Controversies in Giving Data Back

Gholson J. Lyon, M.D. Ph.D.
Conflicts of Interest

• I do not accept $$$ as salary from anyone other than my current employer, CSHL.

• I also work with the **nonprofit** Utah Foundation for Biomedical Research (UFBR) and the Institute for Genomic Medicine (IGM).

• Any revenue that I earn from providing medical consultation to people is donated to UFBR and IGM for the genetics research.
“Prevention” has been and will continue to be the best way to improve health.

Better sanitation = reduce infectious disease
Iodine supplementation = eliminate cretinism
Folate during pregnancy = reduce neural tube defects
PAP smears = detect pre-cancerous lesions
Reduce cigarette smoking = decrease cancer
From Base Pair to Body Plan: Celebrating 60 years of DNA

Organizers:
Alex Gann, Cold Spring Harbor Laboratory
Robert Martienssen, Cold Spring Harbor Laboratory/HHMI
“We don’t have to look for a model organism anymore, because we are the model organisms.”

– **Sydney Brenner**, Nobel Laureate, quote in 2008
Complexity

• There are ~25-100 TRILLION cells in each human body, with ~6 billion nucleotides per cell.
• There is extensive modification of DNA, RNA and proteins both spatially and temporally.
• There are higher level mechanisms of somatic mosaicism, heterosis, and likely ancestral inheritance.
Circular RNAs Are the Predominant Transcript Isoform from Hundreds of Human Genes in Diverse Cell Types

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Abstract

Most human pre-mRNAs are spliced into linear molecules that retain the exon order defined by the genomic sequence. By deep sequencing of RNA from a variety of normal and malignant human cells, we found RNA transcripts from many human genes in which the exons were arranged in a non-canonical order. Statistical estimates and biochemical assays provided strong evidence that a substantial fraction of the spliced transcripts from hundreds of genes are circular RNAs. Our results suggest that a non-canonical mode of RNA splicing, resulting in a circular RNA isoform, is a general feature of the gene expression program in human cells.


Editor: Thomas Preiss, The John Curtin School of Medical Research, Australia

Received November 7, 2011; Accepted December 28, 2011; Published February 1, 2012
We found the SAME mutation in two unrelated families, with a very similar phenotype in both families, helping prove that this genotype contributes to the phenotype observed.
The mutation disrupts the N-terminal acetylation machinery (NatA) in human cells.

Slide courtesy of Thomas Arnesen
New Syndrome with Dysmorphology, Mental Retardation, “Autism”, “ADHD”

Could be X-linked, Autosomal Recessive, multi-allelic or polygenic threshold effect?
Workup Ongoing for past 10 years

- Numerous genetic tests negative, including negative for Fragile X and many candidate genes.
- No obvious pathogenic CNVs – several microarrays without any definitive result.
- Sequenced whole genomes of Mother, Father and Two Boys, using Complete Genomics, version 2.0 CG pipeline.
- But VERY difficult to prove in this instance that any mutation (or mutations) are definitely contributing to the illness.
Worldwide Database?

• We need at least ONE Million humans with detailed phenotype, genomic, and other data followed longitudinally, and all available for analysis to anyone online.

• A “Medical Donor Information Network”, in the words of Maynard Olson.
But how do ever achieve this?
Clinical Validity with Worldwide Human Genetic Variation “database”? 

ancestry.com®  PatientsLikeMe 

Google™  23andMe 

Million Veteran Program: A Partnership with Veterans 

100,000 British Genomes
Networking of Science Model

http://lyonlab.cshl.edu/publications.html
The dreaded “Reviewer #3”
The dreaded “Reviewer #3”

• “The authors should stop and take a breath. Topol’s book (The Creative Destruction of Medicine) is a popularization of the “flying cars” variety.”
Scenic drive: In this undated photo, Dr. Paul Moller stands with two prototypes of his SkyCar and his company's flying saucer, the Neuera, which he helped develop and himself piloted in the 1970s.

[Link](http://www.dailymail.co.uk/news/article-2268402/Flying-car-developer-says-hes-80-million-closer-making-sci-fi-dream-reality.html#ixzz2Ld7gJqiT)
It takes a LONG time and Persistence to introduce new technologies and change the Status Quo
First Human Image July 3, 1977, showing heart, lungs, vertebra, musculature.

