

Extra Views

Reversing Drug Resistance In Vivo

Hans-Guido Wendel

Scott W. Lowe*

Cold Spring Harbor Laboratory; Cold Spring Harbor, New York USA

*Correspondence to: Scott W. Lowe; CSHL 1 Bungtown Rd.; Cold Spring Harbor, New York 11724 USA; Tel.: 516.367.8406x8408; Fax: 516.367.8454 Email: lowe@cshl.edu

Received 05/11/04; Accepted 05/14/04

Previously published online as a *Cell Cycle* E-Publication:
<http://www.landesbioscience.com/journals/cc/abstract.php?id=976>

KEY WORDS

apoptosis, drug resistance, PI3 kinase, Akt, rapamycin, eIF4E, mouse model

ABBREVIATIONS

eIF4E	eukaryotic translation initiation factor 4E
mTOR	mammalian target of rapamycin
PI3K	phosphatidylinositol 3-kinase
PTEN	phosphatase and tensin homolog
TSC1/2	tuberous sclerosis 1/2 gene

ABSTRACT

Apoptotic defects occur in oncogenesis and contribute to drug resistance. We have shown that Bcl-2, Akt, and the translational regulator eIF4E cooperate with Myc during lymphomagenesis and are potent inducers of drug resistance. Interestingly, lymphomas expressing Akt, but not those expressing Bcl-2 are sensitized to chemotherapy-induced apoptosis by the mTOR inhibitor rapamycin, an effect that is countered by eIF4E. These results provide in vivo validation for a strategy to reverse drug resistance in human cancers and highlight the potential role of translational deregulation in oncogenesis and resistance. They also illustrate the importance of tailoring cancer therapy based on tumor genotype.

Apoptotic defects are a common event in oncogenesis, as disruption of apoptotic programs is required to counteract the pro-apoptotic effects of oncogene signaling and other stresses encountered during tumorigenesis.¹ Since chemotherapeutic drugs also engage these apoptotic pathways, their disruption frequently causes drug resistance.² In many cancers, apoptosis is disabled through mutations that ultimately produce hyperactivation of survival signaling molecules such as Bcl-2 or Akt. As a consequence, there are now intense efforts to understand and short-circuit the effects of deregulated survival signaling in cancer.

Our laboratory examines how oncogene and tumor suppressor networks that control cell survival are disrupted during the course of tumor evolution and how mutations in these networks impact tumor cell responses to conventional and targeted therapeutics in vivo. To this end, we have used the E μ -*myc* mouse model of B-cell lymphomagenesis—not to study lymphomagenesis and lymphoma therapy per se—but to exploit an extremely powerful system for studying genetic interactions relevant to tumorigenesis and treatment responses in vivo. To facilitate these efforts, we developed stem cell manipulation/transplantation protocols to rapidly produce lymphomas with compound genotypes, and employed methods to study treatment responses using real endpoints such as remission and survival in a manner that parallels human clinical trials. Using this system, we have previously shown that disruption of apoptosis, for example through loss of *p53* or overexpression of Bcl-2, dramatically accelerates *myc*-induced lymphomagenesis and promotes drug resistance. Moreover, despite the fact that lymphomas are initiated with the same oncogene and treated in the same way, we note an enormous heterogeneity in response, which is dependent on other genetic alterations in the tumor.

We recently examined the consequences of deregulated Akt signaling on lymphomagenesis and treatment responses in the E μ -*myc* model. The PI3K/Akt pathway is activated in many tumors through loss of the tumor suppressors PTEN, TSC1/2 or via amplification of Akt or constitutive activation of *ras* (reviewed in ref. 3). Normally the PI3K/Akt pathway transmits growth and survival signals from surface receptors to affect cellular physiology in multiple ways. However, when we directly compared overexpression of Akt to the strictly anti-apoptotic *bcl-2*, we found that both oncogenes had an identical effect on *myc*-induced lymphomagenesis. Both caused the rapid onset of aggressive, multidrug resistant lymphomas of a primordial B cell type. Therefore, despite the myriad of effects Akt has on cellular physiology, its antiapoptotic function appears critical for oncogenesis and drug resistance.

