A genetic locus targeted to the nuclear periphery in living cells maintains its transcriptional competence

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The peripheral nuclear lamina, which is largely but not entirely associated with inactive chromatin, is considered to be an important determinant of nuclear structure and gene expression. We present here an inducible system to target a genetic locus to the nuclear lamina in living mammalian cells. Using three-dimensional time-lapse microscopy, we determined that targeting of the locus requires passage through mitosis. Once targeted, the locus remains anchored to the nuclear periphery in interphase as well as in daughter cells after passage through a subsequent mitosis. Upon transcriptional induction, components of the gene expression machinery are recruited to the targeted locus, and we visualized nascent transcripts at the nuclear periphery. The kinetics of transcriptional induction at the nuclear lamina is similar to that observed at an internal nuclear region. This new cell system provides a powerful approach to study the dynamics of gene function at the nuclear periphery in living cells.

Introduction

The paradoxical nature of the mammalian nucleus as a compartmentalized, yet dynamic structure has been well established based on studies of nuclear domains (for review see Spector, 1993, 2006) and nuclear protein dynamics (for review see Misteli, 2001). The nuclear envelope marks the nuclear periphery in mammalian cells and is composed of an outer and inner nuclear membrane interrupted in places by nuclear pore complexes (for reviews see Goldman et al., 2002; Hetzer et al., 2005). A nuclear lamina consisting of type V intermediate filament proteins called lamins and other lamin-associated proteins underlies the inner nuclear membrane (for review see Goldman et al., 2002). The lamin proteins form a meshwork between the inner nuclear membrane and chromatin. Lamins play an important role in nuclear envelope assembly/disassembly during mitosis and are considered to be an important determinant of nuclear architecture and gene expression (for review see Goldman et al., 2002). Microscopic analyses of fixed cells (Belmont et al., 1993) and in vitro binding assays with lamins (Taniura et al., 1995) have suggested that lamins interact with chromatin either directly or indirectly via lamina-associated polypeptide (LAP) 2 (for reviews see Goldman et al., 2002; Schirmer and Foisner, 2007). In Drosophila melanogaster, distinct regions of chromatin have been shown to directly interact with the nuclear lamina (Marshall et al., 1996; Zhao et al., 1996; Pickersgill et al., 2006). Most interestingly, cells expressing a mutant form of lamin A (LA\Delta050) associated with Hutchinson-Gilford progeria syndrome exhibit a loss of perinuclear heterochromatin, further emphasizing the role of the nuclear lamina in chromatin organization and function (Shumaker et al., 2006).

The chromosomes in an interphase nucleus occupy a discrete spatially defined subvolume of the nucleus called the chromosome territories (for review see Cremer and Cremer, 2001). In mammalian cells, chromosomes show probabilistic (Parada et al., 2003), yet nonrandom radial positions with gene-rich chromosomes at the nuclear interior and gene-poor chromosomes at the periphery (Croft et al., 1999; Cremer et al., 2001). However, the broad morphological classification of chromatin into euchromatin (less-condensed or open chromatin) and heterochromatin (more-condensed or closed chromatin) does not always correlate with gene activity (Gilbert et al., 2004).

The view that individual genes show preferred nuclear positions (i.e., interior vs. peripheral) based on their transcriptional status has been supported by a variety of evidence that the nuclear envelope is primarily a silent compartment. However, recent findings in Saccharomyces cerevisiae and D. melanogaster indicate that the nuclear periphery can be associated with both transcriptionally active and inactive genes (Andrulis et al., 1998; Gerasimova et al., 2000; Ishii et al., 2002; Brickner and Walter, 2004; Casolari et al., 2004; Gartenberg et al., 2004; Mendjan et al., 2006;
time readout of transcription (Janicki et al., 2004) and mRNP trafficking (Shav-Tal et al., 2004) after the expected trends of these processes based on molecular analyses. Using this newly developed system, we directly visualized that targeting of a genetic locus to the nuclear lamina required passage through mitosis. Upon transcriptional induction, components of the gene expression machinery were recruited to the targeted locus and nascent transcripts were visualized at the transcription site. This new cell system is a powerful tool to study the dynamics of gene function at the nuclear periphery during normal physiology and in disease states, such as envelopathies, in living cells.

