.
263. Sterckx S, Cockbain J, Howard H, Huys I, Borry P: "Trust is not something you can reclaim easily": patenting in the field of direct-to-consumer genetic testing. *Genetics in medicine : official journal of the American College of Medical Genetics* 2013, 15:382-387.
264. Allain D, Friedman S, Senter L: Consumer awareness and attitudes about insurance discrimination post enactment of the Genetic Information Nondiscrimination Act. *Familial Cancer* 2012, 11:637-644.
265. Bakker E: Is the DNA Sequence the Gold Standard in Genetic Testing? Quality of Molecular Genetic Tests Assessed. *Clinical Chemistry* 2006, 52:557-558.
266. Individual Genome Sequencing (IGS) Test
[http://www.illumina.com/clinical/illumina_clinical_laboratory.ilmn]
267. Hunter LE, Hopfer C, Terry SF, Coors ME: Reporting Actionable Research Results: Shared Secrets Can Save Lives. *Science translational medicine* 2012, 4:143cm148.
268. 23andMe [<https://http://www.23andme.com/>]
269. Personal Genome Project [<http://www.personalgenomes.org/about.html>]
270. Fu W, O'Connor TD, Jun G, Kang HM, Abecasis G, Leal SM, Gabriel S, Altshuler D, Shendure J, Nickerson DA, et al: Analysis of 6,515 exomes reveals the recent origin of most human protein-coding variants. *Nature* 2013, 493:216-220.
271. An integrated map of genetic variation from 1,092 human genomes. *Nature* 2012, 491:56-65.
272. McGuire AL, Gibbs RA: No Longer De-Identified. *Science* 2006, 312:370-371.
273. Gymrek M, McGuire AL, Golan D, Halperin E, Erlich Y: Identifying Personal Genomes by Surname Inference. *Science* 2013, 339:321-324.
274. Lowrance WW, Collins FS: Identifiability in Genomic Research. *Science* 2007, 317:600-602.
275. Radick G: Other Histories, Other Biologies. In *Philosophy, Biology and Life*. Cambridge: Cambridge University Press; 2005: 21-47
276. Scientific Inheritance - an Inaugural Lecture from Gregory Radick
[<http://www.youtube.com/watch?v=D3nyB2lqmRo>]
277. Goddard KA, Whitlock EP, Berg JS, Williams MS, Webber EM, Webster JA, Lin JS, Schrader KA, Campos-Outcalt D, Offit K, et al: Description and pilot results from a novel method for evaluating return of incidental findings from next-generation sequencing technologies. *Genetics in medicine : official journal of the American College of Medical Genetics* 2013.
278. Berg JS, Adams M, Nassar N, Bizon C, Lee K, Schmitt CP, Wilhelmsen KC, Evans JP: An informatics approach to analyzing the incidentalome. *Genetics in medicine : official journal of the American College of Medical Genetics* 2013, 15:36-44.
279. Green RC, Berg JS, Berry GT, Biesecker LG, Dimmock DP, Evans JP, Grody WW, Hegde MR, Kalia S, Korf BR, et al: Exploring concordance and discordance for return of incidental findings from clinical sequencing. *Genetics in medicine : official journal of the American College of Medical Genetics* 2012, 14:405-410.
280. ACMG: ACMG Recommendations for Reporting of Incidental Findings in Clinical Exome and Genome Sequencing. 2013.
281. Mole: How we know II: bad dreams *Journal of Cell Science* 2006, 119:197-198