In principle, reversing apoptotic defects through pharmacological or genetic approaches should sensitize drug resistant tumors to cytotoxic therapy. Akt has been proposed to mediate cell survival by phosphorylating proteins directly involved in apoptosis (Caspase 9, Bad, Mdm-2, reviewed in ref. 3), but it also signals transcriptional and translational changes that may indirectly promote survival. However, the relevant contributions of each effector process to oncogenesis and drug resistance, and hence the best molecular targets, remain unclear. Pharmacologically, the choices are currently limited, since Akt inhibitors

are not yet available and drugs targeting the PI3K are toxic. However, one drug—rapamycin—has been widely used as an immunosuppressant in the clinic and is a potent inhibitor of the Akt effector mTOR. Although not typically associated with the Akt-mediated survival signal, mTOR inhibitors are being developed as anti-cancer drugs and have shown some activity towards tumors with defects in the PI3Kinase pathway both in vitro and in vivo.⁴⁻⁷ Therefore, we tested whether rapamycin could reverse the Akt mediated apoptotic block in our system.

While rapamycin alone had little effect on lymphomas of any genotype, it markedly sensitized Akt overexpressing tumors to cytotoxic chemotherapy.⁸ In combination with chemotherapy, rapamycin induced massive apoptosis and lasting remissions without causing increased toxicity. Therefore, targeting a survival pathway can indeed reverse drug resistance in tumors.

Do our studies predict that rapamycin combinations should be universally beneficial? Clearly, they reveal that rapamycin combined with conventional chemotherapy can be synergistic in eliminating tumor cells with mutational activation of the Akt pathway and, indeed, others have observed similar results using targeted drugs.⁹ Our results also raise the possibility that rapamycin may enhance drug sensitivity when Akt is activated by growth or survival factors that would otherwise create local sanctuaries of drug resistant cells. Unfortunately, our experiments do not support a broad use of rapamycin to combat drug resistance, since its beneficial effects did not extend to tumors with other apoptotic defects. Furthermore, in chemosensitive control tumors lacking the ARF tumor suppressor but retaining p53, the combination of rapamycin and doxorubicin was antagonistic. This effect occurred only if rapamycin preceded doxorubicin treatment (unpublished observations), and may be the consequence of a rapamycin induced cell cycle arrest.¹⁰ Clearly, these findings imply that rapamycin/chemotherapy combinations are not a 'one-size-fits-all' tumor therapy, but require careful, molecular identification of patients who would benefit from a therapy targeting the PI3K/Akt pathway.

The fact that rapamycin—a specific inhibitor of the translational regulator mTOR—potently reverses the Akt survival signal suggests that control of translation may be important for Akt-mediated cell survival. Consistent with this idea, eIF4E—a translation factor downstream of mTOR—can co-operate in the transformation of primary cells and block apoptosis induced by *myc* or *ras* in vitro.¹¹⁻¹³ Also, in a glioblastoma model, expression of Akt induced translational effects, evident as the rapamycin-sensitive recruitment of certain capped mRNAs into the polyribosome fraction.¹⁴ However, it is also possible that Akt driven lymphomas become “addicted” to the mTOR signal and undergo catastrophic death in its absence. Such a possibility implies that the increased chemosensitivity of the Akt tumors in the presence of rapamycin does not reflect actual reversal of an apoptotic defect, but rather a hypersensitivity to the loss of mTOR signaling.

To further investigate the role of translational deregulation in oncogenesis, we tested the translation initiation factor eIF4E (reviewed in ref. 15) directly in our system. Indeed, we found that eIF4E co-operates with *myc* in a manner that is comparable to Akt, leading to the rapid development of lymphomas. Furthermore, expression of eIF4E was sufficient to confer rapamycin resistance to Akt tumors. This effect was mediated by eIF4E's ability to block apoptosis and compensate for loss of p53 in tumorigenesis.⁸ Consistent with our results, ubiquitous expression of eIF4E in transgenic mice promotes tumorigenesis in the lung, liver and

hematopoietic compartment, and co-operates with *myc* in lymphomagenesis.¹⁶ Clearly, the translation factor eIF4E is oncogenic in vivo and sufficient to replace either Akt or p53 loss in *myc*-driven tumors, and implies that translational regulation can mediate—or at least compensate for—the Akt survival signal.