Results

Development of an inducible nuclear lamina–targeting gene expression system

To address the influence of the nuclear lamina on gene expression in living human cells, we developed an inducible nuclear lamina–targeting gene expression system. For this, we took advantage of the existing U2OS-2-6-3 stable cell line developed in our laboratory to visualize gene expression in living cells (Janicki et al., 2004). Although this cell line contains a 200-copy tandem array of a reporter transgene, extensive analysis has indicated that it faithfully reports real-time readout of transcription (Janicki et al., 2004) and mRNP trafficking (Shav-Tal et al., 2004) after the expected trends of these processes based on molecular analyses. Using this newly developed system, we directly visualized that targeting of a genetic locus to the nuclear lamina required passage through mitosis. Upon transcriptional induction, components of the gene expression machinery were recruited to the targeted locus and nascent transcripts were visualized at the transcription site. This new cell system is a powerful tool to study the dynamics of gene function at the nuclear periphery during normal physiology and in disease states, such as envelopathies, in living cells.
inducible clones required two rounds of stable selection (Fig. 2 A). U2OS-2-6-3 cells were transfected with pCMV5-CymR repressor plasmid and selected in the presence of G418 to obtain the first stable line U2OS-2-6-3-CymR constitutively expressing the CymR repressor. U2OS-2-6-3-CymR cells were then transfected in parallel with one of two cumate-inducible plasmids, pCMVCuO-LacI-mCherry (control) or pCMVCuO-LacI-mCherry–Lamin B1 (targeting), and selected in the presence of puromycin and absence of cumate to obtain two cumate-inducible stable lines, namely U2OS-2-6-3-CymR-CuO-LacI-mCherry–Lamin B1 (control cell line) and U2OS-2-6-3-CymR-CuO-LacI-mCherry–Lamin B1 (targeting cell line). Both puromycin-resistant lines were screened to obtain cumate-inducible control cell line (red, nontargeted locus) and targeting cell line (red, lamina-targeted locus). (B and C) Immunofluorescence analysis of control and targeting cell lines in the absence (a–c) or presence (d–f) of cumate. Anti–lamin B1 immunolabeling marks the nuclear periphery (B [b, c, e, and f] and C [b, c, e, and f] green). (B) In the control cell line, when lacI-mCherry was not expressed the locus was not visible (a and c). In the absence of cumate and lacI-mCherry expression, the locus was visualized [d and f, red, arrow] internal to the nuclear periphery [f]. (C) In the targeting cell line, in the absence of lacI-mCherry–lamin B1 (targeting fusion) expression there was no signal for the locus (a and c). Cumate-induced targeting fusion expression resulted in targeting the locus to the lamina [d, red, arrow] and the targeted locus colocalized with endogenous lamin B1 [f, yellow, arrow]. Bars, 5 μm.

Figure 2. Generation and characterization of control and targeting cell lines. (A) Parental U2OS-2-6-3 cells were stably transfected with pCMV5-CymR repressor plasmid and selected in the presence of G418 to obtain the first stable line U2OS-2-6-3-CymR constitutively expressing the CymR repressor. U2OS-2-6-3-CymR cells were then transfected in parallel with one of two cumate-inducible plasmids, pCMVCuO-LacI-mCherry (control) or pCMVCuO-LacI-mCherry–Lamin B1 (targeting), and selected in the presence of puromycin and absence of cumate to obtain two cumate-inducible stable lines, namely U2OS-2-6-3-CymR-CuO-LacI-mCherry–Lamin B1 (control cell line) and U2OS-2-6-3-CymR-CuO-LacI-mCherry–Lamin B1 (targeting cell line). Both puromycin-resistant lines were screened to obtain cumate-inducible control cell line (red, nontargeted locus) and targeting cell line (red, lamina-targeted locus). (B and C) Immunofluorescence analysis of control and targeting cell lines in the absence (a–c) or presence (d–f) of cumate. Anti–lamin B1 immunolabeling marks the nuclear periphery (B [b, c, e, and f] and C [b, c, e, and f] green). (B) In the control cell line, when lacI-mCherry was not expressed the locus was not visible (a and c). In the absence of cumate and lacI-mCherry expression, the locus was visualized [d and f, red, arrow] internal to the nuclear periphery [f]. (C) In the targeting cell line, in the absence of lacI-mCherry–lamin B1 (targeting fusion) expression there was no signal for the locus (a and c). Cumate-induced targeting fusion expression resulted in targeting the locus to the lamina [d, red, arrow] and the targeted locus colocalized with endogenous lamin B1 [f, yellow, arrow]. Bars, 5 μm.

visualized by a lac repressor–mCherry fusion protein (Fig. 1 B, a), RNA transcripts by the MS2 coat–YFP fusion protein (Fig. 1 B, b), and the protein product as a CFP-SKL protein (Fig. 1 B, c) that is targeted to cytoplasmic peroxisomes.

We used the U2OS-2-6-3 cells to develop cumate-inducible cell lines stably expressing lacI-mCherry or lacI-mCherry–lamin B1. The targeting fusion protein (lacI-mCherry–lamin B1) associates with the stably integrated locus in U2OS-2-6-3 cells via a lac operator–repressor interaction and with the nuclear lamina via a lamin B1 protein. The cumate expression system has two components: a repressor plasmid (pCMV5-CymR repressor) and a cumate-inducible response plasmid (pCMVCuO–lamin B1 promoter [CuO]). The response plasmid has a CMV promoter and a CymR repressor binding site (CuO). Development of stable cumate-inducible clones required two rounds of stable selection (Fig. 2 A). U2OS-2-6-3 cells were transfected with pCMV5-CymR repressor plasmid (Fig. 2 A) and stable G418-resistant clones were screened by transient transfection with pCMVCuO–lamin B1 in the presence or absence of cumate to obtain a tightly regulated cell line. This first stable cell line was referred to as U2OS-2-6-3-CymR (Fig. 2 A). Subsequently, U2OS-2-6-3-CymR was transfected with either pCMVCuO–lamin B1 (control) or pCMVCuO–lamin B1 (targeting) plasmids and stable puromycin resistance lines were screened in the absence or presence of cumate to obtain tightly regulated cell lines. These stable cumate-inducible cell lines were referred to as U2OS-2-6-3-CymR–CuO–lamin B1 (Fig. 2 A, control cell line) or U2OS-2-6-3-CymR–CuO–lamin B1 (Fig. 2 A, targeting cell line).
To ascertain whether the targeting fusion exhibited properties similar to endogenous lamin B1, we examined its localization in regard to the localization of lamin B–interacting proteins, such as lamin A and LAP2\textsubscript{H9252}. We transiently expressed YFP–lamin A or YFP–LAP2\textsubscript{H9252} (Fig. 3, A or B) in targeting cells and induced targeting fusion expression for 12 h with cumate. The targeted locus was enriched for YFP–lamin A (Fig. 3 A, b and c, arrows) and YFP–LAP2\textsubscript{H9252} (Fig. 3 B, b and c, arrows), indicating that the targeting fusion behaved analogous to endogenous lamin B1 protein.