Figure 9.6  First MRI seen of a live human body, further computer-enhanced. Cross-section of Lawrence Minkoff's chest. Top of the image is anterior boundary of the chest wall. Left area is left side of chest. The heart is the principal structure in the middle and the lungs (black cavities) are on either side. More posteriorly on the left, the circular structure corresponds to the descending aorta. In the body wall, the sternum is seen anteriorly and proceeding around the ellipse alteration of light, and dark areas correspond to the intercostal muscles with rib. [RD]

From *Prize Fight: The Race and Rivalry to be First in Science*, by Morton Meyers, M.D., 2012
Present Day 2013, ~35 years later
Industrialization of Sequencing
Autonomy vs. Privacy vs. Bureaucracy
PRIVACY and PROGRESS in Whole Genome Sequencing

Presidential Commission for the Study of Bioethical Issues

October 2012
Family in Utah - Ogden Syndrome.
Policy and Governance

“If you sequence people’s exomes you’re going to find stuff,” said Gholson Lyon, a physician and researcher previously at the University of Utah, now at Cold Spring Harbor Laboratory.

As part of his research, Dr. Lyon worked with a family in Ogden, Utah. Over two generations, four boys had died from an unknown disease with a distinct combination of symptoms—an aged appearance, facial abnormalities, and developmental delay. Dr. Lyon sought to identify the genetic cause of this disease, and collected blood samples from 12 family members who had signed consent forms. The family members understood these forms to mean that they would have access to their results.

Dr. Lyon conducted exon capture and sequencing of the X chromosome—a process that analyzes specific regions of the X chromosome and is a less expensive alternative to whole genome sequencing—to analyze the blood samples. Dr. Lyon and his colleagues identified a genetic mutation, and named the disease Ogden Syndrome after the family’s hometown.

After Dr. Lyon and his team identified the genetic basis of Ogden Syndrome, one of the family members contacted him. This young mother of one daughter had submitted a blood sample for Dr. Lyon’s research. She had not been pregnant at the time, but was now four months pregnant with her second child. She knew that she was carrying a boy and wanted to know if she was a carrier of the mutation. She wanted to be able to mentally and emotionally prepare herself and her family.

By reexamining his research data, Dr. Lyon was able to see that the expectant mother was a carrier of Ogden Syndrome. This meant that her son had a 50 percent chance of being born with the disease. Dr. Lyon could not, however, legally share this important information with the family because he had conducted the original sequencing in a research laboratory that had not satisfied federally mandated standards designed to ensure the accuracy of clinical genetic results.

Instead, Dr. Lyon worked to have the mutation validated at a laboratory that satisfied those federal standards; this involved overcoming substantial bureaucratic hurdles and other obstacles that held up the process. During this time, the baby boy was born and died of Ogden Syndrome at four months of age. While knowing the results would not have changed the outcome, Dr. Lyon feels he should have been able to do more for the family.

Dr. Lyon has become an outspoken advocate for conducting whole genome sequencing in laboratories that satisfy the federal standards so that researchers can return results to participants, if appropriate. Dr. Lyon wants clear guidance for laboratories conducting genetic research and clear language in consent forms that clarifies the results that participants should expect to have returned from the researchers.
Recommendation 4.1

Funders of whole genome sequencing research, relevant clinical entities, and the commercial sector should facilitate explicit exchange of information between genomic researchers and clinicians, while maintaining robust data protection safeguards, so that whole genome sequence and health data can be shared to advance genomic medicine.

Performing all whole genome sequencing in CLIA-approved laboratories would remove one of the barriers to data sharing. It would help ensure that whole genome sequencing generates high-quality data that clinicians and researchers can use to draw clinically relevant conclusions. It would also ensure that individuals who obtain their whole genome sequence data could share them more confidently in patient-driven research initiatives, producing more meaningful data. That said, current sequencing technologies and those in development are diverse and evolving, and standardization is a substantial challenge. Ongoing efforts, such as those by the Standardization of Clinical Testing working group are critical to achieving standards for ensuring the reliability of whole genome sequencing results, and facilitating the exchange and use of these data.²¹⁶
Applied & Translational Genomics

Practical, ethical and regulatory considerations for the evolving medical and research genomics landscape.

Gholson J. Lyon¹,²* and Jeremy P. Segal³*

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Table 1. Processes involved in a CLIA-certified genetic test.