Do these results mean that only the eIF4E-mediated translational effects are important for Akt's function? Not necessarily. Indeed, it will be interesting to see whether eIF4E can co-operate in oncogenesis with, e.g., S6-kinase, inactivation of caspases or Bad, or how eIF4E's effects on translation intersect with the metabolic alterations that distinguish the survival activities of Akt and Bcl-x_L in FL5-12 cells.¹⁷ Conversely, non-Akt mediated signals may also intersect at the level of eIF4E regulation. For example, the PIM-2 kinase produces a rapamycin/mTOR independent signal that can activate eIF4E.¹⁸ Presumably, the relevant contribution of different survival pathways and their effectors may depend on the cellular context. Yet, together with recent evidence implying defects in mRNA processing in dyskeratosis¹⁹ (reviewed in ref. 20) and amplification of eIF4E in several cancers,²¹⁻²³ the surprising ability of eIF4E to impact lymphomagenesis in our model hints at a broader role for translational deregulation in cancer.

OUTLOOK

Although our study provides a compelling example of how reversing apoptotic defects can also reverse drug resistance, several important questions remain. To what extent do Eμ-*myc* lymphomas reflect the behavior of human tumors? If clinical studies recapitulate our results, this will validate genetically controlled tumor models, for uncovering drug resistance mechanisms and as preclinical models for testing new therapeutic approaches. Which patients will benefit from an mTOR-based therapy and how can they be identified? In our study, lymphomas could have virtually identical pathology yet respond very differently to rapamycin—presumably molecular markers that reflect tumor genotype will be required. Are there better drug targets in the Akt pathway? Based on our model, mTOR is a good target, but others may be better. For example direct inhibition of Akt would target a broader range of effectors while, conversely, inhibiting translational initiation downstream of mTOR may provide less opportunities for resistance. However, it is also possible that interfering with upstream and downstream targets may increase toxicity. Finally, can other survival pathways be targeted? Presumably yes, as Bcl-2 antisense oligonucleotides have shown promise in pre-clinical and clinical studies (reviewed in ref. 24). Clearly, targeted therapeutics hold great promise in cancer therapy and genetically controlled tumor models, like the Eμ-*myc* model, may prove to be particularly valuable in their preclinical development.

References

- Hanahan D, Weinberg RA. The hallmarks of cancer. *Cell* 2000; 100:57-70.
- Lowe SW, Ruley HE, Jacks T, Housman DE. p53-dependent apoptosis modulates the cytotoxicity of anticancer agents. *Cell* 1993; 74:957-67.
- Vivanco I, Sawyers CL. The phosphatidylinositol 3-Kinase AKT pathway in human cancer. *Nat Rev Cancer* 2002; 2:489-501.
- Grunwald V, DeGraffenried L, Russel D, Friedrichs WE, Ray RB, Hidalgo M. Inhibitors of mTOR reverse doxorubicin resistance conferred by PTEN status in prostate cancer cells. *Cancer Res* 2002; 62:6141-5.
- Neshat MS, Mellinger IK, Tran C, Stiles B, Thomas G, Petersen R, Frost P, Gibbons JJ, Wu H, Sawyers CL. Enhanced sensitivity of PTEN-deficient tumors to inhibition of FRAP/mTOR. *Proc Natl Acad Sci USA* 2001; 98:10314-9.
- Shi Y, Gera J, Hu L, Hsu JH, Bookstein R, Li W, Lichtenstein A. Enhanced sensitivity of multiple myeloma cells containing PTEN mutations to CCI-779. *Cancer Res* 2002; 62:5027-34.