Targeting the locus to the nuclear lamina requires passage through mitosis

Having obtained cumate-inducible stable cell lines, we were interested in determining the temporal window during which the locus is targeted to the nuclear periphery. A cumate induction time course study showed that the percentage of targeted cells increased with the passage of time, indicating that targeting could be a postmitotic event (unpublished data). To test this possibility, we performed a G1/S-phase block by treating the targeting cells with hydroxyurea (Fig. 4 A). If cells required passage through mitosis for the locus to be targeted to the nuclear lamina, cells blocked at G1/S and induced with cumate would have a high percentage of nontargeted cells (locus internal to the nuclear lamina) compared with targeted cells (locus targeted to the nuclear lamina). However, in the absence of a G1/S block, the cumate-induced targeting cells would divide asynchronously and, therefore, more targeted loci should be observed.

Fig. 4 B shows representative images of cells used for counting targeted loci in G1/S-blocked or asynchronous cells. When cells were blocked in G1/S phase, there was an approximately twofold increase in the number of nontargeted cells compared with targeted cells (Fig. 4 C). However, when cells were allowed to divide asynchronously there was a twofold increase in the number of targeted cells compared with non-targeted cells. These results indicated that targeting likely required passage through cell division. Furthermore, and consistent with
Figure 4. **Targeting the locus to the nuclear lamina requires passage through mitosis.** (A) G1/S-phase block using hydroxyurea in targeting cell line from day 0 (D0) to day 3.5 (D3.5). (B) Representative images of LacI-mCherry–Lamin B1–expressing cells used for counting to quantify G1/S-blocked non-targeted (a, red, arrow) and targeted (b, red, arrow) cells and asynchronous nontargeted (c, red, arrow) and targeted (d, red, arrow) cells. Bar, 5 μm. (C) Quantification of the percentage of nontargeted and targeted cells in G1/S-blocked and asynchronous cells. The graphical data represents the mean and standard deviation of three experiments (n = 100 nuclei per experiment). Approximately 10% of the cells did not exhibit a nuclear mCherry signal. (D) Still images (a–o) from time series (0–90 min, G2 to early G1 phase) of targeting clone H2A-YFP going through mitosis. Targeting cells stably expressing histone H2A-YFP (green) were induced to express LacI-mCherry–Lamin B1 (red, arrow). Images are projections of deconvolved z stacks taken after every 10 min. Bar, 5 μm. See video 1 [available at http://www.jcb.org/cgi/content/full/jcb.200706060/DC1] for the entire time series.
the nuclear lamina during early G1 (Moir et al., 2000). Furthermore, chromatin is most dynamic during early G1 phase of the cell cycle before the establishment of chromosome territories (Chubb et al., 2002; Walter et al., 2003). Based on these observations, we hypothesized that the best opportunity for the locus to be targeted could be during mitosis, at the transition from telophase to early G1 phase. To test this hypothesis we followed the dynamics of the locus during mitosis in the targeting cell line stably expressing histone H2A-YFP in the absence of hydroxyurea. To minimize phototoxicity associated with cell imaging, we reduced the time window of imaging by roughly preselecting cells based on cell size (large nucleus with a spherical nontargeted locus indicative of a cell in G2 phase) before this possibility, analysis of the dynamics of loci associated with the targeting fusion protein for up to 24 h, during interphase and in the absence of hydroxyurea, did not result in the observation of targeted loci that were stably associated with the nuclear periphery (Fig. 5). Interestingly, at the 5-h 20-min time point the locus shown in Fig. 5 transiently contacts the peripheral lamina, creating tension at the point of contact (Fig. 5, d, arrow); however, it subsequently moves away from the periphery. These results indicate that binding of the lacI-mCherry–lamin B1 targeting fusion to the locus is not sufficient for its stable targeting to the peripheral nuclear lamina during interphase.

Prior studies have shown that lamin B1 is targeted to the nuclear envelope in late telophase and is polymerized as part of the nuclear lamina during early G1 (Moir et al., 2000). Furthermore, chromatin is most dynamic during early G1 phase of the cell cycle before the establishment of chromosome territories (Chubb et al., 2002; Walter et al., 2003). Based on these observations, we hypothesized that the best opportunity for the locus to be targeted could be during mitosis, at the transition from telophase to early G1 phase. To test this hypothesis we followed the dynamics of the locus during mitosis in the targeting cell line stably expressing histone H2A-YFP in the absence of hydroxyurea. To minimize phototoxicity associated with cell imaging, we reduced the time window of imaging by roughly preselecting cells based on cell size (large nucleus with a spherical nontargeted locus indicative of a cell in G2 phase) before