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<th>Preanalytic System:</th>
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<tbody>
<tr>
<td>1) test request and specimen collection criteria</td>
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<tr>
<td>2) specimen submission, handling and referral procedures</td>
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<tr>
<td>3) preanalytic systems assessment</td>
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</tbody>
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<table>
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<tr>
<th>Analytic System:</th>
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</thead>
<tbody>
<tr>
<td>1) a detailed step-by-step procedure manual</td>
</tr>
<tr>
<td>2) test systems, equipment, instruments, reagents, materials and supplies</td>
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<tr>
<td>3) establishment and verification of performance specifications</td>
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<tr>
<td>4) maintenance and function checks</td>
</tr>
<tr>
<td>5) calibration and calibration verification procedures</td>
</tr>
<tr>
<td>6) control procedures, test records, and corrective actions</td>
</tr>
<tr>
<td>7) analytic systems assessment</td>
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</tbody>
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<table>
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<tr>
<th>Post-Analytic System:</th>
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<tbody>
<tr>
<td>1) test report, including (among other things):</td>
</tr>
<tr>
<td>a) interpretation</td>
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<tr>
<td>b) reference ranges and normal values</td>
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<tr>
<td>2) Post-analytic systems assessment</td>
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</tbody>
</table>

I have ordered and obtained whole genome sequencing from the CLIA-certified WGS lab at Illumina. It ALREADY exists.
Will results from my blood tests be forwarded to me?

It will not be possible to give participants results of the blood tests. Due to regulations under the Clinical Laboratory Improvement Amendments (CLIA), we are legally unable to return research results to participants. Results from the blood tests will not be placed in participants' electronic health record. Participants should discuss any health concerns with their doctor or other health care provider, who can arrange any necessary and appropriate tests.

http://www.research.va.gov/mvp/veterans.cfm
accessed March 6, 2013

“A partnership is an arrangement where parties agree to cooperate to advance their mutual interests.” - Wikipedia
Dealing with the unexpected: consumer responses to direct-access BRCA mutation testing

Uta Francke¹,², Cheri Dijamco¹, Amy K. Kiefer¹, Nicholas Eriksson¹, Bianca Moiseff¹, Joyce Y. Tung¹, and Joanna L. Mountain¹

¹ 23andMe, Inc., Mountain View, CA, USA
² Department of Genetics, Stanford University School of Medicine, Stanford, CA, USA

204 BRCA1 (185delAG or 5382insC) or BRCA2 6174delT mutation carriers (130 males and 74 females) in the 23andMe database of 114,627 customers who were at least 18 years of age and had consented to participate in research.
Clinical Validity with “Worldwide Human Genetic Variation Database” and/or “Medical Donor Information Network”?

PatientsLikeMe

Google

23andMe

Facebook

Million Veteran Program: A Partnership with Veterans

100,000 British Genomes
Our Mission

• Implement an infrastructure for clinical genomic sequencing and interpretation.
• Build public trust in genomic medicine.
• Urge insurance companies to reimburse genome sequencing in clinical settings.

Figure 4. NAT activity of recombinant hNaa10p WT or p.Ser37Pro towards synthetic N-terminal peptides. A) and B) Purified MBP-hNaa10p WT or p.Ser37Pro were mixed with the indicated oligopeptide substrates (200 µM for SESSS and 250 µM for DDDIA) and saturated levels of acetyl-CoA (400 µM). Aliquots were collected at indicated time points and the acetylation reactions were quantified using reverse phase HPLC peptide separation. Error bars indicate the standard deviation based on three independent experiments. The five first amino acids in the peptides are indicated, for further details see materials and methods. Time dependent acetylation reactions were performed to determine initial velocity conditions when comparing the WT and Ser37Pro NAT-activities towards different oligopeptides. C) Purified MBP-hNaa10p WT or p.Ser37Pro were mixed with the indicated oligopeptide substrates (200 µM for SESSS and AVFAD, and 250 µM for DDDIA and EEEIA) and saturated levels of acetyl-CoA (400 µM) and incubated for 15 minutes (DDDIA and EEEIA) or 20 minutes (SESSS and AVFAD), at 37°C in acetylation buffer. The acetylation activity was determined as above. Error bars indicate the standard deviation based on three independent experiments. Black bars indicate the acetylation capacity of the MBP-hNaa10p wild type (WT), while white bars indicate the acetylation capacity of the MBP-hNaa10p mutant p.Ser37Pro. The five first amino acids in the peptides are indicated.
Further Acknowledgements
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- [@Katy_Read](https://twitter.com/Katy_Read): Like many writers, I have rituals. Before writing, I pour some coffee, open the window by my desk, and attempt to read the entire internet.