7. Podsypanina K, Lee RT, Politis C, Hennessy I, Crane A, Puc J, et al. An inhibitor of mTOR reduces neoplasia and normalizes p70/S6 kinase activity in *Pten*^{-/-} mice. *Proc Natl Acad Sci USA* 2001; 98:10320-5.
8. Wendel HG, De Stanchina E, Fridman JS, Malina A, Ray S, Kogan S, et al. Survival signalling by Akt and eIF4E in oncogenesis and cancer therapy. *Nature* 2004; 428:332-7.
9. Mohi MG, Boulton C, Gu TL, Sternberg DW, Neuberger D, Griffin JD, et al. Combination of rapamycin and protein tyrosine kinase (PTK) inhibitors for the treatment of leukemias caused by oncogenic PTKs. *Proc Natl Acad Sci USA* 2004; 101:3130-5.
10. Hosoi H, Dilling MB, Shikata T, Liu LN, Shu L, Ashmun RA, et al. Rapamycin causes poorly reversible inhibition of mTOR and induces p53-independent apoptosis in human rhabdomyosarcoma cells. *Cancer Res* 1999; 59:886-94.
11. Polunovsky VA, Rosenwald IB, Tan AT, White J, Chiang L, Sonenberg N, et al. Translational control of programmed cell death: eukaryotic translation initiation factor 4E blocks apoptosis in growth-factor-restricted fibroblasts with physiologically expressed or deregulated Myc. *Mol Cell Biol* 1996; 16:6573-81.
12. Li S, Takasu T, Perlman DM, Peterson MS, Burrichter D, Avdulov S, et al. Translation factor eIF4E rescues cells from Myc-dependent apoptosis by inhibiting cytochrome c release. *J Biol Chem* 2003; 278:3015-22.
13. Lazaris-Karatzas A, Montine KS, Sonenberg N. Malignant transformation by a eukaryotic initiation factor subunit that binds to mRNA 5' cap. *Nature* 1990; 345:544-7.
14. Rajasekhar VK, Viale A, Succi ND, Wiedmann M, Hu X, Holland EC. Oncogenic Ras and Akt signaling contribute to glioblastoma formation by differential recruitment of existing mRNAs to polysomes. *Mol Cell* 2003; 12:889-901.
15. Harris TE, Lawrence JC Jr. TOR signaling. *Sci STKE* 2003; 212:re15.
16. Ruggero D, Montanaro L, Ma L, Xu W, Londei P, Cordon-Cardo C, et al. The translation factor eIF-4E promotes tumor formation and cooperates with c-Myc in lymphomagenesis. *Nat Med* 2004; 10:484-6.
17. Plas DR, Talapatra S, Edinger AL, Rathmell JC, Thompson CB. Akt and Bcl-x_L promote growth factor-independent survival through distinct effects on mitochondrial physiology. *J Biol Chem* 2001; 276:12041-8.
18. Fox CJ, Hammerman PS, Cinalli RM, Master SR, Chodosh LA, Thompson CB. The serine/threonine kinase Pim-2 is a transcriptionally regulated apoptotic inhibitor. *Genes Dev* 2003; 15:1851-4.
19. Ruggero D, Grisendi S, Piazza F, Rego E, Mari F, Rao PH, et al. Dyskeratosis congenita and cancer in mice deficient in ribosomal RNA modification. *Science* 2003; 299:259-62.
20. Ruggero D, Pandolfi PP. Does the ribosome translate cancer? *Nat Rev Cancer* 2003; 3:179-92.
21. Haydon MS, Googe JD, Sorrells DS, Ghali GE, Li BD. Progression of eIF4e gene amplification and overexpression in benign and malignant tumors of the head and neck. *Cancer* 2000; 88:2803-10.
22. Li BD, Liu L, Dawson M, De Benedetti A. Overexpression of eukaryotic initiation factor 4E (eIF4E) in breast carcinoma. *Cancer* 1997; 79:2385-90.
23. Rosenwald IB, Chen JJ, Wang S, Savas L, London IM, Pullman J, Upregulation of protein synthesis initiation factor eIF-4E is an early event during colon carcinogenesis. *Oncogene* 1999; 18:2507-17.
24. Manion MK, Hockenbery DM. Targeting BCL-2-related proteins in cancer therapy. *Cancer Biol Ther* 2003; 2:S105-14.