![Figure 5. Targeting of lamin B1-associated loci does not take place during interphase.](image-url)
imaging, we used short exposure times and rapid shutters, and we placed a UV-blocking filter in the light path of our imaging system. Fig. 4D shows still images (a–o) from a time series (0–90 min, G2 to early G1 phase) of the targeting line H2A-YFP progressing through mitosis (Video 1, available at http://www.jcb.org/cgi/content/full/jcb.200706060/DC1). At 0 min, the locus was spherical (Fig. 4D, a and c, red, arrows) and localized away from the nuclear lamina. Chromatin marked by H2A-YFP (Fig. 4D, b and c, green) was homogenously distributed within the large nucleus, which is indicative of a G2 cell. At the 10-min time point, chromatin began to condense, which is indicative of cells in early prophase (Fig. 4D, e and f, green). The locus was still spherical and was not targeted to the intact lamina (Fig. 4D, d and f, red, arrows). By 20 min, the locus was still visible as a round spot, whereas the lamina started to dissociate and the chromosomes exhibited a prometaphase configuration (Video 1). By 30 min, the locus continued to appear as a round spot, whereas the lamina was completely dissociated and the chromosomes began to congress toward the metaphase plate (Video 1). At 40 min, the locus (Fig. 4D, g and i, red, arrows) was observed aligned at the metaphase plate (Fig. 4D, h and i, green). At the 50-min time point, chromatids (Fig. 4D, k and l, green) started moving toward the poles and loci appeared as coaligned doublets (Fig. 4D, j and l, red, arrows) that were slightly stretched along their long axis, possibly because of torsional force generated during chromosome segregation at anaphase. By 70 min, which marks late telophase, both loci were targeted to the nuclear lamina (Video 1). At the 80- and 90-min time points, which mark late telophase, both loci were targeted to the nuclear lamina (Fig. 4D, m and o, red, arrows; and Video 1).

Figure 6. The targeted locus is stably anchored to the nuclear lamina during interphase. Targeting cells stably expressing histone H2A-YFP (green) were induced to express LacI-mCherry-Lamin B1 (red, arrow) and cells with a targeted locus were selected for imaging. Still images (a–o) are shown from an interphase time series (0–19 h, G1 to G2/M phase). Images are projections of deconvolved z stacks taken every 20 min. Bar, 5 μm.
mRNA splicing factor SF2/ASF, were recruited to the targeted locus at the nuclear lamina. To address this possibility, we took advantage of the transcriptional inducibility of the locus by Dox. We transiently coexpressed Tet-On with either YFP–RNA pol II or YFP–SF2/ASF (Fig. 8, A or B) in targeting cells and induced targeting-fusion expression for 12 h with cumate (Fig. 8, A or B). In the absence of Dox (Fig. 8, A [a–d] or B [a–d]), YFP–RNA pol II and YFP–SF2/ASF were not recruited to the targeted loci (Fig. 8, A [b and d] or B [b and d], no green enrichment, arrows) and there was no CFP-SKL protein at the cytoplasmic peroxisomes (Fig. 8, A [c and d] or B [c and d]). However, in the presence of Dox (Fig. 8, A [e–h] or B [e–h]) for 5 h, YFP–RNA pol II and YFP–SF2/ASF were recruited to the targeted loci (Fig. 8, A [g and h] or B [g and h]). Quantitation showed that 76% (n = 50) of the control loci, 68% (n = 50) of lamin B1–bound internal loci in targeting cells, and 64% (n = 50) of targeted loci in targeting cells showed recruitment of YFP–RNA pol II in the presence of Dox. Also, ~80% (n = 50) of the control cells, 76% (n = 50) of

Although in the 2D projection it appears that only one locus is at the edge of the nucleus, the targeting of both loci is obvious when examined in individual Z sections (Fig. S2 and Video 2). Thus, our live cell data showed that the temporal window for targeting the locus to the nuclear lamina was during the transition from late telophase to early G1 phase of the cell cycle.

Next, we were interested in examining the stability of the targeted locus with the nuclear periphery during interphase. As shown in Fig. 6, the targeted locus was stably anchored to the peripheral lamina throughout the 19-h period of interphase observation. Subsequently, we examined cells containing a targeted locus as the cells progressed through mitosis, and we found that targeted loci remained targeted in daughter cells (Fig. 7 and Video 3, available at http://www.jcb.org/cgi/content/full/jcb.200706060/DC1).

Transcriptional induction at the nuclear periphery

Next, we addressed whether components of the gene expression machinery, such as RNA polymerase II (RNA pol II) and the pre-mRNA splicing factor SF2/ASF, were recruited to the targeted locus at the nuclear lamina. To address this possibility, we took advantage of the transcriptional inducibility of the locus by Dox. We transiently coexpressed Tet-On with either YFP–RNA pol II or YFP–SF2/ASF (Fig. 8, A or B) in targeting cells and induced targeting-fusion expression for 12 h with cumate (Fig. 8, A or B). In the absence of Dox (Fig. 8, A [a–d] or B [a–d]), YFP–RNA pol II and YFP–SF2/ASF were not recruited to the targeted loci (Fig. 8, A [b and d] or B [b and d], no green enrichment, arrows) and there was no CFP-SKL protein at the cytoplasmic peroxisomes (Fig. 8, A [c and d] or B [c and d]). However, in the presence of Dox (Fig. 8, A [e–h] or B [e–h]) for 5 h, YFP–RNA pol II and YFP–SF2/ASF were recruited to the targeted loci (Fig. 8, A [g and h] or B [g and h]). Quantitation showed that ~76% (n = 50) of the control loci, 68% (n = 50) of lamin B1–bound internal loci in targeting cells, and 64% (n = 50) of targeted loci in targeting cells showed recruitment of YFP–RNA pol II in the presence of Dox. Also, ~80% (n = 50) of the control cells, 76% (n = 50) of
lamin B1–bound internal loci in targeting cells, and 74% (n = 50) of targeted loci in targeting cells showed recruitment of YFP-SF2/ASF in the presence of Dox. In the absence of Dox, ~96% (n = 50) of the control or targeting loci did not show recruitment of YFP–RNA pol II or YFP-SF2/ASF. Because control and targeting cells showed a relatively similar percentage of recruitment of the gene expression machinery, association with the nuclear lamina does not appear to preclude accessibility to the gene expression machinery.

Because we observed that the gene expression machinery was recruited to the targeted locus, we were next interested in directly visualizing transcription at the targeted locus. To do so, we took advantage of the fact that the mRNA that is transcribed from the genetic locus contains 24 tandem MS2 translational operators (MS2 repeats; Janicki et al., 2004). As the MS2 coat protein directly binds to these repeats, we were able to use an MS2-YFP fusion protein to localize RNA transcripts at the locus in living cells. To directly visualize transcription, the control or targeting cells were transiently transfected with a dual promoter vector expressing both Tet-On and MS2-YFP simultaneously (Fig. 9, A and B, pVitro2–Tet-On + MS2-YFP, green), as shown in representative images. The cells were first induced for 12 h with cumate to express the targeting fusion protein (red, arrows), and then incubated in the absence of Dox (A [a–d] or B [a–d]) or presence of Dox (A [e–h] or B [e–h]) for 5 h. In Dox (−) condition, YFP–RNA pol II or YFP-SF2/ASF was not recruited (A [b and d] or B [b and d], arrows) and CFP-SKL protein was not observed (A [c and d] or B [c and d]). In Dox (+) condition, YFP–RNA pol II and YFP-SF2/ASF were recruited (A [f] or B [f], arrows) and localized with the locus (A [h] or B [h], arrows) and CFP-SKL protein was observed at cytoplasmic peroxisomes (A [g and h] or B [g and h]). Bars, 5 μm.

Figure 8. Components of the gene expression machinery are recruited to the lamina-targeted locus upon gene activation. Recruitment of YFP–RNA pol II (A) or YFP–SF2/ASF (B) to targeted and activated locus at the lamina. Targeting cells were transiently transfected with pTet-On and YFP–RNA pol II or YFP-SF2/ASF [green], induced for 12 h with cumate to express the targeting fusion protein (red, arrows), and then incubated in the absence of Dox (A [a–d] or B [a–d]) or presence of Dox (A [e–h] or B [e–h]) for 5 h. In Dox (−) condition, YFP–RNA pol II or YFP-SF2/ASF was not recruited (A [b and d] or B [b and d], arrows) and CFP-SKL protein was not observed (A [c and d] or B [c and d]). In Dox (+) condition, YFP–RNA pol II and YFP-SF2/ASF were recruited (A [f] or B [f], arrows) and localized with the locus (A [h] or B [h], arrows) and CFP-SKL protein was observed at cytoplasmic peroxisomes (A [g and h] or B [g and h]). Bars, 5 μm.
Figure 9. **Transcription of the targeted locus at the nuclear lamina.** (A and B) Representative images used for counting cells to quantify the percentage of cells showing transcription in control or targeting conditions. Control cells or targeting cells were transiently transfected with pVitro2–tet-on + MS2-YFP (green), induced for 12 h with cumate for targeting fusion expression (red, arrows), and then incubated in the absence (a–c) or presence (d–f) of Dox for 5 h. In the Dox (+) condition, MS2-YFP accumulated at the site of the targeted locus (B, e, green enrichment, arrow) and MS2-YFP localized with the locus (B, f, arrow). In the absence of Dox, ~90% of control cells and ~70% of targeting cells were transcriptionally active. Arrows in A (b) and B (b) represent absence of accumulation of MS2-YFP. n = 100 nuclei per experiment for each control and test. Bars, 5 μm. (C) Still images (a–p) from time series (0–5 h) of targeting cells showing transcription at the lamina. At 0 h, in Dox (−) condition, targeted locus (a and d, arrows) showed no accumulation of MS2-YFP (b and e, arrows) and no CFP-SKL protein in peroxisomes (c and d). In Dox (+) condition, transcription was observed at the targeted nuclear lamina as observed by the accumulation of MS2-YFP (f, j, and n, arrows) and CFP-SKL localization (k, l, o, and p) over time. Images are projections of deconvolved z stacks taken every 20 min. Arrows: (e, i, and m) decondensed targeted locus; (h, l, and p) accumulation of MS2-YFP. Bar, 5 μm. See Video 4 (available at http://www.jcb.org/cgi/content/full/jcb.200706060/DC1) for the entire time series.
loci were transcriptionally active based on MS2-YFP accumulation in the presence of Dox. These results indicated that although association of lamin B with a locus, internal or targeted, had some effect on the percentage of loci that were able to be transcriptionally induced, a significant percentage of gene loci anchored to the nuclear lamina could be induced. However, because the transcriptional analysis was performed 12 h after cumate-induced expression of lacI-mCherry–lamin B1, it is not possible, based on fixed-cell analysis, to distinguish whether Tet-On–induced transcription initiated before or after the locus reached the nuclear lamina.

To directly visualize transcription at the nuclear lamina in living mammalian cells, targeting cells were transiently transfected with pVitro2–Tet-Off + MS2-YFP and incubated with cumate for 12 h to induce targeting to the nuclear lamina. Fig. 9 C shows still images (a–p) from a time series (0–5 h) of targeting cells during transcriptional activation at the nuclear lamina. Cells that exhibited the locus targeted and plastered onto the nuclear lamina and expressed Tet-Off and MS2-YFP (based on YFP signal) were selected for imaging. At 0 h, in the absence of Dox, the locus was condensed (Fig. 9 C, a and d, arrows), there was no transcription, and no MS2-YFP accumulation was visualized at the locus (Fig. 9 C, b and d, green, arrows). At 20 min after the addition of Dox, MS2-YFP was first observed at the locus with no visible decondensation of the locus or accumulation of CFP-SKL protein in the peroxisomes (Video 4, available at http://www.jcb.org/cgi/content/full/jcb.200706060/DC1). By 1 h in Dox (+) condition, the targeted locus started to decondense (Fig. 9 C, e and h, arrows) and an increase of MS2-YFP was observed at the locus (Fig. 9 C, f, arrow). CFP-SKL protein was first observed at 1 h 20 min after induction (Video 4), although it was more apparent at later time points, and by 2 h peroxisome labeling was very apparent (Fig. 9 C, k and l, and Video 4).

By 5 h in the presence of Dox, the targeted locus continued to be transcribed at the nuclear lamina (Fig. 9 C, n, arrow) and an increased CFP-SKL protein signal marked the peroxisomes (Fig. 9 C, o and p). Thus, our live-cell data shows that transcription can be induced and RNA synthesis visualized at the nuclear periphery in mammalian cells.

Having visualized transcription at the nuclear periphery, we were next interested in determining whether there was a significant difference in the kinetics of transcription when the locus was located at an internal nuclear site versus at the periphery. To do so, control or targeting cells were transiently transfected with pVitro2–Tet-Off + MS2-YFP and induced for 12 h with cumate for control or targeting fusion expression and cells were imaged in the YFP and mCherry channels. The ratio of MS2-YFP fluorescence intensity of the locus to the entire nucleoplasm (mean values) was expressed as a percentage and was plotted against time of Dox induction (each time point is represented by the mean and standard deviation).

Discussion

At the nuclear envelope, lamins bind to chromatin either directly or indirectly through LAPs (for reviews see Goldman et al., 2002; Schirmer and Foisner, 2007) and hence are thought to regulate higher order chromatin structure. In addition to the lamins and various lamina-associated proteins, a multitude of nuclear envelope transmembrane proteins of unknown function have recently been identified (for review see Schirmer and
Gerace, 2005), indicating an increased complexity of the nuclear envelope. In vitro studies showed that lamin B1 binds to native core histones (Taniura et al., 1995) and to specific DNA sequences called matrix attachment regions (Luderus et al., 1992). Analysis of D. melanogaster B-type lamin (Dmo) revealed TRAT amino acid and NLS sequences in the tail domain that directly interact with histone H2A (Mattout et al., 2007). Recently, a genome-wide screen in D. melanogaster Kc cells using DNA adenine methyltransferase identification technology showed that ~500 genes interact with Dmo (Pickersgill et al., 2006). However, these genes show no consensus DNA-binding sequence for B-type lamin that would enable anchoring of perinuclear chromatin to the lamina. In the absence of such a DNA-consensus sequence, it becomes difficult to study the anchoring process. However, the targeting system that we developed has allowed us to address important issues relating to the spatial and temporal aspects of targeting as well as its functional consequences.

Live-cell studies from different organisms in interphase nuclei showed that chromatin, in general, exhibits constrained diffusional random walk motion, with a mean radius of confinement of ~0.5 μm in the mammalian nucleus (for reviews see Spector, 2003; Lanctot et al., 2007). These studies also indicated that unlike in yeast and D. melanogaster, wherein the dynamic nature of chromosomes allows their probing of significant portions of the nuclear volume, the mammalian chromosome visits a limited nuclear volume during interphase. Furthermore, the diffusion coefficient of human chromatin is fourfold lower than that of yeast (Chubb et al., 2002). This constraint on mammalian chromatin could be a reflection of the complexity of the genome and the constraint posed by the physical attachment of chromatin to nuclear compartments (Chubb et al., 2002). Although chromosomes are organized as distinct territories in interphase nuclei, several studies have shown that they are dynamic in early G1 and can be repositioned with respect to each other or other nuclear compartments (for reviews see Spector, 2003; Thomson et al., 2004; Lanctot et al., 2007). Furthermore, chromatin has been shown to frequently exhibit long-range movements of >2 μm during this phase of the cell cycle (Walter et al., 2003). This is also the time window when lamin B1 is incorporated into the polymerizing lamina (Moir et al., 2000). It is very likely that the constrained diffusion of chromatin results from interaction of chromatin to structural elements such as the nuclear lamina or the nuclear pore complex (Marshall, 2002). It has been suggested that the chromatin–nuclear lamina interactions, which are disrupted during mitosis, are reformed during early G1 phase of the cell cycle and, indeed, this temporal window of increased chromatin mobility is in fact when many aspects of nuclear organization are established (Parada et al., 2003; for review see Spector, 2003). We observed targeting of the locus to the nuclear lamina during the same temporal window, correlating with the dynamics of the lamin B1 protein and bulk chromatin. Hence, we propose, based on our targeting study, that the contribution by the lamina in establishing nuclear architecture and chromatin organization occurs during the early G1 phase of the cell cycle.

An important question regarding the role of spatial positioning to gene function is whether subnuclear position of a gene, relative to nuclear landmarks, influences gene activity (for reviews see Spector, 2003; Misteli, 2005). Although a classical case involving the phenomenon of position-effect variegation of the brown locus in D. melanogaster indicates that repositioning to a nearby heterochromatic region results in trans-silencing (Csink and Henikoff, 1996; Dernburg et al., 1996), A5 transgene insertion into heterochromatin in mouse pre-B cells does not result in silencing, and repositioning away from heterochromatin was not required for gene activation (Lundgren et al., 2000). Together, these studies indicate that absolute gene position may not be critical for gene activation or that position may be critical for some genes and not for others. The accessibility of chromosome territories (Verschure et al., 1999; for review see Cremer and Cremer, 2001) and the dynamic nature of chromatin and nucleoplasmic proteins (Becker et al., 2002; Dundr et al., 2002; Kimura et al., 2002; Phair et al., 2004; Chen et al., 2005) provides the opportunity for genes anywhere within a chromosome territory to be easily accessible to both gene activators and repressors (Meaburn and Misteli, 2007). Our findings are consistent with the dynamic accessibility by nuclear proteins, wherein upon transcriptional activation the transcription machinery, as well as the pre-mRNA processing machinery, was recruited to the locus irrespective of its nuclear location, resulting in location-independent gene expression. However, we are not excluding the possibility that there may be other microenvironments along the nuclear periphery, or for that matter at internal nuclear regions, that may be more attuned to transcriptional silencing. It is also interesting to consider that the large size of the 200-copy gene array could generate a microenvironment. Nonetheless, our study shows that there is not a discernible effect on expression of the peripheral versus internal nuclear compartments on this array.

Perhaps the best cases that correlate changes in gene position to gene activation/expression in relation to nuclear periphery in mammals occur during differentiation of specialized cell types, such as erythroid cells, lymphocytes, or embryonic stem cells (for review see Lanctot et al., 2007). For example, the repositioning of genes, such as immunoglobulin heavy chain in B lymphocytes, C-maf in T cells, Mash1 in neuronal cells, C/EBP in adenocarcinoma cells, and MyoD in myoblasts, from the nuclear periphery to the nuclear interior upon activation has been well documented (Kosak et al., 2002; Hewitt et al., 2004; Zink et al., 2004; Lee et al., 2006; Williams et al., 2006; Lanctot et al., 2007). However, gene activation is not always associated with movement away from the nuclear periphery, as was observed in other studies based on interferon γ, β-globin, and the CD8 locus during differentiation or microarray gene expression analysis on lamin B1−/− mouse fibroblasts (Hewitt et al., 2004; Kim et al., 2004; Ragoczky et al., 2006; Malhas et al., 2007). This is consistent with our observation that a lamina-targeted genetic locus can be activated and transcribed at the lamina. Most importantly, although we observed a similar kinetics of transcriptional induction between the targeted and nontargeted loci, we did observe some differences in the percentage of cells that showed recruitment of the gene expression machinery. However, the complexities of the mammalian nuclear periphery and the fact that we are comparing data points from two different cell lines (although with the same genomic integration site of lac operator)
makes it difficult to determine the absolute significance of the observed differences. Therefore, as is the case in yeast and *D. melanogaster*, it is likely that there will not be a single rule that dictates the relationship between the nuclear periphery and transcription.

In summary, we have developed an inducible live-cell system in which the position of a genetic locus can be targeted from a more internal nuclear region to the nuclear periphery in mammalian cells. Interestingly, the mechanism of targeting of the locus to the nuclear periphery requires one round of cell division and nuclear reassembly. Transcriptional analysis indicated that the targeted locus retained its transcriptional activity, which is similar to the locus at an internal nuclear region. This new cell system is a powerful tool to study the dynamics of gene function at the nuclear periphery during normal physiology and in disease states, such as envelopathies, in living cells.

**Materials and methods**

**Plasmid constructs**

pTet-On was purchased from Clontech Laboratories, Inc., pVitro2 was purchased from Invitrogen, pmCV5-CymR and pmCV-CuO were purchased from Qbiogene, and pCMV-lamin B1 and pCMV-lamin A were purchased from Open Biosystems. pmCV-CuO-lacI-mCherry was constructed by cloning lacI-mCherry fragment (mCherry was a gift from R. Tsien [University of California, San Diego, La Jolla, CA] and was initially PCR amplified) into pCMV-CuO. pCMV-CuO-lacI-mCherry–lamin B1 B1 was constructed by cloning lamin B1 into pCMV-CuO-lacI-mCherry. pSV2-YFP–lamin A was obtained by cloning lamin A into pSV2-YFP-C1 [Janicki et al., 2004]. pVitro2-Tet-On + MS2-YFP was made by sequential cloning of Tet-On and MS2-YFP into pVitro2 dual promoter vector. pCMV-M52-YFP, pSV2-YFP–RNA poly II, pSV2-YFP–SF2/ASF, and pYFP–TAN1 plasmids were previously described [Janicki et al., 2004]. pCMV–HA2–YFP and pSV2–YFP–AP2β were subcloned by S. Janicki (The Wistar Institute, Philadelphia, PA).

**Cell culture and transfection**

Cells were grown in DME supplemented with penicillin-streptomycin (Invitrogen) and transfected with 100 μg/ml each of penicillin-streptomycin and hygromycin B (Invitrogen) and 10% tet system–approved FBS (Clontech Laboratories, Inc.). Transient transfection was performed on cells growing on glass coverslips using Fugene 6 reagent (Roche) as per the manufacturer’s protocol or by electroporation (Janicki et al., 2004) using a Gene Pulser II (Bio-Rad Laboratories, Hercules, CA). Because immediate addition of cumate (Qbiogene) drug seemed to decrease transfection efficiency, 200 μg/ml each of penicillin-streptomycin and hygromycin B (Invitrogen) for 7 d. 100 individual colonies for each transfection were subcloned by S. Janicki (The Wistar Institute, Philadelphia, PA).

**Live-cell deconvolution microscopy**

For time-lapse imaging of transcriptional activation, targeting cells grown on coverslips were transiently transfected with pVitro2–Tet-On + MS2-YFP and induced with cumate as before. Cells were then placed in an FCS2 chamber (Biopetech), and phenol red–free Leibovitz’s L15 (live cell) medium with 200 μg/ml cumate and 1 μg/ml Dox was also attached to the FCS2 chamber through an additional inlet (initially closed). The whole setup was mounted on a restoration microscope system (DeltaVision RT; Applied Precision) equipped with an inverted microscope [IX70; Olympus], rapid shutters, and a 60×/1.4 NA oil immersion objective lens (planApo; Olympus) using CFP/YFP/mCherry filters (Chroma Technology Corp.), a cooled charge-coupled device camera (Photometrics Cool SNAP HQ; Roper Scientific) in a thermally insulated temperature-controlled chamber at 37°C. To minimize photosensitivity/phototoxicity from blue light during live-cell imaging, our DeltaVision RT has a sharp cut-off (long-pass) filter at ~420 nm. Z stacks (0.4 μm, 1×1 binning) of transcriptionally inactive cells with a plastered locus (lamin B channel), a uniform diffuse nucleoplasmic signal (CFP channel) with no peroxisomal signal (CPF channel), were collected at 0 min immediately after imaging. Live-cell medium with cumate and Dox from the second syringe was perfused into the FCS2 chamber and corrected for any drift before the next time frame of imaging began at 20 min, and imaging continued with an occasional perfusion of fresh medium. For time-lapse imaging during mitosis, targeting cells H2A-YFP were grown on coverslips and preinduced with 70 μg/ml cumate for 2–3 h and then placed in an FCS2 chamber, and live-cell medium containing 70 μg/ml cumate was perfused into the chamber from a syringe. To minimize the exposure of cells to fluorescent light, cells showing a nontargeted spherical locus at around G2/M phase were chosen by visualization of the H2A-YFP chromatin pattern and nuclear size and imaged through mitosis. Z stacks (0.4 μm, 2×2 binning) were collected (mCherry and YFP channel) every 10 min with occasional perfusion of fresh medium. Z stacks were deconvolved.
using SoftWoRx 2.50. In-focus sections of deconvolved images were projected, saved in tif format, and processed using Photoshop 7.0 (Adobe) and Illustrator 10.0 (Adobe). tif images from the time series were then combined using QuickTime Pro (Apple) to make videos.

**Quantitation of RNA synthesis at the locus**

Control cells or targeting cells grown on live-cell coverslips were transiently transfected with pVitro2–Tet-on + MS2-YFP and induced for 12 h with cumatra for targeting fusion expression. The transfected cells were then imaged in the mCherry and YFP channels in Dox (−) condition (0 h), followed by imaging in Dox (+) condition every 20 min for a total period of 5 h. Raw images were then projected and the mean fluorescence intensity of the locus and of the whole nuclei were extracted using edit polygon and edit 2D polygon finder programs in SoftWoRx 2.50. The ratio of fluorescence intensity of the locus to the entire nucleus (mean values) was expressed as a percentage and was plotted against time of Dox induction.

**Online supplemental material**

Fig. S1 shows that the expressed targeting fusion is associated with the lac operator DNA at the targeted locus at the nuclear periphery. Fig. S2 shows serial z sections, which show targeting of the locus to the nuclear periphery. Video 1 shows that targeting of the locus to the lamina occurs at the end of mitosis. Video 2 shows serial z sections at a single time point, which show that the locus is targeted to the nuclear periphery. Video 3 shows that the targeted locus at the nuclear lamina remains targeted upon passage through mitosis. Video 4 shows transcription from the targeted locus at the lamina. Online supplemental material is available at http://www.jcb.org/cgi/content/full/jcb.200706060/DC1.

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